Judul Artikel	: The residue of formalin in catfish (Clarias gariepinus)after processing and storage for short periods
Nama Jurnal	: AACL Bioflux
ISSN	: 1844-9166
Tahun Terbit	: 2020
Penerbit	: Bioflux Publishing House
Volume	: 13
Nomor	: 6
No. Halaman	: 3441-3446
Url Cek Plagiasi	: http://reposister.almaata.ac.id/id/eprint/2542/1/P1_turnitin.pdf
Url Index	: https://www.scopus.com/sourceid/19300156808
Url Jurnal	: http://bioflux.com.ro/docs/2020.3441-3446.pdf

No Kegiatan Tanggal Submit 3 Oktober 2020 Preliminary acceptance (Lolos untuk Direviu) 3 November 2020 2 Pembayaran Proses Publikasi 13 November 2020 3 4 Pemberitahuan Hasil Reviu Artikel 29 November 2020 Pengiriman Revisi Artikel 4 Desember 2020 5 Pemberitahuan Revisi 2 7 Desember 2020 6 7 Penetapan Revisi 1 13 Desember 2020 Pengiriman Hasil Penetapan Revisi 1 8 15 Desember 2020 9 Penetapan Revisi 2 15 Desember 2020 Pengiriman Hasil Penetapan Revisi 2 10 15 Desember 2020 11 Pengiriman Bukti Pembayaran 16 Desember 2020 Pengiriman Draft Final 20 Desember 2020 12 Revisi Draft Final 23 Desember 2020 13 Balasan Revisi Draft Final 27 Desember 2020 14 15 Persetujuan Publikasi 28 Desember 2020 16 Pemberitahuan Publikasi Manuskrip 29 Desember 2020

Rekapitulasi Proses Penerbitan Artikel



verianiaprilia verianiaprilia <verianiaprilia@almaata.ac.id>

Submit Artikel: 3 Oktober 2020

manuscript submission

2 pesan

verianiaprilia verianiaprilia <verianiaprilia@almaata.ac.id> Kepada: zoobiomag2004@yahoo.com 3 Oktober 2020 pukul 02.45

Dear Dr. Ioan Valentin Petrescu-Mag

Hereby I would like to submit the manuscript entitled "**The residue of formalin in catfish** (*Clarias gariepinus*) **after a short period processing and storage**" to Aquaculture, Aquarium, Conservation & Legislation - International Journal of the Bioflux Society.

I attach the manuscript with the submission letter. Please give me feedback whether this can be processed or not or maybe there is other uncomplete requirement.

Please do not hesitate to contact me at

email: verianiaprilia@almaata.ac.id phone number: +6285802681312

Thank you

Regards, Veriani Aprilia

2 lampiran

Submission letter AACL Bioflux.pdf

isny maulidevi_AACL Bioflux.doc 165K

verianiaprilia verianiaprilia <verianiaprilia@almaata.ac.id> Kepada: zoobiomag2004@yahoo.com 15 Oktober 2020 pukul 12.43

[Kutipan teks disembunyikan]

2 lampiran

Bubmission letter AACL Bioflux.pdf

isny maulidevi_AACL Bioflux.doc 165K



verianiaprilia verianiaprilia <verianiaprilia@almaata.ac.id>

AACL Bioflux

2 pesan

Eni Kovacs <ek.bioflux@gmail.com> Kepada: verianiaprilia@almaata.ac.id 15 Oktober 2020 pukul 23.49

Dear Dr. Veriani Aprilia,

Thank you for your interest in publishing with us.

I will be your editor and I will stay at your disposition for all the aspects concerning your manuscript.

The editorial team will assess your manuscript and I will get back to you as soon as possible with the result.

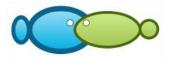
--Kind regards, Editor AACL Bioflux Eniko Kovacs

verianiaprilia verianiaprilia <verianiaprilia@almaata.ac.id> Kepada: Eni Kovacs <ek.bioflux@gmail.com>

Dear Kovacks,

Thank you for your providing us to be our editor. I am waiting for your feed back.

Regards, Aprilia [Kutipan teks disembunyikan] 16 Oktober 2020 pukul 08.07



Submission letter

Article title: The residue of formalin in catfish (*Clarias gariepinus*) after a short period processing and storage

Name of the authors: Isny Maulidevi, Nurul Andriyani, Ryan Salfarino, Veriani Aprilia

Hereby I would like to submit the manuscript entitled "**The residue of formalin in catfish** (*Clarias gariepinus*) **after a short period processing and storage**" to Aquaculture, Aquarium, Conservation & Legislation - International Journal of the Bioflux Society.

This manuscript was not submitted or published to any other journal. The authors declare that the manuscript is an original paper and contain no plagiarised text. All authors declare that they are not currently affiliated or sponsored by any organization with a direct economic interest in subject of the article. My co-authors have all contributed to this manuscript and approve of this submission.

Corresponding author Dr. Veriani Aprilia

Date October 1st, 2020

The residue of formalin in catfish (*Clarias* gariepinus) after a short period processing and storage

¹Isny Maulidevi, ¹Nurul Andriyani, ¹Ryan Salfarino, ¹Veriani Aprilia

Department of Nutrition, Faculty of Health Sciences, Universitas Alma Ata, Jl. Brawijaya 99, Yogyakarta 55183, Indonesia. Corresponding author: verianiaprilia@almaata.ac.id

Abstract. Fresh fish is a highly perishable product due to its high protein content. A preservative may be needed to inhibit it. Nowadays, many fishes were reported to be exposed to formalin. This study aimed to determine the effect of short period processing (washing, frying, boiling in short-period) and storage (in the room and freezing temperature) on the residue of formalin in catfish, as a food model. Catfish were divided into 8 groups, e.g. before treatment, washing in airflow for 1 minute, frying in 200°C for 8 minutes, boiling in 100°C for 7 minutes, storage at room temperature (29±1°C) for 24 and 48 hours, storage at freezing temperature (-3°C) for 24 and 48 hours. Before treatment, the catfish was soaked in 1480 ppm of formalin solutions. The residue of formalin was measured at the end of each treatment by using a spectrophotometer. Data were then analyzed with analysis of variance with SPSS software. The results showed that there were significant effects of processing and storage treatment on the formalin residue of catfish. The formalin residue of all treatments was lower than that of before treatments. The frying process and storage at freezing temperatures were effective treatments to reduce the formalin residue.

Keyword: formalin, fish, processing, storage

Background. Fresh fish is a highly perishable product due to its high protein content for microbial activity (Wijayanti & Lukitasari, 2016). A preservative maybe needs to keep the freshness, inhibit the decay, and longer the shelf life (Antoni, 2010). Nowadays, there are many kinds of food-grade preservatives available in the market. Unfortunately, formalin is still frequently found in food products, including fresh fish. The main reason for the use among fishermen is cheaper, easy to find, and its effectivity to preserve the product (Male et al., 2017).

Formalin is a forty percent solution of formaldehyde in water. This compound is detected in the animal body as an intermediary metabolite. It also uses for several purposes, such as in medicine for specimen preservation and the treatment of hemorrhage (Pandey *et al.*, 2000). The preservatives activity of formalin because of its ability to react with protein generating methylene compound which is more difficult to be decayed by microbes (Ichya'uddin, 2016). In low concentration, formalin is not toxic for humans, but in higher concentrations, it had negative effects such as acute toxicity and death (Pandey *et al.*, 2000). It also decreased the levels of body antioxidants that resulting in liver damage (Yulisa et al., 2014). Other studies also reported the cases of eczema, eye irritation, respiratory tract irritation, cancer, increased the growth rate of malignant neoplastic disease, and denaturation of DNA (Suwanaruang, 2018; Wilianto & Yudianto, 2013). Therefore, its uses are forbidden.

Formalin solution was not stable during storage and exposed to high temperatures (Kaneko et al., 1977; Laksmiani et al., 2015). Those are caused by desolvation or evaporation of formalin during washing, boiling, or frying (Fellows, 2015; Harmita, 2016; Muntaha et al., 2015; Sundari et al., 2015) and enzymatic process in storage (Rachmawati et al., 2007). Formalin that was exposed to the fish decreased up to 43-53% after 5 times rinsing in 250 mL of distilled water and boiling for 30 minutes (Yusuf et al., 2015). Other studies were also reported that soaking in warm water and frying decreased the concentration of formalin up to 63,27% and 83.03% (Levita et al., 2010). In daily practices, fish is usually washed in flowing water before processing, continued to frying/boiling in a shorter time. However, there was no study explained the influences of these processing and storage on the concentration of formalin.

The purpose of this study was to know the effect of processing in a short time and storage (room temperature and freezing) on the residue of formalin. This study used catfish as a food model.

Material and Method

Material. The study used fresh catfish, purchased from the traditional market in Yogyakarta. Another ingredients used were formalin solution and Schiff's reagent. Formalin solution was made from 4 mL of formalin 37% in 1,000 mL of distilled water (Rachmawati, Riyanto, and Ariyani, 2007).

Method. The study was divided into two sections, i.e. 1) study the effect of processing and 2) storage treatment. Fresh catfish were slaughtered, eviscerated, cleaned, and soaked in 1,480 ppm formalin solution (Rachmawati et al., 2007; Sugiarti et al., 2014). For the first study, catfish were divided into four groups, i.e.: controls (before treatment), washing, frying, and boiling treatments. Washing was done by watering the samples with running water for 1 minute, while frying was done by frying the samples in oil 200°C for 8 minutes. Boiling treatment was done by putting the samples into boiled water (100°C) for 7 minutes (Sugiarti, Anggo, and Riyadi, 2014). For the second study, storage treatments, the catfish was also divided into four groups, they were room temperature storage (29°C) for 24 hours and 48 hours and freezing temperatures storage (-3°C) for 24 hours and 48 hours (Handayani et al., 2010). At the end of treatments, the concentration of formalin in samples was analyzed using a spectrophotometer. As the preparation step, a 5 g sample was dissolved in 25 mL of distilled water. One mL of supernatant was mixed with 2 mL of Schiff's reagent, then heated in a water bath at 60°C for 5 minutes. It was incubated for 10 minutes before measuring the absorbance at 520 nm (Shita, 2016; Mudaffar, 2018).

Statistical Analysis. Data were presented in mean±SD and analyzed with One-Way analysis of variance (ANOVA) to know the differences among all processing treatments and the effect of storage periods on the concentration of formalin; paired sample T-test to know the differences between the same samples before and after processing treatments; independent sample t-test to know the effect of temperature at the same storage period, and two-way ANOVA to know the combination effect of storage temperature and period on the concentration of formalin. All data were analyzed with SPSS software (version 16.0 for Windows; SPSS Inc., Chicago, IL, USA) at the 5% significance level.

Results. *Effect of processing on the formalin residue of catfish.* Table 1 shows that all treatments had significant effects on the residue of formalin (p<0.05). It can be seen from the p-value between the same samples before and after treatments. Among all treatments, washing shows the lowest residue of formalin (p<0.05). Both boiling and frying had a similar residue of formalin (p>0.05). The higher residue of formalin in boiling and frying may be due to heating exposure.

					Table 1.
	Effect of process	ing on the formali	n residue of cat	fish	
	Residue of for	malin (ppm)	Δ residue of	p-v	alue
Processing treatments	Before treatment	After treatment	formalin before and after treatment (%)	Among all samples after treatments	Between the same samples before and after treatments
Washing	1,420.17±66.94 ^c	310.42±77.61 ^b	78.14	0.01	0.00
Frying	1,420.17±66.94 ^c	14.18 ± 6.69^{a}	99.01		0.02
Boiling	1,420.17±66.94 ^c	27.50±9.11ª	98.10		0.02

Note: Mean \pm SD in the same column with different superscripts are significantly different (p<0.05). p-value among all samples was obtained from one-way ANOVA analyses,

while p-value between the samples before and after treatments were obtained from paired T-test analyses.

Effect of storage temperatures and periods on the formalin residue of catfish. Table 2 shows the effect of storage temperatures, periods, and the combination of both on the concentration of formalin. Based on this data, the storage period influenced the residue of formalin (p<0.05). It could be seen from the decrease of formalin residue during 24 hours of storage. However, the decrease did not significantly different when the storage continued to 48 hours (p>0.05).

On the other hand, there were no differences in the residue of formalin at room and freezing temperature. It was indicated that the temperature of storage did not affect the formalin residue (p>0.05). Although there were differences in statistical results between the effect of temperature and periods of storage, the combined effect of both variables showed no impact on the concentration of formalin (p>0.05)

Tabel	2.
-------	----

Effect of storage temperature and period on the formalin residue of catfish	
---	--

Temperature	Periods (t) (hours)				p-value	
(T)	0	24	48	Т	t	T*t
Room (29°C)	1,420.17±66.94 ^{a1}	52.18±28.78 ^{a2}	40.64±3.93 ^{a2}	0.20	0.00	0.76
Freezing (-3°C)	1,420.17±66.94 ^{a1}	39.04±7.07 ^{a2}	13.65±4.23 ^{a2}			

Note: Mean±SD in the same column with different alphabetical superscripts (a, b, or c) are significantly different (p<0.05) and so are in the same row with different number superscript (1,2, or 3). The p-value for T, t, and T*t was obtained from independent T-test, one-way ANOVA, and two-way ANOVA, respectively.

Discussion

The residue of formalin before treatment. The residue of formalin before and after treatment (either fish processing or storage) was significantly different (Table 1 and Table 2). In this study, 1,480 ppm of formalin solution was added to the catfish and submerged for 60 minutes. The 60 minutes submersion was chosen because the aldehyde compound may be readily bonded to the amino acids of fish in the small amount (Rahmadhani et al, 2017; Baroroh & Kresnadipayana, 2016). Formalin would attack lysin for the first time, followed by histidine and tyrosine, which led to the formation of methylene compound (Ichya'uddin, 2016). The protein-containing methylene compounds could not be digested by living things and microbes, especially pathogens, so that the shelf life would be longer (Purawisastra & Sahara, 2016). The beneficial bacteria in the fish may also be killed by dehydration mechanism as another effect of the existence of formalin in the body (Mudzkirah, 2016).

The effect of processing on the formalin residue of catfish. The residue of formalin reduced to 78.14% during washing treatment. The reduction was significant (p< 0.05). The formalin was easily washed out because of its high solubility in the water, which is 24 x 10⁵ mg L⁻¹ at 20°C (BPOM, 2008; Sugiarti et al, 2014; Yusuf et al., 2015). Another study conducted by Yusuf et al (2015) also showed the same trend, but the reduction was lower (43%). The differences in the level of reduction can be affected by the length of formalin submersion. The longer the submersion time, the smaller the decreasing as there was more formalin bonded with protein in the fish.

In this study, frying also significantly reduced the residue of formalin (p<0.05). The reduction level was 99% that indicated most formalin was disappeared during the frying process. Formalin was included in a volatile compound, especially at the above of its boiling point (Joshi et al., 2015). This study was also the same with Sugiarti et al (2014) that found the reduction level of formalin up to 99.28% after 1 hour of submersion followed by the frying process (no information about the period of frying process) in squid. Frying could denature the protein and also hydrolyze its bonding with formalin (Levita et al., 2010). The high temperature in frying increased the kinetic energy and led to faster movement of the molecules that could damage the molecular bonds (Sugiarti et al., 2014).

Boiling was another heat processing that was done in this study. The results showed that this process also reduced the residue of formalin up to 98.1%. The residue of formalin after treatment was different from the sample after the washing process (p<0.05), but the same with that after the frying process (p>0.05). In the boiling process, the reduction of formalin was higher than that was in only washing or soaking treatment because there was more formalin dissolved in higher temperature water (Annisak, 2019). The amount of loosing would be higher when it combined with the opened pan during the boiling (Kamal et al., 2017).

The effect of storage on the formalin residue of catfish. This research found that only a period of storage that influenced the residue of formalin (p<0.05), while the temperature of storage and combination between temperature and period of storage did not (Table 2). The previous different studies showed that either room temperature or freezing temperature of storage had a similar effect on the decreasing of formalin residue (Jawahar, et al, 2017; Murtini et al., 2014). It may be due to the degradation of protein during both in the room and freezing temperature, resulting in the loss of the bonds between protein and formalin and releasing free compounds. In the room temperature, the protein degradation was more caused by microbial activity (Murtini et al., 2014), while in the freezing temperature was more caused by physical destruction leading to the conformational and functional changes (Yeasmin et al., 2010). Formalin was easier to break down into another compound or reacts with other compounds in the free form (Riyanto et al., 2006; Rachmawati et al., 2007).

Conclusion. This research concluded that short time process and storage treatments decreased the concentration of formalin. Heat involvement made the decrease of concentration became more effective. In the storage study, the residue of formalin was influenced significantly by the period of storage, mainly during 24 hours of storage.

Thanks to. The authors are grateful to the Indonesia Ministry of Research, Technology, and Higher Education fro providing the financial support in pursuing this research project (Program Kreativitas Mahasiswa Kemeristekdikti 2019 Program).

Reference

- Annisak S. K., Indrivanti N. Y., and Mulyani B., 2019 [Constructive Controversy and Incubation Presented According to The Representation of Tetrahedral Chemical Learning According to Critical Thinking Ability]. Jurnal Inovasi Pendidikan IPA, 5(1), pp. 10–22. [in Indonesian].
- Antoni S, 2010 [Analysis of Formaldehyde Content on Salty Fish by Spectrophotometri Method in Tampan District Pekanbaru]. Analisa Kandungan Formalin Pada Ikan Asin dengan Metoda Spektrofotometri di Kecamatan Tampan Pekanbaru. Universitas Islam Negeri Sultan Syarif Kasim Riau. [in Indonesian].
- Baroroh R. D., and Kresnadipayana D., 2016 [The Effect of Variety Consentration of Starfruit Juice (Averrhoa bilimbi L .) on soaked tofu for Formaldehyde Reduction]. [in Indonesian].

BPOM 2008 [Formaldehyde]. Jakarta: Badan POM RI. [In Indonesian].

Fellows P. J., 2015 [Food Processing Technology]. Jakarta: EGC. [In Indonesian].

Handayani A., Alimin., and Rustiah W. O., 2010 [The Effect of Low Temperature Storage (Freezer -3oC) on Water and Fat Content in Lemuru Fish (Sardinella longiceps)]. Al-Kimia, (3), pp. 64–75. [In Indonesian].

Harmita 2016 [Determination Substance and Availability of Pharmaceuticals Content]. Jakarta: EGC. [in Indonesian].

- Ichya'uddin M., 2014 [Analysis of Formaldehyde Content and Organoleptic Test on Salty Fish in Several Traditional Markets in Tuban]. Universitas Islam Negeri Maulana Malik Ibrahim. [in Indonesian].
- Jawahar L. S., et al., 2017 Retention of Residual Formaldehyde in Treated Indian Mackerel (Rastrelliger kanagurta) Under Iced Storage and Related Food Safety Concern, Indian J. Fish 64(4), pp. 87–93. doi: 10.21077/ijf.2017.64.4.61228-12.
- Joshi et al., 2015 Formaldehyde content of selected fish from the wet markets of Kathmandu valley, International Food Research Journal, 22(4).
- Kamal A. S., et al.. 2017 Time and Temperature Effect on the Residual Concentration of Formaldehyde in Formalin Treated Samples of Labeo rohita, in International Conference on Food Security and Nutrition.
- Levita, J., et al., 2010 The Effect Of Soaking, Washing And Frying On The Concentration Of Formaldehyde In Sange Belah Salty Fish, Jurnal Ilmu-ilmu Hayati dan Fisik 12(1), pp. 31–34.
- Male Y. T., Letsoin L. I., and Siahaya N. A., 2017 [Analysis of Formaldehyde Content on Noodle in Several Area in Ambon]. Jurnal Kementerian Perindustrian 13(2), pp. 5– 10. doi: 10.29360/mb.v13i2.3530. [in Indonesian].
- Mudaffar R. A., 2018 [Qualitative and Quantitative Test of Formaldehyde on Apple, Grape, and Litchi for sale in Makassar]. Jurnal Perbal 6(3). [in Indonesian].
- Mudzkirah, I. (2016) [Identification of Borax and Formalin on Snack Food in UIN Alaudin Makassar Canteen]. UIN Alauddin Makassar. [in Indonesian].
- Muntaha A., Haitami., and Hayati N., 2015 [Comparison of Reduction Formaldehyde Level on Boiled and Hot Water Soaked Tofu]. Medical Laboratory Technology Journal 1(2), pp. 84–90. [in Indonesian].
- Murtini J. T., et al., 2014 [Development of Formaldehyde on Some Kinds of Seawater Fish During Storage in Crushed Ice. JPB Perikanan 9(2), pp. 143–151. [in Indonesian].
- Pandey C. K., et al., 2000 Toxicity of ingested formalin and its management Human and Experiment Toxicology 19(November 1999), pp. 360–366.
- Purawisastra S., and Sahara E., 2011 [Absorption to Formalin by Certain Foodstuff and Disappearance Through Hot Water Soaked]. PGM 34(1), pp. 63–74. [in Indonesian].
- Rachmawati N., Riyanto R., and Ariyani F., 2007 [Identification of Formaldehyde on Kerapu Macan Fish (Ephinephelus fuscoguttatus) During Cold Storage]. Jurnal Pascapanen dan Bioteknologi Kelautan dan Perikanan 2(2), pp. 137–145. [in Indonesian].
- Rahmadhani F., Safrida., and Djufri., 2017 [The Effect of Soaking a Various of Solution for Reduction Formalin Level on Kembung Salty Fish (Scomber canagorta) in Lambaro Traditional Market Aceh]. Jurnal Ilmiah Mahasiswa Keguruan dan Ilmu Pendidikan Unsyiah. [in Indonesian].
- Riyanto R., Kusmarwati A., and Dwiyitno., 2006 [Identification of Formaldehyde Foorming on Kerapu Fish (Epinephelus fuscoguttatus) During Room Temperature Storage]. Jurnal Pascapanen dan Bioteknologi Kelautan dan Perikanan 1(2), pp. 111–116. [in Indonesian]
- Shita A. E., 2016 [Selectivity of Spectrophotometri Analysis with Schiff's Reagen]. Universitas Negeri Yogyakarta. [in Indonesian].
- Sugiarti M., Anggo A. D., and Riyadi P. H., 2014 [The Effect of Soaking in Undercooking Temperature and Cooking Method for Reduction Formalin Level on Cuttlefish]. Jurnal Pengolahan dan Bioteknologi Hasil Perikanan 3(2), pp. 90–98. [in Indonesian].

- Sundari D., Almasyhuri., and Lamid A., 2015 [The Effect of Cooking Process for Nutrition Composition of Protein Source]. Media Litbangkes 25(4), pp. 235–242. [in Indonesian].
- Suwanaruang T., 2018 Formalin Contaminated in Seafood and Frozen Meat at Somdet Market , Kalasin Province. Journal of Environmental Protection 9, pp. 1286–1293. doi: 10.4236/jep.2018.912080.
- Wijayanti N. S., and Lukitasari M., 2016 [Analysis of Formalin Level and Organoleptic Test in Salty Fish in Madiun Market]. Jurnal Florea 3(1), pp. 59–64. [in Indonesian].
- Wilianto W., and Yudianto A., 2013 [The Effect of Formalin Exposure on Various Level for DNA Psoas Major by PCR Lokus CSF1PO, D5S818, D13S317, D21S11 Analysis]. Jurnal Kedokteran Forensik Indonesia 15(1), pp. 11–16. [in Indonesian].
- Yeasmin T., et al., 2010 Quality Changes in Formalin Treated Rohu Fish (Labeo rohita, Hamilton) During Ice Storage Condition. Asian Journal of Agricultural Science 2(4), pp. 158–163.
- Yulisa N., Asni E., and Azrin M., 2014 [Formaldehyde Test of Gurame Fish in Pekanbaru Traditional Market]. 1(2). [in Indonesian].
- Yusuf Y., et al., 2015 [The Effect of Various Treatment for Reduction Formalin Level on Fish by Spectrophotometri Method]. J. Ris. Kim 8(2), pp. 182–188.



Preliminary acceptance (Lolos untuk Direviu): 3 November 2020

Preliminary acceptance

8 pesan

Eni Kovacs <ek.bioflux@gmail.com> Kepada: verianiaprilia@almaata.ac.id 3 November 2020 pukul 03.21

Dear Veriani Aprilia,

We would like to inform you that after the preliminary evaluation your paper is qualified for processing (reviewing). However, the manuscript does not respect the AACL formatting requirements. Please find the model on our website and <u>replaceeachsectionwithyourtext</u>, leaving the footer exactly as it is, including the page numbering. Also, the references from the text and from the reference list must be adjusted according to the requirements.

Before proceeding to the review please consider that a processing publication fee of 250 USD is required. The average duration of the publication process is 10 weeks, but it can be reduced in exchange of a priority tax of 50 USD (<u>http://www.bioflux.com.ro/journal/</u>).

In case the manuscript is not published, the author or his/her institution is reimbursed (exceptions are made in the cases of poor feedback from authors or withdrawal/rejection due to multiple submissions).

Concerning the payment procedure, you will find all the details bellow and I would like to kindly ask you to send me the scanned receipt of the payment in order to start the publishing process. Upon reception of your payment receipt we will send you the invoice.

1st payment option: Beneficiary: Bioflux SRL City: Cluj-Napoca, Country: Romania, European Union;

SWIFT CODE of the bank: BTRLRO22

Account USD: RO68BTRL01302202L28614XX

Bank: BANCA TRANSILVANIA

Important! When bank transfer is used to pay a publication fee, please choose the right option, which is "US"! (among the three options you have: Ben, Our, Us).

The option "Us" means that **all the transfer fees are at your charge**. Please note that in case you do not specify the "Us" option, the payment will not be valid.

2nd payment option:

via Paypal. If you can process such a payment we can start the online form.

If you prefer this option please let us know and we will send you the invoice via Paypal and afterwards you can process the payment.

For this payment option you need a credit card (international, non-iranian card). Please take into account that all bank taxes are in your concern.

We recommend not to send your manuscript to more than one editor, in order to avoid being double charged.

Do not hesitate to contact me for any kind of questions related to the process.

Thank you for publishing with us!

Email Universitas Alma Ata - Preliminary acceptance

Kind regards, Editor AACL Bioflux Eniko Kovacs, PhD

Visit our journals:

Aquaculture, Aquarium, Conservation & Legislation <u>www.bioflux.com.ro/aacl</u> AACL Bioflux Advances in Environmental Sciences <u>www.aes.bioflux.com.ro</u> AES Bioflux Human & Veterinary Medicine <u>www.hvm.bioflux.com.ro</u> HVM Bioflux Advances in Agriculture & Botanics <u>www.aab.bioflux.com.ro</u> AAB Bioflux Animal Biology & Animal Husbandry <u>www.abah.bioflux.com.ro</u> ABAH Bioflux Extreme Life, Biospeology & Astrobiology <u>www.elba.bioflux.com.ro</u> ELBA Bioflux Porcine Research <u>www.porc.bioflux.com.ro</u> Porc Res Rabbit Genetics <u>www.rg.bioflux.com.ro</u> Rabbit Gen Poeciliid Research <u>www.pr.bioflux.com.ro</u> Poec Res



Preliminary_acceptance_AACL_Aprilia.pdf 157K

verianiaprilia verianiaprilia <verianiaprilia@almaata.ac.id> Kepada: Eni Kovacs <ek.bioflux@gmail.com>

Dear Kovacs,

Thank you for your nice news for us about the preliminary acceptance. Would i know, how long will the article be published if i prefer the priority option? thank you for your information

Regards Veriani Aprilia [Kutipan teks disembunyikan]

verianiaprilia verianiaprilia <verianiaprilia@almaata.ac.id> Kepada: Eni Kovacs <ek.bioflux@gmail.com>

Dear Kovacs,

Thank you for your great information about the preliminary acceptance. Please let me know, how long does the processing of publication with the priority term? We also prefer to transfer via pay pal. Would you like to give me the account number? Thank you for your great assistance.

Regards, Veriani Aprilia

Pada tanggal Sel, 3 Nov 2020 pukul 03.21 Eni Kovacs <<u>ek.bioflux@gmail.com</u>> menulis: [Kutipan teks disembunyikan]

Eni Kovacs <ek.bioflux@gmail.com>

Kepada: verianiaprilia verianiaprilia <verianiaprilia@almaata.ac.id>

4 November 2020 pukul 00.35

Dear Veriani Aprilia,

Thank you for your email. First of all, the manuscript must respect the formatting requirements of the journal, for this reason we kindly asked to use the model from our website.

We can't provide an exact publication duration, it depends on several factors such as: the quality of the paper, the reviewers' feedback, the authors response time etc. But in the case of a priority procedure, we do our best to publish as

3 November 2020 pukul 09.05

3 November 2020 pukul 10.36

7/18/23, 1:43 PM

soon as possible.

If you would like to pay via paypal please inform us about which payment fee do you prefer.

Thank you. [Kutipan teks disembunyikan] --[Kutipan teks disembunyikan]

verianiaprilia verianiaprilia <verianiaprilia@almaata.ac.id> Kepada: Eni Kovacs <ek.bioflux@gmail.com>

Dear Kovacs,

Here is our manuscript. Please give me the invoice or account of paypal. We prefer the priority terms. Thank you for your great work.

Regards Veriani Aprilia [Kutipan teks disembunyikan]

isny maulidevi_AACL Bioflux2.doc 155K

Eni Kovacs <ek.bioflux@gmail.com> Kepada: verianiaprilia verianiaprilia <verianiaprilia@almaata.ac.id>

Thank you.

The editor in chief will send you the information.

Kind regards! [Kutipan teks disembunyikan]

verianiaprilia verianiaprilia <verianiaprilia@almaata.ac.id> Kepada: Eni Kovacs <ek.bioflux@gmail.com>

Dear Kovacs,

I have paid the processing publication fee. The invoice is attached. I hope the reviewing process is already started. Thank you

Regards, Veriani Aprilia [Kutipan teks disembunyikan]

Invoice - 0576_BUKTI PEMBAYARAN.pdf

Eni Kovacs <ek.bioflux@gmail.com> Kepada: verianiaprilia verianiaprilia <verianiaprilia@almaata.ac.id>

Thank you for the payment.

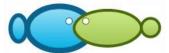
We will further process your manuscript and contact you when we receive the reviewers feedback.

4 November 2020 pukul 12.49

6 November 2020 pukul 02.17

13 November 2020 pukul 22.58

14 November 2020 pukul 03.46



Bioflux (publishing house) 54 Ceahlau Street, Cluj-Napoca 400488, Romania, European Union

Certificate/Letter of preliminary acceptance

This certificate shows that your paper: The residue of formalin in catfish (*Clarias gariepinus*) after a short period processing and storage

Authors: Isny Maulidevi, Nurul Andriyani, Ryan Salfarino, Veriani Aprilia

was preliminary accepted for publication, with revision, in volume 13 (2020) of the scientific/academic journal: Aquaculture, Aquarium, Conservation & Legislation – International Journal of the Bioflux Society.

AACL Bioflux is covered by Thomson ISI Web of Knowledge via:

- Zoological Record (Biosis) and
- CAB Abstracts (CABI)

Thank you for publishing with us!

Sincerely yours, Editor Researcher Eniko Kovacs, PhD



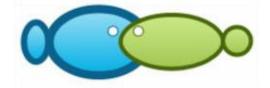
verianiaprilia verianiaprilia <verianiaprilia@almaata.ac.id>

Invoice from Bioflux SRL (0576)

1 pesan

Bioflux SRL <service@paypal.com> Balas Ke: Bioflux SRL <zoobiomag2004@yahoo.com> Kepada: "verianiaprilia@almaata. ac. id" <verianiaprilia@almaata.ac.id> 13 November 2020 pukul 15.15

Hello, verianiaprilia@almaata.ac.id



Here's your invoice

Bioflux SRL sent you an invoice for \$323.00 USD

Due date: December 13, 2020

View and Pay Invoice

Note from Bioflux SRL

The extra fee is for bank transfer





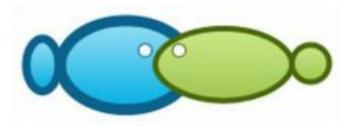
PayPal is committed to preventing fraudulent emails. Emails from PayPal will always contain your full name. Learn to identify phishing.

Please don't reply to this email. To get in touch with us, click Help & Contact.

Not sure why you received this email? Learn more

Copyright © 1999-2020 PayPal, Inc. All rights reserved. PayPal is located at 2211 N. First St., San Jose, CA 95131.

PayPal PPC000977:1.7.1:eb5d07b22a219



Bioflux SRL

Ioan Valentin Petrescu-Mag 54 Ceahlau Street Cluj-Napoca, Cluj 400488 Romania

Phone: +40 0040744470794 zoobiomag2004@yahoo.com www.bioflux.com.ro

INVOICE

Paid

Invoice #: 0576 Invoice Date: Nov 13, 2020 Due date: Dec 13, 2020

> Amount due: **\$0.00**



Bill To:

verianiaprilia@almaata.ac.id

Description	Quantity	Price	Amount
Publication fee The residue of formalin in catfish (Clarias gariepinus) after a short period processing and storage. Isny Maulidevi, Nurul Andriyani, Ryan Salfarino, Veriani Aprilia	1	\$300.00	\$300.00
		Subtotal	\$300.00
			\$23.00
		Total	\$323.00
		Amount paid	-\$323.00
	A	mount due	\$0.00 USD

Notes

The extra fee is for bank transfer



verianiaprilia verianiaprilia <verianiaprilia@almaata.ac.id>

Pemberitahuan Hasil Reviu Artikel: 29 November 2020

Revision

2 pesan

Eni Kovacs <ek.bioflux@gmail.com> Kepada: verianiaprilia verianiaprilia <verianiaprilia@almaata.ac.id> 29 November 2020 pukul 17.56

Dear Dr. Aprilia,

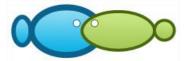
Regarding your manuscript submitted to AACL Bioflux, the editorial team has some requests prior to acceptance (please see the attachment).

Please note: Always operate corrections/additions (or deletions) in the manuscript we are sending you, by **highlighting with a bright color** (for an easy identification). We never work on the manuscript you send back, just identify the corrections and operate them on our document (to avoid any undesirable accidental operations like changed page set up, otherwise the editors have to start all the work from the beginning, and we cannot ask them to re-check every manuscript word by word to identify unmarked modifications).

Thank you!

Kind regards, Editor AACL Bioflux Eniko Kovacs

Aprilia_Clarias gariepinus_wip.doc 183K



The residue of formalin in catfish (*Clarias* gariepinus) after a short period processing and storage for short periods

Isny Maulidevi, Nurul Andriyani, Ryan Salfarino, Veriani Aprilia

Department of Nutrition, Faculty of Health Sciences, Alma Ata University, Jakarta, Indonesia. Corresponding author: V. Aprilia, verianiaprilia@almaata.ac.id

Abstract. Fresh fish is a highly perishable product due to its high protein content. A preservative may be needed to inhibit it. Nowadays, many fishes were reported to be exposed to formalin. This study aimed to determine the effect of short period processing (washing, frying, boiling in short-period) and storage (in-at the room and at freezing temperatures) on the residue of formalin in catfish, as a food model. Catfish, <u>Clarias gariepinus</u>, were divided into 8 groups, e.g. before treatment, washing in airflow for 1 minute, frying in 200°C for 8 minutes, boiling in 100°C for 7 minutes, storage at room temperature ($29\pm1°C$) for 24 and 48 hours, storage at freezing temperature (-3°C) for 24 and 48 hours. Before treatment, the catfish was soaked in 1,480 ppm of formalin solutions. The residue of formalin was measured at the end of each treatment by using a spectrophotometer. Data were then analyzed with analysis of variance with SPS software. The results showed that there were significant effects of all treatments was lower than that of before treatments. The frying processing and storage at freezing temperatures were the most effective treatments to in reducinge the formalin residue. **Key Words**: formalin, fish, processing, storage.]

Introduction. Fresh fish is a highly perishable product due to its high protein content for microbial activity (Wijayanti & Lukitasari 2016). A preservative maybe needs to keep the freshness, inhibit the decay₇ and prolonger the shelf life (Antoni 2010). Nowadays, there are many kinds of food-grade preservatives available in the market. Unfortunately, formalin is still frequently found in food products, including fresh fish-, being used among fishermen The-mainly becausereason for the use among fishermen is cheaper, easy to find, and it is effectivity effective to in preserve-preserving the product, cheaper and accessible (Male et al 2017).

Formalin is a forty percent solution of formaldehyde in water. This compound is detected in the animal body as an intermediary metabolite. It also uses for several purposes, such as in medicine for specimen preservation and the treatment of hemorrhage (Pandey et al 2000). The preservatives activity of formalin because of due to its ability to react with protein generating methylene compounds, which is are more difficult to be decayed by microbes (Ichya'uddin 2016). In low concentrations, formalin is not toxic for humans, but in higher concentrations, it had negative effects such as acute toxicity and death (Pandey et al 2000). It also decreased decreases the levels of body antioxidants that, resulting in liver damage (Yulisa et al 2014). Other studies also reported the cases of eczema, eye irritation, respiratory tract irritation, cancer, increased the growth rate of malignant neoplastic diseases; and denaturation of DNA (Suwanaruang 2018; Wilianto & Yudianto 2013). Therefore, its uses are forbidden.

Formalin solution was-is not stable during storage and exposed exposure to high temperatures (Kaneko et al 1977; Laksmiani et al 2015). Those are caused by desolvation or evaporation of formalin during washing, boiling, or frying (Fellows 2015; Harmita 2016; Muntaha et al 2015; Sundari et al 2015) and enzymatic process in storage (Rachmawati et al 2007). Formalin that was exposed to thein contact with fish decreased up to 43-53% after 5 times rinsing in 250 mL of distilled water and boiling for 30 minutes

AACL Bioflux, 2020, Volume 13, Issue X. http://www.bioflux.com.ro/aacl

1090

Commented [WU1]: According to the international standards in scientific writing it is desirable to display 5 key words which do not appear in the title. This will increase findings via key words and implicit citations.

Commented [WU2]: Not listed in the references. Commented [WU3]: Not listed in the references. (Yusuf et al 2015). Other studies were also reported that soaking in warm water and frying decreased the concentration of formalin up to $63_{7_a}27\%$ and 83.03%, respectively (Levita et al 2010). In daily practices, fish is usually washed in flowing water before processing, continued-then to fryingfryed/boiling-boiled in afor a short_er-time. However, there was no study explained explaining the influences of these processing and storage methods on the concentration of formalin.

The purpose of this study was to <u>know_identify</u> the effect of processing <u>in_for</u> a short time and storage (<u>at</u> room <u>and freezing</u> temperature<u>s</u>-<u>and freezing</u>) on the residue of formalin. This study used catfish as a food model.

Material and Method

Material. The study used fresh catfish, purchased from the traditional market in YogyakartaJakarta. <u>TheAn</u> other <u>used</u> ingredients <u>used</u> were formalin solution and Schiff's reagent. Formalin solution was made from 4 mL of formalin 37% in 1,000 mL of distilled water (Rachmawati et al 2007).

Method. The study was divided into two sections, i.e. 1) study_analyzing_the effect of processing and 2) analyzing the effect of storage treatment. Fresh catfish were slaughtered, eviscerated, cleaned, and soaked in 1,480 ppm formalin solution (Rachmawati et al 2007; Sugiarti et al 2014). For the first study, catfish were divided into four groups, i.e.: controls (before treatment), washing, frying, and boiling treatments. Washing was done by watering the samples with running water for 1 minute, while frying was done by frying the samples in oil at 200°C for 8 minutes. Boiling treatment was done by putting the samples into boiled water (100°C) for 7 minutes (Sugiarti et al 2014). For the second study, storage treatments, the catfish was also divided into four groups, they: were room temperature storage (29°C) for 24 hours and 48 hours and freezing temperatures storage (-3°C) for 24 hours and 48 hours (Handayani et al 2010). At the end of treatments, the concentration of formalin in samples was analyzed using a spectrophotometer. As the preparation step, a 5 g sample was dissolved in 25 mL of distilled water. One mL of supernatant was mixed with 2 mL of Schiff's reagent, then heated in a water bath at 60°C for 5 minutes. It was incubated for 10 minutes before measuring the absorbance at 520 nm (Shita 2016; Mudaffar 2018).

Statistical analysis. Data were presented in mean±SD and analyzed with: (1) one-way analysis of variance (ANOVA) to know_determine_the differences among all processing treatments and the effect of storage periods on the concentration of formalin; (2) paired sample T-test to determineknow the differences between the same samples before and after processing treatments; (3) independent sample t-test to know the effect of temperature at the same storage period, and (4) two-way ANOVA to determineknow the concentration of formalin. All data were analyzed with SPSS software (version 16.0 for Windows; SPSS Inc., Chicago, IL, USA) at the 5% significance level.

Results

Effect of processing on the formalin residue of catfish. Table 1 shows that all treatments had significant effects on the residue of formalin (p<0.05). It can be seen from the p-value between the same samples before and after treatments. Among all treatments, washing shows the <u>lowest highest</u> residue of formalin (p<0.05). Both boiling and frying had a<u>n almost</u> similar residue of formalin (p>0.05). The <u>higher-lower</u> residue of formalin in boiling and frying may be due to heating exposure.

Commented [WU4]: The text was aligned to table 1.

AACL Bioflux, 2020, Volume 13, Issue X. http://www.bioflux.com.ro/aacl

1091

Table 1

Effect of processing on the formalin residue of catfish

	Residue of formalin (ppm)		Aracidua	p-value		
Processing treatments	<i>Before treatment</i>	After treatment	∆ residue of formalin before and after treatment (%)	Among all samples after treatments	<i>Between the same samples before and after</i>	
			(%)		treatments	
Washing	1,420.17±66.94°	310.42±77.61 ^b	78.14		0.00	
Frying	1,420.17±66.94°	14.18±6.69ª	99.01	0.01	0.02	
Boiling	1,420.17±66.94°	27.50±9.11 ^a	98.10		0.02	

Mean \pm SD in the same column with different superscripts are significantly different (p<0.05); p-value among all samples was obtained from one-way ANOVA analyses, while p-value between the samples before and after treatments were obtained from paired T-test analyses.

Effect of storage temperatures and periods on the formalin residue of catfish. Table 2 shows the effect of storage temperatures, <u>of periods</u>₇ and <u>of their</u> combination—of both on the concentration of formalin. Based on this data, the storage period influenced the residue of formalin <u>decrease (p<0.05)</u>—<u>during the first 24 hoursIt could be seen from the decrease of formalin residue during 24 hours of storage. However, but after a storage period of 48 hours the decrease <u>did</u>—was not significantly different <u>when the</u>,—storage continued to 48 hours (p>0.05).</u>

On the other hand, there were no differences in the residue of formalin <u>stored</u> at room and freezing temperature. <u>It was indicated</u>, <u>suggesting</u> that the temperature of storage did not affect the formalin residue (p>0.05). Although there were differences in <u>the</u> statistical results between the <u>effect of</u> temperature <u>effects</u> and <u>the storage</u> periods <u>of storage effects</u>, the combined effect of both variables showed no impact on the concentration of formalin (p>0.05).

Table 2

Effect of storage temperature and period on the formalin residue of catfish

Temperature	Per	riods (t) (hours)			p-value	9
<i>(T)</i>	0	24	48	Т	t	T*t
Room (29°C)	1,420.17±66.94 ^{a1}	52.18±28.78 ^{a2}	40.64±3.93 ^{a2}	0.20	0.00	0.76
Freezing (-3°C)	1,420.17±66.94 ^{a1}	39.04±7.07 ^{a2}	13.65±4.23 ^{a2}	0.20	0.00	0.76
Mean±SD in the same column with different alphabetical superscripts (a, b, or c) are significantly different						

(p<0.05) and so are in the same row with different number superscript (1, 2, or 3). The p-value for T, t, and T*t was obtained from independent T-test, one-way ANOVA, and two-way ANOVA, respectively.

Discussion

The residue of formalin before treatment. The residue of formalin before and after treatment (either fish processing or storage) was significantly different (Table 1 and Table 2). In this study, the catfish specimens were submerged for 60 minutes in 1,480 ppm of formalin solution of 1,480 ppm.was added to the catfish and submerged for 60 minutes. The 60 minutes submersion period was chosen because the aldehyde compound may be readily bonded to the amino acids of fish in the <u>a</u> small amount (Rahmadhani et al 2017; Baroroh & Kresnadipayana 2016). Formalin would attack lysin for the first time, followed by histidine and tyrosine, which ledleading to the formation of <u>a</u> methylene compound (Ichya'uddin 2016). The protein-containing methylene compounds could not be digested by living things and microbes, especially pathogens, so that the shelf life would be longer (Purawisastra & Sahara 2016). The beneficial bacteria in the fish may

1092

Commented [WU5]: Please explain and give an interpretation, it does not seem very clear.

Commented [WU6]: 2011 in the reference list.

also be killed by dehydration mechanism, as another effect of the existence of formalin in the body (Mudzkirah 2016).

The effect of processing on the formalin residue of catfish. The residue of formalin was reduced to by 78.14% during washing treatment. The reduction was significant (p<0.05). The formalin was easily washed out because of its high solubility in the water, which is 24 x 105 mg L⁻¹ at 20°C (BPOM 2008; Sugiarti et al 2014; Yusuf et al 2015). Another study conducted by Yusuf et al (2015) also showed the same trend, but the reduction was lower (with only 43%). The differences in the level of reduction can be affected by the length-duration of formalin submersion. The longer the submersion time, the smaller the decreasing decrease, as there was more formalin was bonded with the fish protein in the fish.

In this study, frying also significantly reduced the residue of formalin (p<0.05). The reduction level was 99% that indicated most <u>of the</u> formalin <u>was</u>-disappeared during the frying process. Formalin was included in a volatile compound, especially at the above of-its boiling point (Joshi et al 2015). This study <u>was also the same withconfirmed the</u> results of Sugiarti et al (2014), showing a that found the reduction level of formalin up to 99.28% after 1 hour of submersion followed by the frying process (no information about the period of frying process) in squid. Frying could denature the protein and also hydrolyze its bonding with formalin (Levita et al 2010). The high temperature in frying increased the kinetic energy and led to faster movement of the molecules that could damage the molecular bonds (Sugiarti et al 2014).

Boiling was another heat-<u>Heat</u> processing <u>by boiling</u> that was done in this study. The results showed that this process also reduced the residue of formalin <u>with</u> up to 98.1%, <u>more than</u>. The residue of formalin after treatment was different from the sample after the washing process (p<0.05), but <u>at</u> the same <u>degree</u> with asthat after the frying process (p>0.05). In the boiling process, tThe high reduction of formalin in the boiling process was higher than that was in only washing or soaking treatment because there was more was due to the accelerated formalin dissolved dissolution rate in at higher temperatures water (Annisak 2019). This effect could be enhanced by amount of losing would be higher when it combined_boiling the sample with thein an opened pan during the boiling (Kamal et al 2017).

The effect of storage on the formalin residue of catfish. This research experiment found_demonstrated_that only a-the_period of storage that significantly_influenced the residue of formalin (p < 0.05), while the temperature of storage and the combination between temperature and period of storage did not (Table 2). The previous different studies showed that either a storage at the room temperature or at the freezing temperature of storage had a similar effects on the decreasing of formalin residue decrease (Jawahar et al 2017; Murtini et al 2014), possibly . It may be due to the same degradation level of protein degradationduring both in the room and freezing temperature, resulting in the loss of by breaking the bonds between protein and formalin and releasing free compounds. In At the room temperature, the protein degradation was more essentially caused by the microbial activity, while in at the freezing temperature it was more caused mostly by physical destruction, leading to the conformational and functional changes (Murtini et al 2014; Yeasmin et al 2010). Formalin in the free form is more reactive and can also be more easily degraded was easier to break down into another compound or reacts with other compounds in the free form (Rivanto et al 2006; Rachmawati et al 2007).

Conclusions. This research concluded that short time processing and storage treatments decreased the concentration of formalin. Heat involvement made determined a more effective the decrease of concentration became more effective. In the storage study, the residue of formalin was influenced significantly reduced by the period of storage, mainly during the first 24 hours, then the formalin concentration reduction was insignificant, up to 48 hours of storage.

Commented [WU7]: Please add some details concerning the above mentioned results.

AACL Bioflux, 2020, Volume 13, Issue X. http://www.bioflux.com.ro/aacl 1093

Acknowledgements. The authors are grateful to the Indonesia Ministry of Research, Technology, and Higher Education for providing the financial support in pursuing this research project (Program Kreativitas Mahasiswa Kemeristekdikti 2019 Program).

Reference

- Annisak S. K., Indriyanti N. Y., and Mulyani B., 2019 [Constructive controversy and incubation presented according to the representation of tetrahedral chemical learning according to critical thinking ability]. Jurnal Inovasi Pendidikan IPA 5(1):10–22. [In Indonesian].
- Antoni S., 2010 [Analysis of formaldehyde content on salty fish by spectrophotometri method in Tampan District Pekanbaru]. ... Type of study, Universitas Islam Negeri Sultan Syarif Kasim Riau, <u>No of pages</u>. [In Indonesian].
- Baroroh R. D., and Kresnadipayana D., 2016 [The effect of variety consentration of starfruit juice (*-Averrhoa bilimbi* L-.) on soaked tofu for formaldehyde reduction]. <u>Publisher, Volume, Issue, Pages.</u> [In Indonesian].

BPOM 2008 [Formaldehyde]. Jakarta: Badan POM RI. <u>No of pages</u> ISBN: 979-979-1269-17-19. [In Indonesian]. <u>What does BPOM stands for?</u>

- Fellows P. J., 2015 [Food processing technology]. <u>Publisher, no of pages</u> Jakarta: EGC. ISBN: 979-044-612-0. [In Indonesian].
- Handayani A., Alimin, and Rustiah W. O., 2010 [The effect of low temperature storage (freezer -3oc) on water and fat content in lemuru fish (*Sardinella longiceps*)]. Al-Kimia (3):64–75. [In Indonesian].
- Harmita, 2016 [Determination substance and availability of pharmaceuticals content]. <u>Publisher</u>, Jakarta: EGC. <u>No of pages</u> ISBN: 979-044-773-8. [In Indonesian].
- Ichya'uddin M., 2014 [Analysis of Formaldehyde content and organoleptic test on salty fish in several traditional markets in Tuban]. <u>Publication type</u>, Universitas Islam Negeri Maulana Malik Ibrahim., <u>no of pages</u>. [In Indonesian].
- Jawahar L. S., et al.all the authors must be listed, 2017 Retention of Residual formaldehyde in treated Indian mackerel (*Rastrelliger kanagurta*) under iced storage and related food safety concern, Indian J. Fishno abbreviations 64(4):87– 93. doi: 10.21077/ijf.2017.64.4.61228-12.
- Joshi-<u>all the authors must be listedet al.</u>, 2015 Formaldehyde content of selected fish from the wet markets of Kathmandu valley<u>-</u>__International Food Research Journal 22(4):...pages. ISSN: 1434-1437.
- Kamal A. S., et al..all the authors must be listed, 2017 Time and Temperature effect on the residual concentration of formaldehyde in formalin treated samples of <u>Labeo</u> rohita____in-International Conference on Food Security and Nutrition, <u>pages</u>.
- Levita₇ J., et al.all the authors must be listed, 2010 The effect of soaking, washing and frying on the concentration of formaldehyde in Sange Belah salty fish₇. Jurnal Ilmuilmu Hayati dan Fisik 12(1):31–34.
- Male Y. T., Letsoin L. I., and Siahaya N. A., 2017 [Analysis of formaldehyde content on noodle in several area in Ambon]. Jurnal Kementerian Perindustrian 13(2):5–10. [In Indonesian].
- Mudaffar R. A., 2018 [Qualitative and quantitative test of formaldehyde on apple, grape, and litchi for sale in Makassar]. Jurnal Perbal 6(3).....pages. [In Indonesian].
- Mudzkirah I., 2016 [Identification of borax and formalin on snack food in UIN Alaudin Makassar Canteen]. <u>Type of publication</u>, UIN Alauddin Makassar., <u>no of pages.</u> [In Indonesian].
- Muntaha A., Haitami-, and Hayati N., 2015 [Comparison of reduction formaldehyde level on boiled and hot water soaked tofu]. Medical Laboratory Technology Journal 1(2):84–90. [In Indonesian].
- Murtini J. T., et al.all authors must be listed, 2014 [Development of Formaldehyde on some kinds of seawater fish during storage in crushed ice]. JPB Perikanan 9(2):143–151. [In Indonesian].

AACL Bioflux, 2020, Volume 13, Issue X. http://www.bioflux.com.ro/aacl Formatted: Font: Italic

Formatted: English (United States)

Formatted: Font: Italic

Formatted: Font: Italic

Formatted: Font: Italic

1094

Pandey C. K., et al.all authors must be listed, 2000 Toxicity of ingested formalin and its management. Human and Experiment Toxicology 19(November 1999), pp. 360– 366.

Purawisastra S., and Sahara E., 2011 [Absorption to Formalin by certain foodstuff and disappearance through hot water soaked]. PGM 34(1):63–74. [In Indonesian].

Rachmawati N., Riyanto R., and Ariyani F., 2007 [Identification of formaldehyde on kerapu macan fish (*Ephinephelus fuscoguttatus*) during cold storage]. Jurnal Pascapanen dan Bioteknologi Kelautan dan Perikanan 2(2):137–145. [In Indonesian].

Rahmadhani F., Safrida-, and Djufri-, 2017 [The effect of soaking a various of solution for reduction formalin level on kembung salty fish (*Scomber canagorta*) in Lambaro traditional market Aceh]. Jurnal Ilmiah Mahasiswa Keguruan dan Ilmu Pendidikan Unsyiah, volume, issue, pages. [In Indonesian].

Riyanto R., Kusmarwati A., and Dwiyitno-, 2006 [Identification of formaldehyde foorming on kerapu fish (*Epinephelus fuscoguttatus*) during room temperature storage]. Jurnal Pascapanen dan Bioteknologi Kelautan dan Perikanan 1(2):111–116. [In Indonesian].

Shita A. E., 2016 [Selectivity of spectrophotometri spectrophotometer analysis with Schiff's Reagen]. Type of publication, Universitas Negeri Yogyakarta, no of pages. [In Indonesian].

Sugiarti M., Anggo A. D., and Riyadi P. H., 2014 [The effect of soaking in undercooking temperature and cooking method for reduction formalin level on cuttlefish]. Jurnal Pengolahan dan Bioteknologi Hasil Perikanan 3(2):90–98. [In Indonesian].

Sundari D., Almasyhuri, and Lamid A., 2015 [The effect of cooking process for nutrition composition of protein source]. Media Litbangkes 25(4):235–242. [In Indonesian].

Suwanaruang T., 2018 Formalin contaminated in seafood and frozen meat at Somdet market-, Kalasin Province. Journal of Environmental Protection 9, pp.-:1286–1293-.

Wijayanti N. S., and-Lukitasari M., 2016 [Analysis of formalin level and organoleptic test in salty fish in Madiun Market]. Jurnal Florea 3(1):59–64. [In Indonesian].

Wilianto W., and Yudianto A., 2013 [The effect of formalin exposure on various level for DNA psoas major by PCR Lokus CSF1PO, D5S818, D13S317, D21S11 Analysis]. Jurnal Kedokteran Forensik Indonesia 15(1):11–16. [In Indonesian].

Yeasmin T., et al.all authors must be listed, 2010 Quality changes in formalin treated rohu fish (*Labeo rohita*, Hamilton) during ice storage condition. Asian Journal of Agricultural Science 2(4):158–163.

Yulisa N., Asni E., and Azrin M., 2014 [Formaldehyde test of gurame fish in Pekanbaru Traditional Market]. <u>Publisher1(2): pages</u>. [In Indonesian].

Yusuf Y., et al.all the authors must be listed, 2015 [The effect of various treatment for reduction formalin level on fish by spectrophotometri spectrophotometry method]. J. Ris. KimNo abbreviations 8(2):182–188. [In Indonesian].

Received: 15 October 2020. Accepted: 2020. Published online: 2020. Authors:

This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution and reproduction in any medium, provided the original author and source are credited.

How to cite this article: Maulidevi I., Andriyani N., Salfarino R., Aprilia V.,2020 The residue of formalin in catfish (*Clarias gariepinus*) after a short period processing and storage. AACL Bioflux 13(x):

AACL Bioflux, 2020, Volume 13, Issue X. http://www.bioflux.com.ro/aacl 1095

Formatted: Font: Italic

Formatted: Font: Italic

Formatted: Font: Italic

Formatted: English (United States)

Formatted: Font: Italic

Commented [WU8]: Information missing.



verianiaprilia verianiaprilia <verianiaprilia@almaata.ac.id>

Pengiriman Revisi Artikel: 4 Desember 2020

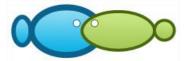
verianiaprilia verianiaprilia <verianiaprilia@almaata.ac.id> Kepada: Eni Kovacs <ek.bioflux@gmail.com> 4 Desember 2020 pukul 00.08

Dear Kovacs

This is my revision article. Please give me the information if the revised part is still not clear. thank you

Regards Veriani Aprilia [Kutipan teks disembunyikan]

rev_Aprilia_Clarias gariepinus_wip.doc 180K



The residue of formalin in catfish (*Clarias* gariepinus) after a short period processing and storage for short periods

Isny Maulidevi, Nurul Andriyani, Ryan Salfarino, Veriani Aprilia

Department of Nutrition, Faculty of Health Sciences, Alma Ata University, Yogyakarta, Indonesia. Corresponding author: V. Aprilia, verianiaprilia@almaata.ac.id

Abstract. Fresh fish is a highly perishable product due to its high protein content. A preservative may be needed to inhibit it. Nowadays, many fishes were reported to be exposed to formalin. This study aimed to determine the effect of short period processing (washing, frying, boiling in short-period) and storage (in-at the room and at freezing temperatures) on the residue of formalin in caffish, as a food model. Catfish, *Clarias qariepinus*, were divided into 8 groups, e.g. before treatment, washing in airflow for 1 minute, frying in 200°C for 8 minutes, boiling in 100°C for 7 minutes, storage at room temperature (29±1°C) for 24 and 48 hours, storage at freezing temperature (-3°C) for 24 and 48 hours. Before treatment, the catfish was soaked in 1,480 ppm of formalin solutions. The residue of formalin was measured at the end of each treatment by using a spectrophotometer. Data were then analyzed with analysis of variance with SPSS software. The results showed that there were significant effects of all treatments was lower than that of before treatments. The frying processing and storage at freezing temperatures were the most effective treatments. The frying processing and storage at freezing temperatures were the most effective treatments. The formalin residue of all treatments was lower than that of before treatments. The formalin residue. **Key Words**: formalin, fish, processing, storage, formaldehyde.

Introduction. Fresh fish is a highly perishable product due to its high protein content for microbial activity (Wijayanti & Lukitasari 2016). A preservative maybe needs to keep the freshness, inhibit the decay₇ and prolong[⊭] the shelf life (Antoni 2010). Nowadays, there are many kinds of food-grade preservatives available in the market. Unfortunately, formalin is still frequently found in food products, including fresh fish, being used among fishermen The-mainly because reason for the use among fishermen is cheaper, easy to find, and it is effectivity effective to in preserve-preserving the product, cheaper and accessible (Male et al 2017).

Formalin is a forty percent solution of formaldehyde in water. This compound is detected in the animal body as an intermediary metabolite. It also uses for several purposes, such as in medicine for specimen preservation and the treatment of hemorrhage (Pandey et al., 2000). The preservatives activity of formalin because of due to its ability to react with protein generating methylene compounds, which is are more difficult to be decayed by microbes (Ichya'uddin 2016). In low concentrations, formalin is not toxic for humans, but in higher concentrations, it had negative effects such as acute toxicity and death (Pandey et al 2000). It also decreased decreases the levels of body antioxidants that, resulting in liver damage (Yulisa et al 2014). Other studies also reported the cases of eczema, eye irritation, respiratory tract irritation, cancer, increased the growth rate of malignant neoplastic diseases; and denaturation of DNA (Suwanaruang 2018; Wilianto & Yudianto 2013). Therefore, its uses are forbidden.

Formalin solution <u>was-is</u> not stable during storage and <u>exposed exposure</u> to high temperatures (Kaneko et al., 1977; Laksmiani et al., 2015). <u>Those are caused by</u> desolvation or evaporation of formalin during washing, boiling, or frying (Fellows 2015; Harmita 2016; Muntaha et al 2015; Sundari et al 2015) and enzymatic process in storage (Rachmawati et al 2007). Formalin that was <u>exposed to thein contact with</u> fish decreased up to 43-53% after 5 times rinsing in 250 mL of distilled water and boiling for 30 minutes

AACL Bioflux, 2020, Volume 13, Issue X. http://www.bioflux.com.ro/aacl

1090

Commented [WU1]: Not listed in the references.

I have added it

Commented [WU2]: Not listed in the references.

I have added it

(Yusuf et al 2015). Other studies were also reported that soaking in warm water and frying decreased the concentration of formalin up to 63_{72} 27% and 83.03%, respectively (Levita et al 2010). In daily practices, fish is usually washed in flowing water before processing, continued_then_to_fryingfryed/boiling_boiled_in_afor a short_er_time. However, there was no study explained_explaining_the influences of these processing and storage methods on the concentration of formalin.

The purpose of this study was to <u>know_identify</u> the effect of processing <u>in_for_a</u> short time and storage (<u>at_room_and_freezing_temperatures_and_freezing</u>) on the residue of formalin. This study used catfish as a food model.

Material and Method

Material. The study used fresh catfish, purchased from the traditional market in YogyakartaYogyakarta, Indonesia. <u>TheAn</u> other used ingredients used—were formalin solution and Schiff's reagent. Formalin solution was made from 4 mL of formalin 37% in 1,000 mL of distilled water (Rachmawati et al 2007).

Method. The study was divided into two sections, i.e. 1) study analyzing the effect of processing and 2) analyzing the effect of storage treatment. Fresh catfish were slaughtered, eviscerated, cleaned, and soaked in 1,480 ppm formalin solution (Rachmawati et al 2007; Sugiarti et al 2014). For the first study, catfish were divided into four groups, i.e.: controls (before treatment), washing, frying, and boiling treatments. Washing was done by watering the samples with running water for 1 minute, while frying was done by frying the samples in oil at 200°C for 8 minutes. Boiling treatment was done by putting the samples into boiled water (100°C) for 7 minutes (Sugiarti et al 2014). For the second study, storage treatments, the catfish was also divided into four groups, they: were room temperature storage (29°C) for 24 hours and 48 hours and freezing temperatures storage (-3°C) for 24 hours and 48 hours (Handayani et al 2010). At the end of treatments, the concentration of formalin in samples was analyzed using a spectrophotometer. As the preparation step, a 5 g sample was dissolved in 25 mL of distilled water. One mL of supernatant was mixed with 2 mL of Schiff's reagent, then heated in a water bath at 60°C for 5 minutes. It was incubated for 10 minutes before measuring the absorbance at 520 nm (Shita 2016; Mudaffar 2018).

Statistical analysis. Data were presented in mean±SD and analyzed with: (1) one-way analysis of variance (ANOVA) to know_determine_the differences among all processing treatments and the effect of storage periods on the concentration of formalin; (2) paired sample T-test to determineknow the differences between the same samples before and after processing treatments; (3) independent sample t-test to know the effect of temperature at the same storage period, and (4) two-way ANOVA to determineknow the concentration of formalin. All data were analyzed with SPSS software (version 16.0 for Windows; SPSS Inc., Chicago, IL, USA) at the 5% significance level.

Results

Effect of processing on the formalin residue of catfish. Table 1 shows that all treatments had significant effects on the residue of formalin (p<0.05). It can be seen from the p-value between the same samples before and after treatments. Among all treatments, washing shows the <u>lowest-highest</u> residue of formalin (p<0.05). Both boiling and frying had a<u>n almost</u> similar residue of formalin (p>0.05). The <u>higher_lower</u> residue of formalin in boiling and frying may be due to heating exposure.

Commented [S3]: Yogyakarta is different from Jakarta

Commented [WU4]: The text was aligned to table 1.
Commented [S5R4]: Thank you for the corrections

AACL Bioflux, 2020, Volume 13, Issue X. http://www.bioflux.com.ro/aacl

1091

Effect of processing on the formalin residue of catfish

Processing	Residue of fo	Δ residue of formalin	
treatments			before and after
liealments	Belore treatment	Alter treatment	treatment (%)
Washing	1,420.17±66.94 ^{c1}	310.42±77.61 ^{b2}	78.14
Frying	1,420.17±66.94 ^{c1}	14.18±6.69 ^{a2}	99.01
Boiling	1,420.17±66.94 ^{c1}	27.50±9.11 ^{a2}	98.10
Mean+SD in the	same column with different :	alphabetical superscripts (a. b	or c) are significantly different

(p+0.05) by using one-way ANOVA statistical analysis. Mean SpetSD in the same row with different number superscripts (1 or 2) are significantly different (p<0.05) by using paired sample T-test statistical analysis.

Effect of storage temperatures and periods on the formalin residue of catfish. Table 2 shows the effect of storage temperatures, <u>of periods</u>, and <u>of their</u> combination—of both on the concentration of formalin. Based on this data, the storage period influenced the residue of formalin <u>decrease</u> (p<0.05)—<u>during the first 24 hours</u>. It could be seen from the decrease of formalin residue during 24 hours of storage. However, but after a storage period of 48 hours the decrease <u>did</u>—was_not significantly different <u>when the</u>,—storage continued to 48 hours (p>0.05).

On the other hand, there were no differences in the residue of formalin <u>stored_at</u> room and freezing temperature. <u>It was indicated, suggesting</u> that the temperature of storage did not affect the formalin residue (p>0.05). Although there were differences in <u>the</u> statistical results between the <u>effect of</u> temperature <u>effects</u> and <u>the storage</u> periods <u>of storage_effects</u>, the combined effect of both variables showed no impact on the concentration of formalin (p>0.05).

Table 2

Table 1

Effect of storage temperature and period on the formalin residue of catfish

Temperature	Per	riods (t) (hours)			p-value	9
(T)	0	24	48	Т	t	T*t
Room (29°C)	1,420.17±66.94 ^{a1}	52.18±28.78 ^{a2}	40.64±3.93 ^{a2}	0.20	0.00	0.76
Freezing (-3°C)	1,420.17±66.94 ^{a1}	39.04±7.07 ^{a2}	13.65±4.23 ^{a2}	0.20	0.00	0.76

Mean \pm SD in the same column with different alphabetical superscripts (a, b, or c) are significantly different (p<0.05) and so are in the same row with different number superscript (1, 2, or 3). The p-value for T, t, and T*t was obtained from independent T-test, one-way ANOVA, and two-way ANOVA, respectively.

Discussion

The residue of formalin before treatment. The residue of formalin before and after treatment (either fish processing or storage) was significantly different (Table 1 and Table 2). In this study, the catfish specimens were submerged for 60 minutes in 1,480 ppm of formalin solution of 1,480 ppm.was added to the catfish and submerged for 60 minutes. The 60 minutes submersion period was chosen because the aldehyde compound may be readily bonded to the amino acids of fish in the <u>a</u> small amount (Rahmadhani et al 2017; Baroroh & Kresnadipayana 2016). Formalin would attack lysine for the first time, followed by histidine and tyrosine, which ledleading to the formation of <u>a</u> methylene compound (Ichya'uddin 2016). The protein-containing methylene compounds could not be digested by living things and microbes, especially pathogens, so that the shelf life would be longer (Purawisastra & Sahara 2011). The beneficial bacteria in the fish may also be killed by dehydration mechanism₄ as another effect of the existence of formalin in the body (Mudzkirah 2016).

The effect of processing on the formalin residue of catfish. The residue of formalin was reduced to-by 78.14% during washing treatment. The reduction was significant

AACL Bioflux, 2020, Volume 13, Issue X. http://www.bioflux.com.ro/aacl

1092

Commented [S7]: I have revised the year. Thank you

Commented [S6]: I have deleted 2 columns of p values and replaced it with the superscript to show the statistical result

(p<0.05). The formalin was easily washed out because of its high solubility in the water, which is 24 x 105 mg L⁻¹ at 20°C (BPOM 2008; Sugiarti et al 2014; Yusuf et al 2015). Another study conducted by Yusuf et al (2015) also-showed the same trend, but the reduction was lower (with only 43%). The differences in the level of reduction can be affected by the length-duration of formalin submersion. The longer the submersion time, the smaller the decreasing decrease, as there was more formalin was bonded with the fish protein in the fish.

In this study, frying also significantly reduced the residue of formalin (p<0.05). The reduction level was 99% that indicated most <u>of the</u> formalin <u>was</u>-disappeared during the frying process. Formalin was included in a volatile compound, especially at the above of-its boiling point (Joshi et al 2015). This study <u>was also the same withconfirmed the</u> <u>results of</u> Sugiarti et al (2014), <u>showing a that found the</u> reduction level of formalin up to 99.28% after 1 hour of submersion followed by the frying process (no information about the period of frying process) in squid. Frying could denature the protein and also hydrolyze its bonding with formalin (Levita et al 2010). The high temperature in frying increased the kinetic energy and led to faster movement of the molecules that could damage the molecular bonds (Sugiarti et al 2014).

Boiling was another heat-Heat processing by boiling that was done in this study. The results showed that this process also reduced the residue of formalin with up to 98.1%, more than. The residue of formalin after treatment was different from the sample after the washing process (p<0.05), but at the same degree with asthat after the frying process (p>0.05). In the boiling process, tThe high reduction of formalin in the boiling process was higher than that was in only washing or soaking treatment because there was morewas due to the accelerated formalin dissolved dissolution rate in at higher temperatures water (Annisak 2019). This effect could be enhanced by amount of losing would be higher when it combined boiling the sample with thein an opened pan during the boiling (Kamal et al 2017).

The effect of storage on the formalin residue of catfish. This research experiment found demonstrated that only a-the period of storage that significantly influenced the residue of formalin (p<0.05), while the temperature of storage and the combination between temperature and period of storage did not (Table 2). The previous different studies showed that either a storage at the room temperature or at the freezing temperature of storage had a similar effects on the decreasing of formalin residue decrease (Jawahar et al 2017; Murtini et al 2014), possibly . It may be due to the same degradation level of protein degradationduring both in the room and freezing temperature, resulting in the loss of by breaking the bonds between protein and formalin and releasing free compounds. In-At the room temperature, the protein degradation was more essentially caused by the microbial activity, while in at the freezing temperature it was more caused mostly by physical destruction, leading to the conformational and functional changes (Murtini et al 2014; Yeasmin et al 2010). Formalin in the free form is more reactive and can also be more easily degraded was easier to break down into another compound or reacts with other compounds in the free form (Riyanto et al 2006; Rachmawati et al 2007).

Conclusions. This research concluded that short time processing and storage treatments decreased the concentration of formalin. Heat involvement, especially during boiling and frying process made determined a more effective the decrease of concentration became more effective (the decrease level was >90%). In the storage study, the residue of formalin was influenced significantly reduced by the period of storage, mainly during the first 24 hours, then the formalin concentration reduction was insignificant, up to 48 hours of storage.

Acknowledgements. The authors are grateful to the Indonesia Ministry of Research, Technology, and Higher Education for providing the financial support in pursuing this research project (Program Kreativitas Mahasiswa Kemeristekdikti 2019 Program).

Commented [WU8]: Please add some details concerning the above mentioned results.

Thank you for the suggestion. Please check the revision. Is this enough or not.

AACL Bioflux, 2020, Volume 13, Issue X. http://www.bioflux.com.ro/aacl

1093

Reference

- Annisak, S. K., Indriyanti, N. Y., Mulyani, B., 2019 Constructive controversy dan inkuiri terbimbing sesuai representasi tetrahedral pembelajaran kimia ditinjau dari kemampuan berpikir kritis. Jurnal Inovaso Pendidikan IPA 5(1):10–22.
- Antoni, S., 2010 Analisa kandungan formalin pada ikan asin dengan metoda spektrofotometri di Kecamatan Tampan Pekanbaru. Universitas Islam Negeri Sultan Syarif Kasim Riau.
- Baroroh, R. D., Kresnadipayana, D., 2016 Pengaruh variasi konsentrasi sari buah belimbing wuluh (Averrhoa bilimbi L.) pada perendaman tahu terhadap penurunan kadar formalin. Universitas Setia Budi.
- BPOM. 2008 Formalin (Larutan Formaldehid). Badan POM RI.
- Fellows, P. J., 2015 Teknologi Pengolahan Pangan. EGC.
- Handayani, A., Alimin, Rustiah, W. O., 2010 Pengaruh penyimpanan pada suhu rendah (freezer -3°c) terhadap kandungan air dan kandungan lemak pada ikan lemuru (Sardinella longiceps). Al-Kimia 3:64–75.
- Harmita, 2016 Penetapan kadar bahan baku obat dan sediaan farmasi. EGC.
- Ichya'uddin, M., 2014 Analisis kadar formalin dan uji organoleptik ikan asin di beberapa pasar tradisional di Kabupaten Tuban. Universitas Islam Negeri Maulana Malik Ibrahim.
- Jawahar, L. S., John, C., Shafeekh, M., Anupama, T. K., Sankar, T. V., 2017 Retention of residual formaldehyde in treated indian mackerel (Rastrelliger kanagurta) under iced storage and related food safety concern. Indian J. Fish 64(4):87–93. https://doi.org/10.21077/ijf.2017.64.4.61228-12.
- Joshi, Bhatta, R., Paudel, P. N., Kafle, B. K., 2015 Formaldehyde content of selected fish from the wet markets of Kathmandu valley. International Food Research Journal 22(4):1434-1437.
- Kamal, A. S., Uddin, M. M., Rahman, R., Samira, S., Rahman, M., Nandy, S., Khan, M. S., 2017 Time and temperature effect on the residual concentration of formaldehyde in formalin treated samples of Labeo rohita. International Conference on Food Security and Nutrition.
- Kaneko, M., Wada, Y., Fukui, S., Kanno, S., 1977 NII-Electronic Library Service. The Journal of Hygienic Chemistry 23(6):393–396.
- Laksmiani, N., Widjaja, I., Sonia, 2015 Stabilitas formalin terhadap pengaruh suhu dan lama pemanasan. Jurnal Farmasi Udayana 4(2):76–81. https://ojs.unud.ac.id/index.php/jfu/article/view/17187/11312
- Levita, J., Musfiroh, I., Indriyati, W., Mustarichie, R., 2010. The effect of soaking, washing, and frying on the concentration of formaldehyde in Sange Belah Salty Fish. Jurnal Ilmu-Ilmu Hayati Dan Fisik 12(1):31–34.
- Male, Y. T., Letsoin, L. I., Siahaya, N. A., 2017 Analisis kandungan formalin pada mie basah pada beberapa lokasi di kota ambon. Jurnal Kementerian Perindustrian 13(2):5-10. https://doi.org/10.29360/mb.v13i2.3530.
- Mudaffar, R. A., 2018 Uji kualitatif dan kuantitatif formalin pada buah apel, anggur, dan lengkeng yang dijual di Kota Makassar. Jurnal Perbal 6(3).
- Mudzkirah, I., 2016 Identifikasi penggunaan zat pengawet boraks dan formalin pada makanan jajanan Di Kantin UIN Alauddin Makassar. UIN Alauddin Makassar.
- Muntaha, A., Haitami, Hayati, N., 2015 Perbandingan penurunan kadar formalin pada tahu yang direbus dan direndam air panas. Medical Laboratory Technology Journal 1(2):84–90.
- Murtini, J. T., Riyanto, R., Priyanto, N., Hermana, I, 2014 Pembentukan Formaldehid alami pada Beberapa Jenis Ikan Laut Selama Penyimpanan dalamn Es Curai. JPB Perikanan 9(2):143–151.
- Pandey, C. K., Agarwal, A., Baronia, A., Singh, N., 2000 Toxicity of ingested formalin and its management. Human and Experiment Toxicology, 19(November 1999):360– 366.
- Purawisastra, S., Sahara, E., 2011. Penyerapan formalin oleh beberapa jenis bahan makanan serta penghilangannya melalui perendaman dalam air panas. PGM

AACL Bioflux, 2020, Volume 13, Issue X. http://www.bioflux.com.ro/aacl 34(1):63-74.

- Rachmawati, N., Riyanto, R., Ariyani, F., 2007 Pembentukan formaldehod pada ikan kerapu macan (Ephinephelus fuscoguttatus) selama penyimpanan suhu dingin. Jurnal Pascapanen Dan Bioteknologi Kelautan Dan Perikanan 2(2):137-145.
- Rahmadhani, F., Safrida, Djufri, 2017 Pengaruh perendaman berbagai larutan terhadap penurunan kadar formalin pada ikan asin kembung (Scomber canagorta) di Pasar Lambaro Aceh. Jurnal Ilmiah Mahasiswa Keguruan Dan Ilmu Pendidikan Unsyiah.
- Riyanto, R., Kusmarwati, A., Dwiyitno., 2006 Pembentukan Formaldehid pada ikan kerapu (Epinephelus fuscoguttatus) selama penyimpanan pada suhu kamar. Jurnal Pascapanen dan Bioteknologi Kelautan dan Perikanan 1(2):111-116.
- Shita, A. E., 2016 Selektivitas metode analisis formalin secara spektrofotometri dengan pereaksi Schiff's. Universitas Negeri Yogyakarta.
- Sugiarti, M., Anggo, A. D., Riyadi, P. H., 2014 Efek perendaman pada suhu undercooking dan metode cooking terhadap pengurangan kadar formalin pada cumi-cumi (Loligo sp.). Jurnal Pengolahan dan Bioteknologi Hasil Perikanan 3(2):90-98.
- Sundari, D., Almasyhuri, Lamid, A., 2015 Pengaruh proses pemasakan terhadap komposisi zat gizi bahan pangan sumber protein. Media Litbangkes 25(4):235-242.
- Suwanaruang, T., 2018 Formalin Contaminated in Seafood and Frozen Meat at Somdet Market, Kalasin Province. Journal of Environmental Protection 9:1286-1293. https://doi.org/10.4236/jep.2018.912080
- Wijayanti, N. S., Lukitasari, M., 2016 Analisis kandungan formalin dan uji organoleptik ikan asin yang beredar di pasar besar madiun. Jurnal Florea 3(1): 59-64.
- Wilianto, W., Yudianto, A., 2013 Pengaruh Paparan Formalin Dalam Berbagai Kadar Terhadap Dna Muskulus Psoas Mayor Dengan Pemeriksaan Metode Pcr Lokus CSF1PO , D5S818 , D13S317 , D21S11. Jurnal Kedokteran Forensik Indonesia 15(1):11-16.
- Yeasmin, T., Reza, M. S., Shikha, F. H., Khan, M. N. A., Kamal, M., 2010 Quality changes in formalin treated rohu fish (Labeo rohita, Hamilton) during ice storage condition. Asian Journal of Agricultural Science 2(4): 158-163.
- Yulisa, N., Asni, E., Azrin, M., 2014 Uji formalin pada ikan asin gurami di Pasar Tradisional Pekabaru 1(2).
- Yusuf, Y., Zuki, Z., MP, Amanda, R. R., 2015 Pengaruh beberapa perlakuan terhadap pengurangan kadar formalin pada ikan yang ditentukan secara spektrofotometri. J. Ris. Kim 8(2):182-188.

Received: 15 October 2020. Accepted: 2020. Published online: 2020.

Authors: Isny Maulidevi, Department of Nutrition, Faculty of Health Sciences, Alma Ata University, Jl. Brawijaya 99, Isny Maulidevi, Department of Nutrition, Faculty of Health Sciences, Alma Ata University, Jl. Brawijaya 99, Yogyakarta 55183 Indonesia, email: <u>isnymaulidevi540@gmail.com</u> Nurul Andriyani, Department of Nutrition, Faculty of Health Sciences, Alma Ata University, Jl. Brawijaya 99, Yogyakarta 55183 Indonesia, email: <u>nurulandriyani96@gmail.com</u> Ryan Salfarino, Department of Nutrition, Faculty of Health Sciences, Alma Ata University, Jl. Brawijaya 99,

Yogyakarta 55183 Indonesia, email: <u>ryan.salfarino@gmail.com</u> Veriani Aprilia, Department of Nutrition, Faculty of Health Sciences, Alma Ata University, Jl. Brawijaya 99, Yogyakarta 55183 Indonesia, email: <u>verianiaprilia@almaata.ac.id</u> This is an open-access article distributed under the terms of the Creative Commons Attribution License, which

permits unrestricted use, distribution and reproduction in any medium, provided the original author and source are credited.

How to cite this article:

Maulidevi I., Andriyani N., Salfarino R., Aprilia V., 2020 The residue of formalin in catfish (Clarias gariepinus) essing and storage for short periods. AACL Bioflux 13(x): after proc

AACL Bioflux, 2020, Volume 13, Issue X. http://www.bioflux.com.ro/aacl

1095

Commented [S9]: I have added this missing information. Thank you



verianiaprilia verianiaprilia <verianiaprilia@almaata.ac.id>

Pemberitahuan revisi 2: 7 Desember 2020

Revision

2 pesan

Eni Kovacs <ek.bioflux@gmail.com> Kepada: verianiaprilia verianiaprilia <verianiaprilia@almaata.ac.id>

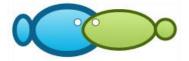
Dear Veriani Aprilia,

Please find the manuscript attached and make the necessary adjustments.

Thank you.

Kind regards, Editor AACL Bioflux Eniko Kovacs

Aprilia_Clarias gariepinus_final.doc 167K 7 Desember 2020 pukul 04.02



The residue of formalin in catfish (*Clarias* gariepinus) after processing and storage for short periods

İsny Maulidevi, Nurul Andriyani, Ryan Salfarino, Veriani Aprilia

Department of Nutrition, Faculty of Health Sciences, Alma Ata University, Yogyakarta, Indonesia. Corresponding author: V. Aprilia, verianiaprilia@almaata.ac.id

Abstract. Fresh fish is a highly perishable product due to its high protein content. A preservative may be needed to inhibit it. Nowadays, many fishes were reported to be exposed to formalin. This study aimed to determine the effect of short period processing (washing, frying, boiling in short-period) and storage (at the room and at freezing temperatures) on the residue of formalin in catfish, as a food model. Catfish, *Clarias gariepinus*, were divided into 8 groups, e.g. before treatment, washing in airflow for 1 minute, frying in 200°C for 8 minutes, boiling in 100°C for 7 minutes, storage at room temperature ($29\pm1°C$) for 24 and 48 hours, storage at freezing temperature (-3°C) for 24 and 48 hours. Before treatment, the catfish was soaked in 1,480 ppm of formalin solutions. The residue of formalin was measured at the end of each treatment by using a spectrophotometer. Data were then analyzed with analysis of variance with SPS software. The results showed that there were significant effects of all treatments was lower than before treatments. The frying processing and storage at freezing temperatures of catfish. The formalin residue of all treatments was lower than before treatments in reducing the formalin residue. **Key Words**: fresh fish, preservative, food model, treatments, formalin residue.

Introduction. Fresh fish is a highly perishable product due to its high protein content for microbial activity (Wijayanti & Lukitasari 2016). A preservative maybe needs to keep the freshness, inhibit the decay and prolong the shelf life (Antoni 2010). Nowadays, there are many kinds of food-grade preservatives available in the market. Unfortunately, formalin is still frequently found in food products, including fresh fish, being used among fishermen mainly because it is effective in preserving the product, cheaper and accessible (Male et al 2017).

Formalin is a forty percent solution of formaldehyde in water. This compound is detected in the animal body as an intermediary metabolite. It also uses for several purposes, such as in medicine for specimen preservation and the treatment of hemorrhage (Pandey et al 2000). The preservatives activity of formalin is due to its ability to react with protein generating methylene compounds, which are more difficult to be decayed by microbes (Ichya'uddin 2016). In low concentrations, formalin is not toxic for humans, but in higher concentrations, it had negative effects such as acute toxicity and death (Pandey et al 2000). It also decreases the levels of body antioxidants, resulting in liver damage (Yulisa et al 2014). Other studies also reported the cases of eczema, eye irritation, respiratory tract irritation, cancer, increased growth rate of malignant neoplastic diseases and denaturation of DNA (Suwanaruang 2018; Wilianto & Yudianto 2013). Therefore, its uses are forbidden.

Formalin solution is not stable during storage and exposure to high temperatures (Kaneko et al 1977; Laksmiani et al 2015). Those are caused by desolvation or evaporation of formalin during washing, boiling, or frying (Fellows 2015; Harmita 2016; Muntaha et al 2015; Sundari et al 2015) and enzymatic process in storage (Rachmawati et al 2007). Formalin that was in contact with fish decreased up to 43-53% after 5 times rinsing in 250 mL of distilled water and boiling for 30 minutes (Yusuf et al 2015). Other studies were also reported that soaking in warm water and frying decreased the

AACL Bioflux, 2020, Volume 13, Issue X. http://www.bioflux.com.ro/aacl concentration of formalin up to 63.27% and 83.03%, respectively (Levita et al 2010). In daily practices, fish is usually washed in flowing water before processing, then fryed/boiled for a short time. However, there was no study explaining the influences of these processing and storage methods on the concentration of formalin.

The purpose of this study was to identify the effect of processing for a short time and storage (at room and freezing temperatures) on the residue of formalin. This study used catfish as a food model.

Material and Method

Material. The study used fresh catfish, purchased from the traditional market in Yogyakarta, Indonesia. The other used ingredients were formalin solution and Schiff's reagent. Formalin solution was made from 4 mL of formalin 37% in 1,000 mL of distilled water (Rachmawati et al 2007).

Method. The study was divided into two sections, i.e. 1) analyzing the effect of processing and 2) analyzing the effect of storage treatment. Fresh catfish were slaughtered, eviscerated, cleaned, and soaked in 1,480 ppm formalin solution (Rachmawati et al 2007; Sugiarti et al 2014). For the first study, catfish were divided into four groups, i.e.: controls (before treatment), washing, frying, and boiling treatments. Washing was done by watering the samples with running water for 1 minute, while frying was done by frying the samples in oil at 200°C for 8 minutes. Boiling treatment was done by putting the samples into boiled water (100°C) for 7 minutes (Sugiarti et al 2014). For the second study, storage treatments, the catfish was also divided into four groups: room temperature storage (29°C) for 24 hours and 48 hours and freezing temperatures storage (-3°C) for 24 hours and 48 hours (Handayani et al 2010). At the end of treatments, the concentration of formalin in samples was analyzed using a spectrophotometer. As the preparation step, a 5 g sample was dissolved in 25 mL of distilled water. One mL of supernatant was mixed with 2 mL of Schiff's reagent, then heated in a water bath at 60°C for 5 minutes. It was incubated for 10 minutes before measuring the absorbance at 520 nm (Shita 2016; Mudaffar 2018).

Statistical analysis. Data were presented in mean±SD and analyzed with: (1) one-way analysis of variance (ANOVA) to determine the differences among all processing treatments and the effect of storage periods on the concentration of formalin; (2) paired sample T-test to determine the differences between the same samples before and after processing treatments; (3) independent sample t-test to know the effect of temperature at the same storage period, and (4) two-way ANOVA to determine the combination effect of storage temperature and period on the concentration of formalin. All data were analyzed with SPSS software (version 16.0 for Windows; SPSS Inc., Chicago, IL, USA) at the 5% significance level.

Results

Effect of processing on the formalin residue of catfish. Table 1 shows that all treatments had significant effects on the residue of formalin (p<0.05). It can be seen from the p-value between the same samples before and after treatments. Among all treatments, washing shows the highest residue of formalin (p<0.05). Both boiling and frying had an almost similar residue of formalin (p>0.05). The lower residue of formalin in boiling and frying may be due to heating exposure.

1091

Effect of processing on the formalin residue of catfish

Processing	Residue of formalin (ppm)		Δ residue of formalin		
treatments	Before treatment	After treatment	<i>before and after treatment</i> (%)		
Washing	1,420.17±66.94 ^{c1}	310.42±77.61 ^{b2}	78.14		
Frying	1,420.17±66.94 ^{c1}	14.18±6.69 ^{a2}	99.01		
Boiling	1,420.17±66.94 ^{c1}	27.50±9.11 ^{a2}	98.10		
Mean+SD in the same column with different alphabetical superscripts (a, b, or c) are significantly different					

(p<0.05) by using one-way ANOVA statistical analysis. Mean \pm SD in the same row with different number superscripts (1 or 2) are significantly different (p<0.05) by using one-way ANOVA statistical analysis.

Effect of storage temperatures and periods on the formalin residue of catfish. Table 2 shows the effect of storage temperatures, of periods and of their combination on the concentration of formalin. Based on this data, the storage period influenced the residue of formalin decrease (p<0.05) during the first 24 hours, but after a storage period of 48 hours the decrease was not significantly different (p>0.05).

On the other hand, there were no differences in the residue of formalin stored at room and freezing temperature, suggesting that the temperature of storage did not affect the formalin residue (p>0.05). Although there were differences in the statistical results between the temperature effects and the storage periods effects, the combined effect of both variables showed no impact on the concentration of formalin (p>0.05).

Table 2

Effect of storage temperature and period on the formalin residue of catfish

Temperature	Periods (t) (hours)				p-value	9
(T)	0	24	48	Т	t	T*t
Room (29°C)	1,420.17±66.94 ^{a1} 1,420.17±66.94 ^{a1}	52.18±28.78 ^{a2}	40.64±3.93 ^{a2}	0.20	0.00	0.76
Freezing (-3°C)	1,420.17±66.94 ^{a1}	39.04±7.07 ^{a2}	13.65±4.23 ^{a2}	0.20	0.00	0.76

Mean±SD in the same column with different alphabetical superscripts (a, b, or c) are significantly different (p<0.05) and so are in the same row with different number superscript (1, 2, or 3). The p-value for T, t, and T*t was obtained from independent T-test, one-way ANOVA, and two-way ANOVA, respectively.

Discussion

The residue of formalin before treatment. The residue of formalin before and after treatment (either fish processing or storage) was significantly different (Table 1 and Table 2). In this study, the catfish specimens were submerged for 60 minutes in formalin solution of 1,480 ppm. The 60 minutes submersion period was chosen because the aldehyde compound may be readily bonded to the amino acids of fish in a small amount (Rahmadhani et al 2017; Baroroh & Kresnadipayana 2016). Formalin would attack lysine for the first time, followed by histidine and tyrosine, leading to the formation of a methylene compound (Ichya'uddin 2016). The protein-containing methylene compounds could not be digested by living things and microbes, especially pathogens, so that the shelf life would be longer (Purawisastra & Sahara 2011). The beneficial bacteria in the fish may also be killed by dehydration mechanism, as another effect of the existence of formalin in the body (Mudzkirah 2016).

The effect of processing on the formalin residue of catfish. The residue of formalin was reduced by 78.14% during washing treatment. The reduction was significant (p<0.05). The formalin was easily washed out because of its high solubility in the water, which is 24 x 105 mg L⁻¹ at 20°C (BPOM 2008; Sugiarti et al 2014). A study conducted by Yusuf et al (2015) showed the same trend, but the reduction was lower (with only 43%). The differences in the level of reduction can be affected by the duration of

AACL Bioflux, 2020, Volume 13, Issue X. http://www.bioflux.com.ro/aacl

1092

Table 1

formalin submersion. The longer the submersion time, the smaller the decrease, as more formalin was bonded with the fish protein.

In this study, frying also significantly reduced the residue of formalin (p<0.05). The reduction level was 99% that indicated most of the formalin disappeared during the frying process. Formalin was included in a volatile compound, especially above its boiling point (Joshi et al 2015). This study confirmed the results of Sugiarti et al (2014), showing a reduction level of formalin up to 99.28% after 1 hour of submersion followed by the frying process (no information about the period of frying process) in squid. Frying could denature the protein and also hydrolyze its bonding with formalin (Levita et al 2010). The high temperature in frying increased the kinetic energy and led to faster movement of the molecules that could damage the molecular bonds (Sugiarti et al 2014).

Heat processing by boiling reduced the residue of formalin with up to 98.1%, more than the washing process (p<0.05), but at the same degree as the frying process (p>0.05).The high reduction of formalin in the boiling process was due to the accelerated formalin dissolution rate at higher temperatures (Annisak 2019). This effect could be enhanced by boiling the sample in an open pan (Kamal et al 2017).

The effect of storage on the formalin residue of catfish. This experiment demonstrated that only the period of storage significantly influenced the residue of formalin (p<0.05), while the temperature of storage and the combination between temperature and period of storage did not (Table 2). The previous different studies showed that a storage at the room temperature or at the freezing temperature had similar effects on the formalin residue decrease (Jawahar et al 2017; Murtini et al 2014), possibly due to the same level of protein degradation by breaking the bonds between protein and formalin and releasing free compounds. At the room temperature, the protein degradation was essentially caused by the microbial activity, while at the freezing temperature it was caused mostly by physical destruction, leading to conformational and functional changes (Murtini et al 2014; Yeasmin et al 2010). Formalin in the free form is more reactive and can also be more easily degraded (Riyanto et al 2006; Rachmawati et al 2007).

Conclusions. This research concluded that short time processing and storage treatments decreased the concentration of formalin. Heat involvement determined a more effective decrease of concentration (the decrease level was >90%). In the storage study, the residue of formalin was significantly reduced by the period of storage, mainly during the first 24 hours, then the formalin concentration reduction was insignificant, up to 48 hours of storage.

Acknowledgements. The authors are grateful to the Indonesia Ministry of Research, Technology, and Higher Education for providing the financial support in pursuing this research project (Program Kreativitas Mahasiswa Kemeristekdikti 2019 Program).

Reference

- Annisak S. K., Indriyanti N. Y., Mulyani B., 2019 [Constructive controversy and incubation presented according to the representation of tetrahedral chemical learning according to critical thinking ability]. Jurnal Inovasi Pendidikan IPA 5(1):10–22. [In Indonesian].
- Antoni S., 2010 [Analysis of formaldehyde content on salty fish by spectrophotometri method in Tampan District Pekanbaru]. ... Type of study, Universitas Islam Negeri Sultan Syarif Kasim Riau, No of pages. [In Indonesian].

Baroroh R. D., Kresnadipayana D., 2016 [The effect of variety consentration of starfruit juice (*Averrhoa bilimbi* L.) on soaked tofu for formaldehyde reduction]. Publisher, Volume, Issue, Pages. [In Indonesian].

BPOM 2008 [Formaldehyde]. Jakarta: Badan POM RI. No of pages [In Indonesian]. What does BPOM stands for?

1093

AACL Bioflux, 2020, Volume 13, Issue X. http://www.bioflux.com.ro/aacl

information required.

Commented [WU1]: Please replace the red text with the

Fellows P. J., 2015 [Food processing technology]. Publisher, no of pages EGC Jakarta. ISBN: 979-044-612-0. [In Indonesian].

- Handayani A., Alimin, Rustiah W. O., 2010 [The effect of low temperature storage (freezer -3oc) on water and fat content in lemuru fish (*Sardinella longiceps*)]. Al-Kimia (3):64–75. [In Indonesian].
- Harmita, 2016 [Determination substance and availability of pharmaceuticals content]. Publisher, Jakarta: EGC. No of pages ISBN: 979-044-773-8. [In Indonesian].
- Ichya'uddin M., 2014 [Analysis of Formaldehyde content and organoleptic test on salty fish in several traditional markets in Tuban]. Publication type, Universitas Islam Negeri Maulana Malik Ibrahim, no of pages. [In Indonesian].
- Jawahar L. S., all the authors must be listed, 2017 Retention of Residual formaldehyde in treated Indian mackerel (*Rastrelliger kanagurta*) under iced storage and related food safety concern, Indian J. Fish. no abbreviations 64(4):87–93.
- Joshiall the authors must be listed, 2015 Formaldehyde content of selected fish from the wet markets of Kathmandu valley. International Food Research Journal 22(4):...pages. ISSN: 1434-1437.
- Kamal A. S., all the authors must be listed, 2017 Time and Temperature effect on the residual concentration of formaldehyde in formalin treated samples of *Labeo rohita*. International Conference on Food Security and Nutrition, pages.
- Kaneko M., Wada Y., Fukui S., Kanno S., 1977 NII-Electronic Library Service. The Journal of Hygienic Chemistry 23(6):393–396.
- Laksmiani N., Widjaja I., Sonia, 2015 [Stabilitas formalin terhadap pengaruh suhu dan lama pemanasan]. PLease translate the title Jurnal Farmasi Udayana 4(2):76–81. https://ojs.unud.ac.id/index.php/jfu/article/view/17187/11312
- Levita J., Musfiroh I., Indriyati W., Mustarichie R., 2010 The effect of soaking, washing and frying on the concentration of formaldehyde in Sange Belah salty fish. Jurnal Ilmu-ilmu Hayati dan Fisik 12(1):31–34.
- Male Y. T., Letsoin L. I., Siahaya N. A., 2017 [Analysis of formaldehyde content on noodle in several area in Ambon]. Jurnal Kementerian Perindustrian 13(2):5–10. [In Indonesian].
- Mudaffar R. A., 2018 [Qualitative and quantitative test of formaldehyde on apple, grape, and litchi for sale in Makassar]. Jurnal Perbal 6(3):....pages. [In Indonesian].
- Mudzkirah I., 2016 [Identification of borax and formalin on snack food in UIN Alaudin Makassar Canteen]. Type of publication, UIN Alauddin Makassar, no of pages. [In Indonesian].
- Muntaha A., Haitami, Hayati N., 2015 [Comparison of reduction formaldehyde level on boiled and hot water soaked tofu]. Medical Laboratory Technology Journal 1(2):84– 90. [In Indonesian].
- Murtini J. T., Riyanto R., Priyanto N., Hermana I., 2014 [Development of Formaldehyde on some kinds of seawater fish during storage in crushed ice]. JPB Perikanan 9(2):143–151. [In Indonesian].
- Pandey C. K., Agarwal A., Baronia A., Singh N., 2000 Toxicity of ingested formalin and its management. Human and Experiment Toxicology 360–366.
- Purawisastra S., Sahara E., 2011 [Absorption to Formalin by certain foodstuff and disappearance through hot water soaked]. PGM 34(1):63–74. [In Indonesian].
- Rachmawati N., Riyanto R., Ariyani F., 2007 [Identification of formaldehyde on kerapu macan fish (*Ephinephelus fuscoguttatus*) during cold storage]. Jurnal Pascapanen dan Bioteknologi Kelautan dan Perikanan 2(2):137–145. [In Indonesian].
- Rahmadhani F., Safrida, Djufri, 2017 [The effect of soaking a various of solution for reduction formalin level on kembung salty fish (*Scomber canagorta*) in Lambaro traditional market Aceh]. Jurnal Ilmiah Mahasiswa Keguruan dan Ilmu Pendidikan Unsyiah, volume, issue, pages. [In Indonesian].
- Riyanto R., Kusmarwati A., Dwiyitno, 2006 [Identification of formaldehyde foorming on kerapu fish (*Epinephelus fuscoguttatus*) during room temperature storage]. Jurnal Pascapanen dan Bioteknologi Kelautan dan Perikanan 1(2):111–116. [In Indonesian].

AACL Bioflux, 2020, Volume 13, Issue X. http://www.bioflux.com.ro/aacl

Shita A. E., 2016 [Selectivity of spectrophotometer analysis with Schiff's Reagen]. Type of publication, Universitas Negeri Yogyakarta, no of pages. [In Indonesian].

Sugiarti M., Anggo A. D., Riyadi P. H., 2014 [The effect of soaking in undercooking temperature and cooking method for reduction formalin level on cuttlefish]. Jurnal Pengolahan dan Bioteknologi Hasil Perikanan 3(2):90–98. [In Indonesian].

Sundari D., Almasyhuri, Lamid A., 2015 [The effect of cooking process for nutrition composition of protein source]. Media Litbangkes 25(4):235–242. [In Indonesian].

Suwanaruang T., 2018 Formalin contaminated in seafood and frozen meat at Somdet market, Kalasin Province. Journal of Environmental Protection 9:1286–1293.

Wijayanti N. S., Lukitasari M., 2016 [Analysis of formalin level and organoleptic test in salty fish in Madiun Market]. Jurnal Florea 3(1):59–64. [In Indonesian].

Wilianto W., Yudianto A., 2013 [The effect of formalin exposure on various level for DNA psoas major by PCR Lokus CSF1PO, D5S818, D13S317, D21S11 Analysis]. Jurnal Kedokteran Forensik Indonesia 15(1):11–16. [In Indonesian].

Yeasmin T., Reza M. S., Shikha F. H., Khan M. N. A., Kamal M., 2010 Quality changes in formalin treated rohu fish (*Labeo rohita*, Hamilton) during ice storage condition. Asian Journal of Agricultural Science 2(4):158–163.

Yulisa N., Asni E., Azrin M., 2014 [Formaldehyde test of gurame fish in Pekanbaru Traditional Market]. Publisher 1(2): pages. [In Indonesian].

Yusuf Y., Zuki Z., Last name MP, Amanda R. R., 2015 [The effect of various treatment for reduction formalin level on fish by spectrophotometry method]. J. Ris. Kim No abbreviations 8(2):182–188. [In Indonesian].

Received: 15 October 2020. Accepted: 2020. Published online: 2020.

Isny Maulidevi, Alma Ata University, Faculty of Health Sciences, Department of Nutrition, Jl. Brawijaya 99, 55183 Yogyakarta, Indonesia, e-mail: isnymaulidevi540@gmail.com

Nurul Andriyani, Alma Ata University, Faculty of Health Sciences, Department of Nutrition, Jl. Brawijaya 99, 55183 Yogyakarta, Indonesia, e-mail: nurulandriyani96@gmail.com

Ryan Salfarino, Alma Ata University, Faculty of Health Sciences, Department of Nutrition, Jl. Brawijaya 99, 55183 Yogyakarta, Indonesia, e-mail: ryan.salfarino@gmail.com

Veriani Aprilia, Alma Ata University, Faculty of Health Sciences, Department of Nutrition, Jl. Brawijaya 99, 55183 Yogyakarta, Indonesia, e-mail: verianiaprilia@almaata.ac.id This is an open-access article distributed under the terms of the Creative Commons Attribution License, which

This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution and reproduction in any medium, provided the original author and source are credited.

How to cite this article:

Maulidevi I., Andriyani N., Salfarino R., Aprilia V., 2020 The residue of formalin in catfish (*Clarias gariepinus*) after processing and storage for short periods. AACL Bioflux 13(x):

AACL Bioflux, 2020, Volume 13, Issue X. http://www.bioflux.com.ro/aacl



verianiaprilia verianiaprilia <verianiaprilia@almaata.ac.id>

Pengiriman revisi 2: 7 Desember 2020

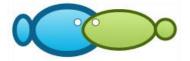
verianiaprilia verianiaprilia <verianiaprilia@almaata.ac.id> Kepada: Eni Kovacs <ek.bioflux@gmail.com>

Dear Kovacs, Here my revised article. thank you very much

Regards Veriani Aprilia [Kutipan teks disembunyikan]

rev_Aprilia_Clarias gariepinus_final (1).doc
165K

7 Desember 2020 pukul 12.03



The residue of formalin in catfish (*Clarias* gariepinus) after processing and storage for short periods

İsny Maulidevi, Nurul Andriyani, Ryan Salfarino, Veriani Aprilia

Department of Nutrition, Faculty of Health Sciences, Alma Ata University, Yogyakarta, Indonesia. Corresponding author: V. Aprilia, verianiaprilia@almaata.ac.id

Abstract. Fresh fish is a highly perishable product due to its high protein content. A preservative may be needed to inhibit it. Nowadays, many fishes were reported to be exposed to formalin. This study aimed to determine the effect of short period processing (washing, frying, boiling in short-period) and storage (at the room and at freezing temperatures) on the residue of formalin in catfish, as a food model. Catfish, *Clarias gariepinus*, were divided into 8 groups, e.g. before treatment, washing in airflow for 1 minute, frying in 200°C for 8 minutes, boiling in 100°C for 7 minutes, storage at room temperature ($29\pm1°C$) for 24 and 48 hours, storage at freezing temperature (-3°C) for 24 and 48 hours. Before treatment, the catfish was soaked in 1,480 ppm of formalin solutions. The residue of formalin was measured at the end of each treatment by using a spectrophotometer. Data were then analyzed with analysis of variance with SPS software. The results showed that there were significant effects of all treatments was lower than before treatments. The frying processing and storage at freezing temperatures of catfish. The formalin residue of all treatments was lower than before treatments in reducing the formalin residue. **Key Words**: fresh fish, preservative, food model, treatments, formalin residue.

Introduction. Fresh fish is a highly perishable product due to its high protein content for microbial activity (Wijayanti & Lukitasari 2016). A preservative maybe needs to keep the freshness, inhibit the decay and prolong the shelf life (Antoni 2010). Nowadays, there are many kinds of food-grade preservatives available in the market. Unfortunately, formalin is still frequently found in food products, including fresh fish, being used among fishermen mainly because it is effective in preserving the product, cheaper and accessible (Male et al 2017).

Formalin is a forty percent solution of formaldehyde in water. This compound is detected in the animal body as an intermediary metabolite. It also uses for several purposes, such as in medicine for specimen preservation and the treatment of hemorrhage (Pandey et al 2000). The preservatives activity of formalin is due to its ability to react with protein generating methylene compounds, which are more difficult to be decayed by microbes (Ichya'uddin 2016). In low concentrations, formalin is not toxic for humans, but in higher concentrations, it had negative effects such as acute toxicity and death (Pandey et al 2000). It also decreases the levels of body antioxidants, resulting in liver damage (Yulisa et al 2014). Other studies also reported the cases of eczema, eye irritation, respiratory tract irritation, cancer, increased growth rate of malignant neoplastic diseases and denaturation of DNA (Suwanaruang 2018; Wilianto & Yudianto 2013). Therefore, its uses are forbidden.

Formalin solution is not stable during storage and exposure to high temperatures (Kaneko et al 1977; Laksmiani et al 2015). Those are caused by desolvation or evaporation of formalin during washing, boiling, or frying (Muntaha et al 2015; Sundari et al 2015) and enzymatic process in storage (Rachmawati et al 2007). Formalin that was in contact with fish decreased up to 43-53% after 5 times rinsing in 250 mL of distilled water and boiling for 30 minutes (Yusuf et al 2015). Other studies were also reported that soaking in warm water and frying decreased the concentration of formalin up to

AACL Bioflux, 2020, Volume 13, Issue X. http://www.bioflux.com.ro/aacl

63.27% and 83.03%, respectively (Levita et al 2010). In daily practices, fish is usually washed in flowing water before processing, then fryed/boiled for a short time. However, there was no study explaining the influences of these processing and storage methods on the concentration of formalin.

The purpose of this study was to identify the effect of processing for a short time and storage (at room and freezing temperatures) on the residue of formalin. This study used catfish as a food model.

Material and Method

Material. The study used fresh catfish, purchased from the traditional market in Yogyakarta, Indonesia. The other used ingredients were formalin solution and Schiff's reagent. Formalin solution was made from 4 mL of formalin 37% in 1,000 mL of distilled water (Rachmawati et al 2007).

Method. The study was divided into two sections, i.e. 1) analyzing the effect of processing and 2) analyzing the effect of storage treatment. Fresh catfish were slaughtered, eviscerated, cleaned, and soaked in 1,480 ppm formalin solution (Rachmawati et al 2007; Sugiarti et al 2014). For the first study, catfish were divided into four groups, i.e.: controls (before treatment), washing, frying, and boiling treatments. Washing was done by watering the samples with running water for 1 minute, while frying was done by frying the samples in oil at 200°C for 8 minutes. Boiling treatment was done by putting the samples into boiled water (100°C) for 7 minutes (Sugiarti et al 2014). For the second study, storage treatments, the catfish was also divided into four groups: room temperature storage (29°C) for 24 hours and 48 hours and freezing temperatures storage (-3°C) for 24 hours and 48 hours (Handayani et al 2010). At the end of treatments, the concentration of formalin in samples was analyzed using a spectrophotometer. As the preparation step, a 5 g sample was dissolved in 25 mL of distilled water. One mL of supernatant was mixed with 2 mL of Schiff's reagent, then heated in a water bath at 60°C for 5 minutes. It was incubated for 10 minutes before measuring the absorbance at 520 nm (Shita 2016; Mudaffar 2018).

Statistical analysis. Data were presented in mean±SD and analyzed with: (1) one-way analysis of variance (ANOVA) to determine the differences among all processing treatments and the effect of storage periods on the concentration of formalin; (2) paired sample T-test to determine the differences between the same samples before and after processing treatments; (3) independent sample t-test to know the effect of temperature at the same storage period, and (4) two-way ANOVA to determine the combination effect of storage temperature and period on the concentration of formalin. All data were analyzed with SPSS software (version 16.0 for Windows; SPSS Inc., Chicago, IL, USA) at the 5% significance level.

Results

Effect of processing on the formalin residue of catfish. Table 1 shows that all treatments had significant effects on the residue of formalin (p<0.05). It can be seen from the p-value between the same samples before and after treatments. Among all treatments, washing shows the highest residue of formalin (p<0.05). Both boiling and frying had an almost similar residue of formalin (p>0.05). The lower residue of formalin in boiling and frying may be due to heating exposure.

Effect of processing on the formalin residue of catfish

Processing	Residue of formalin (ppm)		Δ residue of formalin		
treatments	Before treatment	After treatment	<i>before and after treatment</i> (%)		
Washing	1,420.17±66.94 ^{c1}	310.42±77.61 ^{b2}	78.14		
Frying	1,420.17±66.94 ^{c1}	14.18±6.69 ^{a2}	99.01		
Boiling	1,420.17±66.94 ^{c1}	27.50±9.11 ^{a2}	98.10		
Mean+SD in the same column with different alphabetical superscripts (a, b, or c) are significantly different					

(p<0.05) by using one-way ANOVA statistical analysis. Mean #SD in the same row with different number superscripts (1 or 2) are significantly different (p<0.05) by using paired sample T-test statistical analysis.

Effect of storage temperatures and periods on the formalin residue of catfish. Table 2 shows the effect of storage temperatures, of periods and of their combination on the concentration of formalin. Based on this data, the storage period influenced the residue of formalin decrease (p<0.05) during the first 24 hours, but after a storage period of 48 hours the decrease was not significantly different (p>0.05).

On the other hand, there were no differences in the residue of formalin stored at room and freezing temperature, suggesting that the temperature of storage did not affect the formalin residue (p>0.05). Although there were differences in the statistical results between the temperature effects and the storage periods effects, the combined effect of both variables showed no impact on the concentration of formalin (p>0.05).

Table 2

Effect of storage temperature and period on the formalin residue of catfish

Temperature	Periods (t) (hours)				p-value	9
(T)	0	24	48	Т	t	T*t
Room (29°C)	1,420.17±66.94 ^{a1} 1,420.17±66.94 ^{a1}	52.18±28.78 ^{a2}	40.64±3.93 ^{a2}	0.20	0.00	0.76
Freezing (-3°C)	1,420.17±66.94 ^{a1}	39.04±7.07 ^{a2}	13.65±4.23 ^{a2}	0.20	0.00	0.76

Mean±SD in the same column with different alphabetical superscripts (a, b, or c) are significantly different (p<0.05) and so are in the same row with different number superscript (1, 2, or 3). The p-value for T, t, and T*t was obtained from independent T-test, one-way ANOVA, and two-way ANOVA, respectively.

Discussion

The residue of formalin before treatment. The residue of formalin before and after treatment (either fish processing or storage) was significantly different (Table 1 and Table 2). In this study, the catfish specimens were submerged for 60 minutes in formalin solution of 1,480 ppm. The 60 minutes submersion period was chosen because the aldehyde compound may be readily bonded to the amino acids of fish in a small amount (Rahmadhani et al 2017; Baroroh & Kresnadipayana 2016). Formalin would attack lysine for the first time, followed by histidine and tyrosine, leading to the formation of a methylene compound (Ichya'uddin 2016). The protein-containing methylene compounds could not be digested by living things and microbes, especially pathogens, so that the shelf life would be longer (Purawisastra & Sahara 2011). The beneficial bacteria in the fish may also be killed by dehydration mechanism, as another effect of the existence of formalin in the body (Mudzkirah 2016).

The effect of processing on the formalin residue of catfish. The residue of formalin was reduced by 78.14% during washing treatment. The reduction was significant (p<0.05). The formalin was easily washed out because of its high solubility in the water, which is 4×10^5 mg L⁻¹ at 20°C (BPOM 2008; Sugiarti et al 2014). A study conducted by Yusuf et al (2015) showed the same trend, but the reduction was lower (with only 43%). The differences in the level of reduction can be affected by the duration of formalin

AACL Bioflux, 2020, Volume 13, Issue X. http://www.bioflux.com.ro/aacl

1092

Table 1

submersion. The longer the submersion time, the smaller the decrease, as more formalin was bonded with the fish protein.

In this study, frying also significantly reduced the residue of formalin (p<0.05). The reduction level was 99% that indicated most of the formalin disappeared during the frying process. Formalin was included in a volatile compound, especially above its boiling point (Joshi et al 2015). This study confirmed the results of Sugiarti et al (2014), showing a reduction level of formalin up to 99.28% after 1 hour of submersion followed by the frying process (no information about the period of frying process) in squid. Frying could denature the protein and also hydrolyze its bonding with formalin (Levita et al 2010). The high temperature in frying increased the kinetic energy and led to faster movement of the molecules that could damage the molecular bonds (Sugiarti et al 2014).

Heat processing by boiling reduced the residue of formalin with up to 98.1%, more than the washing process (p<0.05), but at the same degree as the frying process (p>0.05).The high reduction of formalin in the boiling process was due to the accelerated formalin dissolution rate at higher temperatures (Annisak 2019). This effect could be enhanced by boiling the sample in an open pan (Kamal et al 2017).

The effect of storage on the formalin residue of catfish. This experiment demonstrated that only the period of storage significantly influenced the residue of formalin (p<0.05), while the temperature of storage and the combination between temperature and period of storage did not (Table 2). The previous different studies showed that a storage at the room temperature or at the freezing temperature had similar effects on the formalin residue decrease (Jawahar et al 2017; Murtini et al 2014), possibly due to the same level of protein degradation by breaking the bonds between protein and formalin and releasing free compounds. At the room temperature, the protein degradation was essentially caused by the microbial activity, while at the freezing temperature it was caused mostly by physical destruction, leading to conformational and functional changes (Murtini et al 2014; Yeasmin et al 2010). Formalin in the free form is more reactive and can also be more easily degraded (Riyanto et al 2006; Rachmawati et al 2007).

Conclusions. This research concluded that short time processing and storage treatments decreased the concentration of formalin. Heat involvement determined a more effective decrease of concentration (the decrease level was >90%). In the storage study, the residue of formalin was significantly reduced by the period of storage, mainly during the first 24 hours, then the formalin concentration reduction was insignificant, up to 48 hours of storage.

Acknowledgements. The authors are grateful to the Indonesia Ministry of Research, Technology, and Higher Education for providing the financial support in pursuing this research project (Program Kreativitas Mahasiswa Kemeristekdikti 2019 Program).

Reference

- Annisak S. K., Indriyanti N. Y., Mulyani B., 2019 [Constructive controversy and incubation presented according to the representation of tetrahedral chemical learning according to critical thinking ability]. Jurnal Inovasi Pendidikan IPA 5(1):10–22. [In Indonesian].
- Antoni S., 2010 [Analysis of formaldehyde content on salty fish by spectrophotometry method in Tampan District Pekanbaru]. Thesis, Universitas Islam Negeri Sultan Syarif Kasim Riau, 1-53. [In Indonesian].

Baroroh R. D., Kresnadipayana D., 2016 [The effect of variety consentration of starfruit juice (*Averrhoa bilimbi* L.) on soaked tofu for formaldehyde reduction]. Thesis. Universitas Setia Budi, 1-2. [In Indonesian].

BPOM (Indonesian Food and Drug Authority) 2008 [Formaldehyde]. *Leaflet.* Jakarta: Badan POM RI. [In Indonesian].

AACL Bioflux, 2020, Volume 13, Issue X. http://www.bioflux.com.ro/aacl

1093

Commented [WU1]: Please replace the red text with the information required.

- Handayani A., Alimin, Rustiah W. O., 2010 [The effect of low temperature storage (freezer -3oc) on water and fat content in lemuru fish (*Sardinella longiceps*)]. Al-Kimia (3):64–75. [In Indonesian].
- Ichya'uddin M., 2014 [Analysis of formaldehyde content and organoleptic test on salty fish in several traditional markets in Tuban]. Thesis, Universitas Islam Negeri Maulana Malik Ibrahim, 1-112. [In Indonesian].
- Jawahar L. S., all the authors must be listed, 2017 Retention of Residual formaldehyde in treated Indian mackerel (*Rastrelliger kanagurta*) under iced storage and related food safety concern, Indian Journal of Fisheries 64(4):87–93.
- Josh, Bhatta, R., Paudel, P.N., Kafle, B.K., 2015 Formaldehyde content of selected fish from the wet markets of Kathmandu valley. International Food Research Journal 22(4):1434-1437.
- Kamal A. S., Uddin, M.M., Rahman, R., Samira, S., Rahman, M., Nandy, S., Khan, M.S., 2017 Time and Temperature effect on the residual concentration of formaldehyde in formalin treated samples of *Labeo rohita*. International Conference on Food Security and Nutrition, 1-6.
- Kaneko M., Wada Y., Fukui S., Kanno S., 1977 NII-Electronic Library Service. The Journal of Hygienic Chemistry 23(6):393–396.
- Laksmiani N., Widjaja I., Sonia, 2015 [Stability of formalin in different temperature and period of heating]. Jurnal Farmasi Udayana 4(2):76–81. [In Indonesian]. https://ojs.unud.ac.id/index.php/jfu/article/view/17187/11312
- Levita J., Musfiroh I., Indriyati W., Mustarichie R., 2010 The effect of soaking, washing and frying on the concentration of formaldehyde in Sange Belah salty fish. Jurnal Ilmu-ilmu Hayati dan Fisik 12(1):31–34.
- Male Y. T., Letsoin L. I., Siahaya N. A., 2017 [Analysis of formaldehyde content on noodle in several area in Ambon]. Jurnal Kementerian Perindustrian 13(2):5–10. [In Indonesian].
- Mudaffar R. A., 2018 [Qualitative and quantitative test of formaldehyde on apple, grape, and litchi for sale in Makassar]. Jurnal Perbal 6(3):59-65. [In Indonesian].
- Mudzkirah I., 2016 [Identification of borax and formalin on snack food in UIN Alaudin Makassar Canteen]. Thesis, UIN Alauddin Makassar, 1-95. [In Indonesian].
- Muntaha A., Haitami, Hayati N., 2015 [Comparison of reduction formaldehyde level on boiled and hot water soaked tofu]. Medical Laboratory Technology Journal 1(2):84– 90. [In Indonesian].
- Murtini J. T., Riyanto R., Priyanto N., Hermana I., 2014 [Development of Formaldehyde on some kinds of seawater fish during storage in crushed ice]. JPB Perikanan 9(2):143–151. [In Indonesian].
- Pandey C. K., Agarwal A., Baronia A., Singh N., 2000 Toxicity of ingested formalin and its management. Human and Experiment Toxicology 360–366.
- Purawisastra S., Sahara E., 2011 [Absorption to Formalin by certain foodstuff and disappearance through hot water soaked]. PGM 34(1):63–74. [In Indonesian].
- Rachmawati N., Riyanto R., Ariyani F., 2007 [Identification of formaldehyde on kerapu macan fish (*Ephinephelus fuscoguttatus*) during cold storage]. Jurnal Pascapanen dan Bioteknologi Kelautan dan Perikanan 2(2):137–145. [In Indonesian].
- Rahmadhani F., Safrida, Djufri, 2017 [The effect of soaking a various of solution for reduction formalin level on kembung salty fish (*Scomber canagorta*) in Lambaro traditional market Aceh]. Jurnal Ilmiah Mahasiswa Pendidikan Biologi, 2(3): 1-12. [In Indonesian].
- Riyanto R., Kusmarwati A., Dwiyitno, 2006 [Identification of formaldehyde foorming on kerapu fish (*Epinephelus fuscoguttatus*) during room temperature storage]. Jurnal Pascapanen dan Bioteknologi Kelautan dan Perikanan 1(2):111–116. [In Indonesian].
- Shita A. E., 2016 [Selectivity of spectrophotometer analysis with Schiff's Reagent]. Thesis, Universitas Negeri Yogyakarta, 1-87. [In Indonesian].
- Sugiarti M., Anggo A. D., Riyadi P. H., 2014 [The effect of soaking in undercooking temperature and cooking method for reduction formalin level on cuttlefish]. Jurnal Pengolahan dan Bioteknologi Hasil Perikanan 3(2):90–98. [In Indonesian].

AACL Bioflux, 2020, Volume 13, Issue X. http://www.bioflux.com.ro/aacl

Sundari D., Almasyhuri, Lamid A., 2015 [The effect of cooking process for nutrition composition of protein source]. Media Litbangkes 25(4):235–242. [In Indonesian].

Suwanaruang T., 2018 Formalin contaminated in seafood and frozen meat at Somdet market, Kalasin Province. Journal of Environmental Protection 9:1286-1293.

Wijayanti N. S., Lukitasari M., 2016 [Analysis of formalin level and organoleptic test in salty fish in Madiun Market]. Jurnal Florea 3(1):59-64. [In Indonesian].

Wilianto W., Yudianto A., 2013 [The effect of formalin exposure on various level for DNA psoas major by PCR Lokus CSF1PO, D5S818, D13S317, D21S11 Analysis]. Jurnal Kedokteran Forensik Indonesia 15(1):11-16. [In Indonesian].

Yeasmin T., Reza M. S., Shikha F. H., Khan M. N. A., Kamal M., 2010 Quality changes in formalin treated rohu fish (Labeo rohita, Hamilton) during ice storage condition. Asian Journal of Agricultural Science 2(4):158–163.

Yulisa N., Asni E., Azrin M., 2014 [Formaldehyde test of gurame fish in Pekanbaru Traditional Market]. Jurnal Online Mahasiswa 1(2): 1-12. [In Indonesian].

Yusuf Y., Zuki Z., Amanda R. R., 2015 [The effect of various treatment for reduction formalin level on fish by spectrophotometry method]. Jurnal Riset Kimia 8(2):182-188. [In Indonesian].

Received: 15 October 2020. Accepted: 2020. Published online: 2020. Authors:

Isny Maulidevi, Alma Ata University, Faculty of Health Sciences, Department of Nutrition, Jl. Brawijaya 99,

55183 Yogyakarta, Indonesia, e-mail: isnymaulidevi540@gmail.com Nurul Andriyani, Alma Ata University, Faculty of Health Sciences, Department of Nutrition, Jl. Brawijaya 99, 55183 Yogyakarta, Indonesia, e-mail: nurulandriyani96@gmail.com

Ryan Salfarino, Alma Ata University, Faculty of Health Sciences, Department of Nutrition, Jl. Brawijaya 99, 55183 Yogyakarta, Indonesia, e-mail: ryan.salfarino@gmail.com

Veriani Aprilia, Alma Ata University, Faculty of Health Sciences, Department of Nutrition, Jl. Brawijaya 99, 55183 Yoqyakarta, Indonesia, e-mail: verianiaprilia@almaata.ac.id

This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution and reproduction in any medium, provided the original author and source are credited.

How to cite this article:

Maulidevi I., Andriyani N., Salfarino R., Aprilia V., 2020 The residue of formalin in catfish (Clarias gariepinus) after processing and storage for short periods. AACL Bioflux 13(x):

AACL Bioflux, 2020, Volume 13, Issue X. http://www.bioflux.com.ro/aacl



verianiaprilia verianiaprilia <verianiaprilia@almaata.ac.id>

Penetapan revisi 1: 13 Desember 2020

Adjustments

4 pesan

7/18/23, 1:45 PM Email Universitas Alma Ata - Adjustments Eni Kovacs <ek.bioflux@gmail.com> Kepada: verianiaprilia verianiaprilia <verianiaprilia@almaata.ac.id>

13 Desember 2020 pukul 03.06

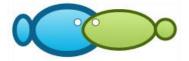
Dear Veriani Aprilia,

Please find the manuscript attached and make the necessary adjustments.

Thank you.

Kind regards, Editor AACL <u>Bioflux</u> Eniko Kovacs

Aprilia_Clarias gariepinus_final.doc 170K



The residue of formalin in catfish (*Clarias* gariepinus) after processing and storage for short periods

İsny Maulidevi, Nurul Andriyani, Ryan Salfarino, Veriani Aprilia

Department of Nutrition, Faculty of Health Sciences, Alma Ata University, Yogyakarta, Indonesia. Corresponding author: V. Aprilia, verianiaprilia@almaata.ac.id

Abstract. Fresh fish is a highly perishable product due to its high protein content. A preservative may be needed to inhibit it. Nowadays, many fishes were reported to be exposed to formalin. This study aimed to determine the effect of short period processing (washing, frying, boiling in short-period) and storage (at the room and at freezing temperatures) on the residue of formalin in catfish, as a food model. Catfish, *Clarias gariepinus*, were divided into 8 groups, e.g. before treatment, washing in airflow for 1 minute, frying in 200°C for 8 minutes, boiling in 100°C for 7 minutes, storage at room temperature ($29\pm1°C$) for 24 and 48 hours, storage at freezing temperature (-3°C) for 24 and 48 hours. Before treatment, the catfish was soaked in 1,480 ppm of formalin solutions. The residue of formalin was measured at the end of each treatment by using a spectrophotometer. Data were then analyzed with analysis of variance with SPS software. The results showed that there were significant effects of all treatments was lower than before treatments. The frying processing and storage at freezing temperatures of catfish. The formalin residue of all treatments was lower than before treatments in reducing the formalin residue. **Key Words**: fresh fish, preservative, food model, treatments, formalin residue.

Introduction. Fresh fish is a highly perishable product due to its high protein content for microbial activity (Wijayanti & Lukitasari 2016). A preservative maybe needs to keep the freshness, inhibit the decay and prolong the shelf life (Antoni 2010). Nowadays, there are many kinds of food-grade preservatives available in the market. Unfortunately, formalin is still frequently found in food products, including fresh fish, being used among fishermen mainly because it is effective in preserving the product, cheaper and accessible (Male et al 2017).

Formalin is a forty percent solution of formaldehyde in water. This compound is detected in the animal body as an intermediary metabolite. It also uses for several purposes, such as in medicine for specimen preservation and the treatment of hemorrhage (Pandey et al 2000). The preservatives activity of formalin is due to its ability to react with protein generating methylene compounds, which are more difficult to be decayed by microbes (Ichya'uddin 2016). In low concentrations, formalin is not toxic for humans, but in higher concentrations, it had negative effects such as acute toxicity and death (Pandey et al 2000). It also decreases the levels of body antioxidants, resulting in liver damage (Yulisa et al 2014). Other studies also reported the cases of eczema, eye irritation, respiratory tract irritation, cancer, increased growth rate of malignant neoplastic diseases and denaturation of DNA (Suwanaruang 2018; Wilianto & Yudianto 2013). Therefore, its uses are forbidden.

Formalin solution is not stable during storage and exposure to high temperatures (Kaneko et al 1977; Laksmiani et al 2015). Those are caused by desolvation or evaporation of formalin during washing, boiling, or frying (Fellows 2015; Harmita 2016; Muntaha et al 2015; Sundari et al 2015) and enzymatic process in storage (Rachmawati et al 2007). Formalin that was in contact with fish decreased up to 43-53% after 5 times rinsing in 250 mL of distilled water and boiling for 30 minutes (Yusuf et al 2015). Other studies were also reported that soaking in warm water and frying decreased the

AACL Bioflux, 2020, Volume 13, Issue X. http://www.bioflux.com.ro/aacl

concentration of formalin up to 63.27% and 83.03%, respectively (Levita et al 2010). In daily practices, fish is usually washed in flowing water before processing, then fryed/boiled for a short time. However, there was no study explaining the influences of these processing and storage methods on the concentration of formalin.

The purpose of this study was to identify the effect of processing for a short time and storage (at room and freezing temperatures) on the residue of formalin. This study used catfish, *Clarias gariepinus*, as a food model.

Material and Method

Material. The study used fresh catfish, purchased from the traditional market in Yogyakarta, Indonesia. The other used ingredients were formalin solution and Schiff's reagent. Formalin solution was made from 4 mL of formalin 37% in 1,000 mL of distilled water (Rachmawati et al 2007).

Method. The study was divided into two sections, i.e. 1) analyzing the effect of processing and 2) analyzing the effect of storage treatment. Fresh catfish were slaughtered, eviscerated, cleaned, and soaked in 1,480 ppm formalin solution (Rachmawati et al 2007; Sugiarti et al 2014). For the first study, catfish were divided into four groups, i.e.: controls (before treatment), washing, frying, and boiling treatments. Washing was done by watering the samples with running water for 1 minute, while frying was done by frying the samples in oil at 200°C for 8 minutes. Boiling treatment was done by putting the samples into boiled water (100°C) for 7 minutes (Sugiarti et al 2014). For the second study, storage treatments, the catfish was also divided into four groups: room temperature storage (29°C) for 24 hours and 48 hours and freezing temperatures storage (-3°C) for 24 hours and 48 hours (Handayani et al 2010). At the end of treatments, the concentration of formalin in samples was analyzed using a spectrophotometer. As the preparation step, a 5 g sample was dissolved in 25 mL of distilled water. One mL of supernatant was mixed with 2 mL of Schiff's reagent, then heated in a water bath at 60°C for 5 minutes. It was incubated for 10 minutes before measuring the absorbance at 520 nm (Shita 2016; Mudaffar 2018).

Statistical analysis. Data were presented in mean±SD and analyzed with: (1) one-way analysis of variance (ANOVA) to determine the differences among all processing treatments and the effect of storage periods on the concentration of formalin; (2) paired sample T-test to determine the differences between the same samples before and after processing treatments; (3) independent sample t-test to know the effect of temperature at the same storage period, and (4) two-way ANOVA to determine the combination effect of storage temperature and period on the concentration of formalin. All data were analyzed with SPSS software (version 16.0 for Windows; SPSS Inc., Chicago, IL, USA) at the 5% significance level.

Results

Effect of processing on the formalin residue of C. gariepinus. Table 1 shows that all treatments had significant effects on the residue of formalin (p<0.05). It can be seen from the p-value between the same samples before and after treatments. Among all treatments, washing shows the highest residue of formalin (p<0.05). Both boiling and frying had an almost similar residue of formalin (p>0.05). The lower residue of formalin in boiling and frying may be due to heating exposure.

Effect of processing on the formalin residue of Clarias gariepinus

Processing	Residue of formalin (ppm)		Δ residue of formalin
treatments	Before treatment	After treatment	<i>before and after treatment</i> (%)
Washing	1,420.17±66.94 ^{c1}	310.42±77.61 ^{b2}	78.14
Frying	1,420.17±66.94 ^{c1}	14.18±6.69 ^{a2}	99.01
Boiling	1,420.17±66.94 ^{c1}	27.50±9.11 ^{a2}	98.10

Mean \pm SD in the same column with different alphabetical superscripts (a, b, or c) are significantly different (p<0.05) by using one-way ANOVA statistical analysis. Mean \pm SD in the same row with different number superscripts (1 or 2) are significantly different (p<0.05) by using paired sample T-test statistical analysis.

Effect of storage temperatures and periods on the formalin residue of C. gariepinus. Table 2 shows the effect of storage temperatures, of periods and of their combination on the concentration of formalin. Based on this data, the storage period influenced the residue of formalin decrease (p<0.05) during the first 24 hours, but after a storage period of 48 hours the decrease was not significantly different (p>0.05).

On the other hand, there were no differences in the residue of formalin stored at room and freezing temperature, suggesting that the temperature of storage did not affect the formalin residue (p>0.05). Although there were differences in the statistical results between the temperature effects and the storage periods effects, the combined effect of both variables showed no impact on the concentration of formalin (p>0.05).

Table 2

Effect of storage temperature and period on the formalin residue of Clarias gariepinus

Temperature	Per	riods (t) (hours)			p-value	9
<i>(T)</i>	0	24	48	Т	t	T*t
Room (29°C)	1,420.17±66.94 ^{a1}	52.18±28.78 ^{a2}	40.64±3.93 ^{a2}	0.20	0.00	0.76
Freezing (-3°C)	1,420.17±66.94 ^{a1}	39.04±7.07 ^{a2}	13.65±4.23 ^{a2}	0.20	0.00	0.76

Mean±SD in the same column with different alphabetical superscripts (a, b, or c) are significantly different (p<0.05) and so are in the same row with different number superscript (1, 2, or 3). The p-value for T, t, and T*t was obtained from independent T-test, one-way ANOVA, and two-way ANOVA, respectively.

Discussion

The residue of formalin before treatment. The residue of formalin before and after treatment (either fish processing or storage) was significantly different (Table 1 and Table 2). In this study, the *C. gariepinus* specimens were submerged for 60 minutes in formalin solution of 1,480 ppm. The 60 minutes submersion period was chosen because the aldehyde compound may be readily bonded to the amino acids of fish in a small amount (Rahmadhani et al 2017; Baroroh & Kresnadipayana 2016). Formalin would attack lysine for the first time, followed by histidine and tyrosine, leading to the formation of a methylene compound (Ichya'uddin 2016). The protein-containing methylene compounds could not be digested by living things and microbes, especially pathogens, so that the shelf life would be longer (Purawisastra & Sahara 2011). The beneficial bacteria in the fish may also be killed by dehydration mechanism, as another effect of the existence of formalin in the body (Mudzkirah 2016).

The effect of processing on the formalin residue of C. gariepinus. The residue of formalin was reduced by 78.14% during washing treatment. The reduction was significant (p<0.05). The formalin was easily washed out because of its high solubility in the water, which is 4 x 10⁵ mg L⁻¹ at 20°C (BPOM 2008; Sugiarti et al 2014). A study conducted by Yusuf et al (2015) showed the same trend, but the reduction was lower (with only 43%). The differences in the level of reduction can be affected by the duration

AACL Bioflux, 2020, Volume 13, Issue X. http://www.bioflux.com.ro/aacl

1092

Table 1

of formalin submersion. The longer the submersion time, the smaller the decrease, as more formalin was bonded with the fish protein.

In this study, frying also significantly reduced the residue of formalin (p<0.05). The reduction level was 99% that indicated most of the formalin disappeared during the frying process. Formalin was included in a volatile compound, especially above its boiling point (Joshi et al 2015). This study confirmed the results of Sugiarti et al (2014), showing a reduction level of formalin up to 99.28% after 1 hour of submersion followed by the frying process (no information about the period of frying process) in squid. Frying could denature the protein and also hydrolyze its bonding with formalin (Levita et al 2010). The high temperature in frying increased the kinetic energy and led to faster movement of the molecules that could damage the molecular bonds (Sugiarti et al 2014).

Heat processing by boiling reduced the residue of formalin with up to 98.1%, more than the washing process (p<0.05), but at the same degree as the frying process (p>0.05). The high reduction of formalin in the boiling process was due to the accelerated formalin dissolution rate at higher temperatures (Annisak 2019). This effect could be enhanced by boiling the sample in an open pan (Kamal et al 2017).

The effect of storage on the formalin residue of *C. gariepinus*. This experiment demonstrated that only the period of storage significantly influenced the residue of formalin (p<0.05), while the temperature of storage and the combination between temperature and period of storage did not (Table 2). The previous different studies showed that a storage at the room temperature or at the freezing temperature had similar effects on the formalin residue decrease (Jawahar et al 2017; Murtini et al 2014), possibly due to the same level of protein degradation by breaking the bonds between protein and formalin and releasing free compounds. At the room temperature, the protein degradation was essentially caused by the microbial activity, while at the freezing temperature it was caused mostly by physical destruction, leading to conformational and functional changes (Murtini et al 2014; Yeasmin et al 2010). Formalin in the free form is more reactive and can also be more easily degraded (Riyanto et al 2006; Rachmawati et al 2007).

Conclusions. This research concluded that short time processing and storage treatments decreased the concentration of formalin. Heat involvement determined a more effective decrease of concentration (the decrease level was >90%). In the storage study, the residue of formalin was significantly reduced by the period of storage, mainly during the first 24 hours, then the formalin concentration reduction was insignificant, up to 48 hours of storage.

Acknowledgements. The authors are grateful to the Indonesian Ministry of Research, Technology, and Higher Education for providing the financial support in pursuing this research project (Program Kreativitas Mahasiswa Kemeristekdikti 2019 Program).

Reference

- Annisak S. K., Indriyanti N. Y., Mulyani B., 2019 [Constructive controversy and incubation presented according to the representation of tetrahedral chemical learning according to critical thinking ability]. Jurnal Inovasi Pendidikan IPA 5(1):10–22. [In Indonesian].
- Antoni S., 2010 [Analysis of formaldehyde content on salty fish by spectrophotometry method in Tampan District Pekanbaru]. Thesis, Universitas Islam Negeri Sultan Syarif Kasim Riau, Indonesia, 53 p. [In Indonesian].
- Baroroh R. D., Kresnadipayana D., 2016 [The effect of variety consentration of starfruit juice (*Averrhoa bilimbi* L.) on soaked tofu for formaldehyde reduction]. Thesis, Universitas Setia Budi, Indonesia, p. [In Indonesian].
- Fellows P. J., 2015 [Food processing technology]. Publisher, no of pages EGC Jakarta. ISBN: 979-044-612-0. [In Indonesian].

AACL Bioflux, 2020, Volume 13, Issue X. http://www.bioflux.com.ro/aacl

- Handayani A., Alimin, Rustiah W. O., 2010 [The effect of low temperature storage (freezer -3oc) on water and fat content in lemuru fish (*Sardinella longiceps*)]. Al-Kimia (3):64–75. [In Indonesian].
- Harmita, 2016 [Determination substance and availability of pharmaceuticals content]. Publisher, Jakarta: EGC. No of pages ISBN: 979-044-773-8. [In Indonesian].
- Ichya'uddin M., 2014 [Analysis of Formaldehyde content and organoleptic test on salty fish in several traditional markets in Tuban]. Thesis, Universitas Islam Negeri Maulana Malik Ibrahim, Indonesia, 112 p. [In Indonesian].
- Jawahar L. S., John C., Shafeekh M., Anupama T. K., Sankar T. V., 2017 Retention of residual formaldehyde in treated Indian mackerel (*Rastrelliger kanagurta*) under iced storage and related food safety concern. Indian Journal of Fisheries 64(4):87– 93.
- Joshi R., Bhatta R., Paudel P. N., Kafle B. K., 2015 Formaldehyde content of selected fish from the wet markets of Kathmandu valley. International Food Research Journal 22(4):1434-1437.
- Kamal A. S., Uddin M. M., Rahman R., Samira S., Rahman M., Nandy S., Khan M. S., 2017 Time and temperature effect on the residual concentration of formaldehyde in formalin treated samples of *Labeo rohita*. International Conference on Food Security and Nutrition, 6 p.
- Kaneko M., Wada Y., Fukui S., Kanno S., 1977 NII-Electronic library service. The Journal of Hygienic Chemistry 23(6):393–396.
- Laksmiani N., Widjaja I., Sonia, 2015 [Stability of formalin in different temperature and period of heating]. Jurnal Farmasi Udayana 4(2):76–81.
- Levita J., Musfiroh I., Indriyati W., Mustarichie R., 2010 The effect of soaking, washing and frying on the concentration of formaldehyde in Sange Belah salty fish. Jurnal Ilmu-ilmu Hayati dan Fisik 12(1):31-34.
- Male Y. T., Letsoin L. I., Siahaya N. A., 2017 [Analysis of formaldehyde content on noodle in several area in Ambon]. Jurnal Kementerian Perindustrian 13(2):5–10. [In Indonesian].
- Mudaffar R. A., 2018 [Qualitative and quantitative test of formaldehyde on apple, grape, and litchi for sale in Makassar]. Jurnal Perbal 6(3):59-65. [In Indonesian].
- Mudzkirah I., 2016 [Identification of borax and formalin on snack food in UIN Alaudin Makassar Canteen]. Thesis, UIN Alauddin Makassar, Indonesia, 95 p. [In Indonesian].
- Muntaha A., Haitami, Hayati N., 2015 [Comparison of reduction formaldehyde level on boiled and hot water soaked tofu]. Medical Laboratory Technology Journal 1(2):84– 90. [In Indonesian].
- Murtini J. T., Riyanto R., Priyanto N., Hermana I., 2014 [Development of Formaldehyde on some kinds of seawater fish during storage in crushed ice]. JPB Perikanan 9(2):143–151. [In Indonesian].
- Pandey C. K., Agarwal A., Baronia A., Singh N., 2000 Toxicity of ingested formalin and its management. Human and Experiment Toxicology 360–366.
- Purawisastra S., Sahara E., 2011 [Absorption to formalin by certain foodstuff and disappearance through hot water soaked]. PGM 34(1):63–74. [In Indonesian].
- Rachmawati N., Riyanto R., Ariyani F., 2007 [Identification of formaldehyde on kerapu macan fish (*Ephinephelus fuscoguttatus*) during cold storage]. Jurnal Pascapanen dan Bioteknologi Kelautan dan Perikanan 2(2):137–145. [In Indonesian].
- Rahmadhani F., Safrida, Djufri, 2017 [The effect of soaking a various of solution for reduction formalin level on kembung salty fish (*Scomber canagorta*) in Lambaro traditional market Aceh]. Jurnal Ilmiah Mahasiswa Pendidikan Biologi 2(3):1-12. [In Indonesian].
- Riyanto R., Kusmarwati A., Dwiyitno, 2006 [Identification of formaldehyde foorming on kerapu fish (*Epinephelus fuscoguttatus*) during room temperature storage]. Jurnal Pascapanen dan Bioteknologi Kelautan dan Perikanan 1(2):111–116. [In Indonesian].
- Shita A. E., 2016 [Selectivity of spectrophotometer analysis with Schiff's reagent]. Thesis, Universitas Negeri Yogyakarta, Indonesia, 87 p. [In Indonesian].

AACL Bioflux, 2020, Volume 13, Issue X. http://www.bioflux.com.ro/aacl

Sugiarti M., Anggo A. D., Riyadi P. H., 2014 [The effect of soaking in undercooking temperature and cooking method for reduction formalin level on cuttlefish]. Jurnal Pengolahan dan Bioteknologi Hasil Perikanan 3(2):90–98. [In Indonesian].

Sundari D., Almasyhuri, Lamid A., 2015 [The effect of cooking process for nutrition composition of protein source]. Media Litbangkes 25(4):235–242. [In Indonesian].
 Suwanaruang T., 2018 Formalin contaminated in seafood and frozen meat at Somdet

market, Kalasin Province. Journal of Environmental Protection 9:1286–1293. Wijayanti N. S., Lukitasari M., 2016 [Analysis of formalin level and organoleptic test in

salty fish in Madiun Market]. Jurnal Florea 3(1):59–64. [In Indonesian].

Wilianto W., Yudianto A., 2013 [The effect of formalin exposure on various level for DNA psoas major by PCR Lokus CSF1PO, D5S818, D13S317, D21S11 Analysis]. Jurnal Kedokteran Forensik Indonesia 15(1):11–16. [In Indonesian].

Yeasmin T., Reza M. S., Shikha F. H., Khan M. N. A., Kamal M., 2010 Quality changes in formalin treated rohu fish (*Labeo rohita*, Hamilton) during ice storage condition. Asian Journal of Agricultural Science 2(4):158–163.

Yulisa N., Asni E., Azrin M., 2014 [Formaldehyde test of gurame fish in Pekanbaru Traditional Market]. Jurnal Online Mahasiswa 1(2):1-12. [In Indonesian].

Yusuf Y., Zuki Z., Amanda R. R., 2015 [The effect of various treatment for reduction formalin level on fish by spectrophotometry method]. Jurnal Riset Kimia 8(2):182–188. [In Indonesian].

*** BPOM, Indonesian Food and Drug Authority, 2008 [Formaldehyde]. Badan POM RI, Jakarta, Indonesia. [In Indonesian]. Commented [WU1]: Please specify what kind ogf thesis: PhD? MSc?

Received: 15 October 2020. Accepted: 2020. Published online: 2020. Authors:

Isny Maulidevi, Alma Ata University, Faculty of Health Sciences, Department of Nutrition, Jl. Brawijaya 99, 55183 Yogyakarta, Indonesia, e-mail: isnymaulidevi540@gmail.com

Nurul Andriyani, Alma Ata University, Faculty of Health Sciences, Department of Nutrition, Jl. Brawijaya 99, 55183 Yogyakarta, Indonesia, e-mail: nurulandriyani96@gmail.com

Ryan Salfarino, Alma Ata University, Faculty of Health Sciences, Department of Nutrition, Jl. Brawijaya 99, 55183 Yogyakarta, Indonesia, e-mail: ryan.salfarino@gmail.com

Veriani Aprilia, Alma Ata University, Faculty of Health Sciences, Department of Nutrition, Jl. Brawijaya 99, 55183 Yogyakarta, Indonesia, e-mail: verianiaprilia@almaata.ac.id This is an open-access article distributed under the terms of the Creative Commons Attribution License, which

This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution and reproduction in any medium, provided the original author and source are credited.

How to cite this article:

Maulidevi I., Andriyani N., Salfarino R., Aprilia V., 2020 The residue of formalin in catfish (*Clarias gariepinus*) after processing and storage for short periods. AACL Bioflux 13(x):

AACL Bioflux, 2020, Volume 13, Issue X. http://www.bioflux.com.ro/aacl



Pengiriman hasil penetapan revisi 1: 15 Desember 2020

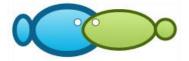
verianiaprilia verianiaprilia <verianiaprilia@almaata.ac.id> Kepada: Eni Kovacs <ek.bioflux@gmail.com> 15 Desember 2020 pukul 02.01

Dear Kovacs

Please find my adjustment below. Thank you

Regards Verian Aprilia [Kutipan teks disembunyikan]

Aprilia_Clarias gariepinus_final_4.doc 172K



The residue of formalin in catfish (*Clarias* gariepinus) after processing and storage for short periods

İsny Maulidevi, Nurul Andriyani, Ryan Salfarino, Veriani Aprilia

Department of Nutrition, Faculty of Health Sciences, Alma Ata University, Yogyakarta, Indonesia. Corresponding author: V. Aprilia, verianiaprilia@almaata.ac.id

Abstract. Fresh fish is a highly perishable product due to its high protein content. A preservative may be needed to inhibit it. Nowadays, many fishes were reported to be exposed to formalin. This study aimed to determine the effect of short period processing (washing, frying, boiling in short-period) and storage (at the room and at freezing temperatures) on the residue of formalin in catfish, as a food model. Catfish, *Clarias gariepinus*, were divided into 8 groups, e.g. before treatment, washing in airflow for 1 minute, frying in 200°C for 8 minutes, boiling in 100°C for 7 minutes, storage at room temperature ($29\pm1°C$) for 24 and 48 hours, storage at freezing temperature (-3°C) for 24 and 48 hours. Before treatment, the catfish was soaked in 1,480 ppm of formalin solutions. The residue of formalin was measured at the end of each treatment by using a spectrophotometer. Data were then analyzed with analysis of variance with SPS software. The results showed that there were significant effects of all treatments was lower than before treatments. The frying processing and storage at freezing temperatures of catfish. The formalin residue of all treatments was lower than before treatments in reducing the formalin residue. **Key Words**: fresh fish, preservative, food model, treatments, formalin residue.

Introduction. Fresh fish is a highly perishable product due to its high protein content for microbial activity (Wijayanti & Lukitasari 2016). A preservative maybe needs to keep the freshness, inhibit the decay and prolong the shelf life (Antoni 2010). Nowadays, there are many kinds of food-grade preservatives available in the market. Unfortunately, formalin is still frequently found in food products, including fresh fish, being used among fishermen mainly because it is effective in preserving the product, cheaper and accessible (Male et al 2017).

Formalin is a forty percent solution of formaldehyde in water. This compound is detected in the animal body as an intermediary metabolite. It also uses for several purposes, such as in medicine for specimen preservation and the treatment of hemorrhage (Pandey et al 2000). The preservatives activity of formalin is due to its ability to react with protein generating methylene compounds, which are more difficult to be decayed by microbes (Ichya'uddin 2016). In low concentrations, formalin is not toxic for humans, but in higher concentrations, it had negative effects such as acute toxicity and death (Pandey et al 2000). It also decreases the levels of body antioxidants, resulting in liver damage (Yulisa et al 2014). Other studies also reported the cases of eczema, eye irritation, respiratory tract irritation, cancer, increased growth rate of malignant neoplastic diseases and denaturation of DNA (Suwanaruang 2018; Wilianto & Yudianto 2013). Therefore, its uses are forbidden.

Formalin solution is not stable during storage and exposure to high temperatures (Kaneko et al 1977; Laksmiani et al 2015). Those are caused by desolvation or evaporation of formalin during washing, boiling, or frying (Fellows 2000; Hayun et al 2017; Muntaha et al 2015; Sundari et al 2015) and enzymatic process in storage (Rachmawati et al 2007). Formalin that was in contact with fish decreased up to 43-53% after 5 times rinsing in 250 mL of distilled water and boiling for 30 minutes (Yusuf et al 2015). Other studies were also reported that soaking in warm water and frying decreased

AACL Bioflux, 2020, Volume 13, Issue X. http://www.bioflux.com.ro/aacl

1090

Commented [S1]: I replace my reference Fellows 2015 with Fellows 2000; and Harmita 2016 with Hayun et al 2017 the concentration of formalin up to 63.27% and 83.03%, respectively (Levita et al 2010). In daily practices, fish is usually washed in flowing water before processing, then fryed/boiled for a short time. However, there was no study explaining the influences of these processing and storage methods on the concentration of formalin.

The purpose of this study was to identify the effect of processing for a short time and storage (at room and freezing temperatures) on the residue of formalin. This study used catfish, *Clarias gariepinus*, as a food model.

Material and Method

Material. The study used fresh catfish, purchased from the traditional market in Yogyakarta, Indonesia. The other used ingredients were formalin solution and Schiff's reagent. Formalin solution was made from 4 mL of formalin 37% in 1,000 mL of distilled water (Rachmawati et al 2007).

Method. The study was divided into two sections, i.e. 1) analyzing the effect of processing and 2) analyzing the effect of storage treatment. Fresh catfish were slaughtered, eviscerated, cleaned, and soaked in 1,480 ppm formalin solution (Rachmawati et al 2007; Sugiarti et al 2014). For the first study, catfish were divided into four groups, i.e.: controls (before treatment), washing, frying, and boiling treatments. Washing was done by watering the samples with running water for 1 minute, while frying was done by frying the samples in oil at 200°C for 8 minutes. Boiling treatment was done by putting the samples into boiled water (100°C) for 7 minutes (Sugiarti et al 2014). For the second study, storage treatments, the catfish was also divided into four groups: room temperature storage (29°C) for 24 hours and 48 hours and freezing temperatures storage (-3°C) for 24 hours and 48 hours (Handayani et al 2010). At the end of treatments, the concentration of formalin in samples was analyzed using a spectrophotometer. As the preparation step, a 5 g sample was dissolved in 25 mL of distilled water. One mL of supernatant was mixed with 2 mL of Schiff's reagent, then heated in a water bath at 60°C for 5 minutes. It was incubated for 10 minutes before measuring the absorbance at 520 nm (Shita 2016; Mudaffar 2018).

Statistical analysis. Data were presented in mean±SD and analyzed with: (1) one-way analysis of variance (ANOVA) to determine the differences among all processing treatments and the effect of storage periods on the concentration of formalin; (2) paired sample T-test to determine the differences between the same samples before and after processing treatments; (3) independent sample t-test to know the effect of temperature at the same storage period, and (4) two-way ANOVA to determine the combination effect of storage temperature and period on the concentration of formalin. All data were analyzed with SPSS software (version 16.0 for Windows; SPSS Inc., Chicago, IL, USA) at the 5% significance level.

Results

Effect of processing on the formalin residue of C. gariepinus. Table 1 shows that all treatments had significant effects on the residue of formalin (p<0.05). It can be seen from the p-value between the same samples before and after treatments. Among all treatments, washing shows the highest residue of formalin (p<0.05). Both boiling and frying had an almost similar residue of formalin (p>0.05). The lower residue of formalin in boiling and frying may be due to heating exposure.

Effect of processing on the formalin residue of Clarias gariepinus

Processing	Residue of formalin (ppm)		Δ residue of formalin
treatments	Before treatment	After treatment	<i>before and after treatment</i> (%)
Washing	1,420.17±66.94 ^{c1}	310.42±77.61 ^{b2}	78.14
Frying	1,420.17±66.94 ^{c1}	14.18±6.69 ^{a2}	99.01
Boiling	1,420.17±66.94 ^{c1}	27.50±9.11 ^{a2}	98.10

Mean \pm SD in the same column with different alphabetical superscripts (a, b, or c) are significantly different (p<0.05) by using one-way ANOVA statistical analysis. Mean \pm SD in the same row with different number superscripts (1 or 2) are significantly different (p<0.05) by using paired sample T-test statistical analysis.

Effect of storage temperatures and periods on the formalin residue of C. gariepinus. Table 2 shows the effect of storage temperatures, of periods and of their combination on the concentration of formalin. Based on this data, the storage period influenced the residue of formalin decrease (p<0.05) during the first 24 hours, but after a storage period of 48 hours the decrease was not significantly different (p>0.05).

On the other hand, there were no differences in the residue of formalin stored at room and freezing temperature, suggesting that the temperature of storage did not affect the formalin residue (p>0.05). Although there were differences in the statistical results between the temperature effects and the storage periods effects, the combined effect of both variables showed no impact on the concentration of formalin (p>0.05).

Table 2

Effect of storage temperature and period on the formalin residue of Clarias gariepinus

Temperature	Periods (t) (hours)				p-value	9
<i>(T)</i>	0	24	48	Т	t	T*t
Room (29°C)	1,420.17±66.94 ^{a1} 1,420.17±66.94 ^{a1}	52.18±28.78 ^{a2}	40.64±3.93 ^{a2}	0.20	0.00	0.76
Freezing (-3°C)	1,420.17±66.94 ^{a1}	39.04±7.07 ^{a2}	13.65±4.23 ^{a2}	0.20	0.00	0.76

Mean±SD in the same column with different alphabetical superscripts (a, b, or c) are significantly different (p<0.05) and so are in the same row with different number superscript (1, 2, or 3). The p-value for T, t, and T*t was obtained from independent T-test, one-way ANOVA, and two-way ANOVA, respectively.

Discussion

The residue of formalin before treatment. The residue of formalin before and after treatment (either fish processing or storage) was significantly different (Table 1 and Table 2). In this study, the *C. gariepinus* specimens were submerged for 60 minutes in formalin solution of 1,480 ppm. The 60 minutes submersion period was chosen because the aldehyde compound may be readily bonded to the amino acids of fish in a small amount (Rahmadhani et al 2017; Baroroh & Kresnadipayana 2016). Formalin would attack lysine for the first time, followed by histidine and tyrosine, leading to the formation of a methylene compound (Ichya'uddin 2016). The protein-containing methylene compounds could not be digested by living things and microbes, especially pathogens, so that the shelf life would be longer (Purawisastra & Sahara 2011). The beneficial bacteria in the fish may also be killed by dehydration mechanism, as another effect of the existence of formalin in the body (Mudzkirah 2016).

The effect of processing on the formalin residue of C. gariepinus. The residue of formalin was reduced by 78.14% during washing treatment. The reduction was significant (p<0.05). The formalin was easily washed out because of its high solubility in the water, which is 4 x 10⁵ mg L⁻¹ at 20°C (BPOM 2008; Sugiarti et al 2014). A study conducted by Yusuf et al (2015) showed the same trend, but the reduction was lower (with only 43%). The differences in the level of reduction can be affected by the duration

AACL Bioflux, 2020, Volume 13, Issue X. http://www.bioflux.com.ro/aacl

1092

Table 1

of formalin submersion. The longer the submersion time, the smaller the decrease, as more formalin was bonded with the fish protein.

In this study, frying also significantly reduced the residue of formalin (p<0.05). The reduction level was 99% that indicated most of the formalin disappeared during the frying process. Formalin was included in a volatile compound, especially above its boiling point (Joshi et al 2015). This study confirmed the results of Sugiarti et al (2014), showing a reduction level of formalin up to 99.28% after 1 hour of submersion followed by the frying process (no information about the period of frying process) in squid. Frying could denature the protein and also hydrolyze its bonding with formalin (Levita et al 2010). The high temperature in frying increased the kinetic energy and led to faster movement of the molecules that could damage the molecular bonds (Sugiarti et al 2014).

Heat processing by boiling reduced the residue of formalin with up to 98.1%, more than the washing process (p<0.05), but at the same degree as the frying process (p>0.05). The high reduction of formalin in the boiling process was due to the accelerated formalin dissolution rate at higher temperatures (Annisak 2019). This effect could be enhanced by boiling the sample in an open pan (Kamal et al 2017).

The effect of storage on the formalin residue of *C. gariepinus*. This experiment demonstrated that only the period of storage significantly influenced the residue of formalin (p<0.05), while the temperature of storage and the combination between temperature and period of storage did not (Table 2). The previous different studies showed that a storage at the room temperature or at the freezing temperature had similar effects on the formalin residue decrease (Jawahar et al 2017; Murtini et al 2014), possibly due to the same level of protein degradation by breaking the bonds between protein and formalin and releasing free compounds. At the room temperature, the protein degradation was essentially caused by the microbial activity, while at the freezing temperature it was caused mostly by physical destruction, leading to conformational and functional changes (Murtini et al 2014; Yeasmin et al 2010). Formalin in the free form is more reactive and can also be more easily degraded (Riyanto et al 2006; Rachmawati et al 2007).

Conclusions. This research concluded that short time processing and storage treatments decreased the concentration of formalin. Heat involvement determined a more effective decrease of concentration (the decrease level was >90%). In the storage study, the residue of formalin was significantly reduced by the period of storage, mainly during the first 24 hours, then the formalin concentration reduction was insignificant, up to 48 hours of storage.

Acknowledgements. The authors are grateful to the Indonesian Ministry of Research, Technology, and Higher Education for providing the financial support in pursuing this research project (Program Kreativitas Mahasiswa Kemeristekdikti 2019 Program).

Reference

- Annisak S. K., Indriyanti N. Y., Mulyani B., 2019 [Constructive controversy and incubation presented according to the representation of tetrahedral chemical learning according to critical thinking ability]. Jurnal Inovasi Pendidikan IPA 5(1):10–22. [In Indonesian].
- Antoni S., 2010 [Analysis of formaldehyde content on salty fish by spectrophotometry method in Tampan District Pekanbaru]. Bachelor Thesis, Universitas Islam Negeri Sultan Syarif Kasim Riau, Indonesia, 53 p. [In Indonesian].
 Baroroh R. D., Kresnadipayana D., 2016 [The effect of variety consentration of starfruit
- Baroroh R. D., Kresnadipayana D., 2016 [The effect of variety consentration of starfruit juice (*Averrhoa bilimbi* L.) on soaked tofu for formaldehyde reduction]. Bachelor Thesis, Universitas Setia Budi, Indonesia, 1-2 p. [In Indonesian].
- Fellows P. J., 2000 [Food processing technology: principle and practice]. Woodhead Publishing Limited and CRC Press LLC, Boca Roton USA, 356 p. ISBN: 0 8493 0887

1093

AACL Bioflux, 2020, Volume 13, Issue X. http://www.bioflux.com.ro/aacl

- Handayani A., Alimin, Rustiah W. O., 2010 [The effect of low temperature storage (freezer -3oc) on water and fat content in lemuru fish (*Sardinella longiceps*)]. Al-Kimia (3):64–75. [In Indonesian].
- Hayun H., Harmita K., Pramudita T.B., 2017. Determination of Formaldehyde Content in Wet Noodles by Thin Layer Chromatography-Densitometry After Derivatization With Nash Reagent. Oriental Journal of Chemistry 33(3):1400-1405.
- Ichya'uddin M., 2014 [Analysis of Formaldehyde content and organoleptic test on salty fish in several traditional markets in Tuban]. Bachelor Thesis, Universitas Islam Negeri Maulana Malik Ibrahim, Indonesia, 112 p. [In Indonesian].
- Jawahar L. S., John C., Shafeekh M., Anupama T. K., Sankar T. V., 2017 Retention of residual formaldehyde in treated Indian mackerel (*Rastrelliger kanagurta*) under iced storage and related food safety concern. Indian Journal of Fisheries 64(4):87– 93.
- Joshi R., Bhatta R., Paudel P. N., Kafle B. K., 2015 Formaldehyde content of selected fish from the wet markets of Kathmandu valley. International Food Research Journal 22(4):1434-1437.
- Kamal A. S., Uddin M. M., Rahman R., Samira S., Rahman M., Nandy S., Khan M. S., 2017 Time and temperature effect on the residual concentration of formaldehyde in formalin treated samples of *Labeo rohita*. International Conference on Food Security and Nutrition, 6 p.
- Kaneko M., Wada Y., Fukui S., Kanno S., 1977 NII-Electronic library service. The Journal of Hygienic Chemistry 23(6):393–396.
- Laksmiani N., Widjaja I., Sonia, 2015 [Stability of formalin in different temperature and period of heating]. Jurnal Farmasi Udayana 4(2):76-81.
- Levita J., Musfiroh I., Indriyati W., Mustarichie R., 2010 The effect of soaking, washing and frying on the concentration of formaldehyde in Sange Belah salty fish. Jurnal Ilmu-ilmu Hayati dan Fisik 12(1):31–34.
- Male Y. T., Letsoin L. I., Siahaya N. A., 2017 [Analysis of formaldehyde content on noodle in several area in Ambon]. Jurnal Kementerian Perindustrian 13(2):5–10. [In Indonesian].
- Mudaffar R. A., 2018 [Qualitative and quantitative test of formaldehyde on apple, grape, and litchi for sale in Makassar]. Jurnal Perbal 6(3):59-65. [In Indonesian].
- Mudzkirah I., 2016 [Identification of borax and formalin on snack food in UIN Alaudin Makassar Canteen]. Bachelor Thesis, UIN Alauddin Makassar, Indonesia, 95 p. [In Indonesian].
- Muntaha A., Haitami, Hayati N., 2015 [Comparison of reduction formaldehyde level on boiled and hot water soaked tofu]. Medical Laboratory Technology Journal 1(2):84– 90. [In Indonesian].
- Murtini J. T., Riyanto R., Priyanto N., Hermana I., 2014 [Development of Formaldehyde on some kinds of seawater fish during storage in crushed ice]. JPB Perikanan 9(2):143–151. [In Indonesian].
- Pandey C. K., Agarwal A., Baronia A., Singh N., 2000 Toxicity of ingested formalin and its management. Human and Experiment Toxicology 360–366.
- Purawisastra S., Sahara E., 2011 [Absorption to formalin by certain foodstuff and disappearance through hot water soaked]. The Journal of Nutrition and Food Research 34(1):63–74. [In Indonesian].
- Rachmawati N., Riyanto R., Ariyani F., 2007 [Identification of formaldehyde on kerapu macan fish (*Ephinephelus fuscoguttatus*) during cold storage]. Jurnal Pascapanen dan Bioteknologi Kelautan dan Perikanan 2(2):137–145. [In Indonesian].
- Rahmadhani F., Safrida, Djufri, 2017 [The effect of soaking a various of solution for reduction formalin level on kembung salty fish (*Scomber canagorta*) in Lambaro traditional market Aceh]. Jurnal Ilmiah Mahasiswa Pendidikan Biologi 2(3):1-12. [In Indonesian].
- Riyanto R., Kusmarwati A., Dwiyitno, 2006 [Identification of formaldehyde foorming on kerapu fish (*Epinephelus fuscoguttatus*) during room temperature storage]. Jurnal Pascapanen dan Bioteknologi Kelautan dan Perikanan 1(2):111-116. [In Indonesian].

AACL Bioflux, 2020, Volume 13, Issue X. http://www.bioflux.com.ro/aacl

Shita A. E., 2016 [Selectivity of spectrophotometer analysis with Schiff's reagent]. Bachelor Thesis, Universitas Negeri Yogyakarta, Indonesia, 87 p. [In Indonesian].

Sugiarti M., Anggo A. D., Riyadi P. H., 2014 [The effect of soaking in undercooking temperature and cooking method for reduction formalin level on cuttlefish]. Jurnal Pengolahan dan Bioteknologi Hasil Perikanan 3(2):90–98. [In Indonesian].

Sundari D., Almasyhuri, Lamid A., 2015 [The effect of cooking process for nutrition composition of protein source]. Media Litbangkes 25(4):235–242. [In Indonesian]. Suwanaruang T., 2018 Formalin contaminated in seafood and frozen meat at Somdet

market, Kalasin Province. Journal of Environmental Protection 9:1286–1293. Wijayanti N. S., Lukitasari M., 2016 [Analysis of formalin level and organoleptic test in

salty fish in Madiun Market]. Jurnal Florea 3(1):59–64. [In Indonesian].

Wilianto W., Yudianto A., 2013 [The effect of formalin exposure on various level for DNA psoas major by PCR Lokus CSF1PO, D5S818, D13S317, D21S11 Analysis]. Jurnal Kedokteran Forensik Indonesia 15(1):11–16. [In Indonesian].

Yeasmin T., Reza M. S., Shikha F. H., Khan M. N. A., Kamal M., 2010 Quality changes in formalin treated rohu fish (*Labeo rohita*, Hamilton) during ice storage condition. Asian Journal of Agricultural Science 2(4):158–163.

Yulisa N., Asni E., Azrin M., 2014 [Formaldehyde test of gurame fish in Pekanbaru Traditional Market]. Jurnal Online Mahasiswa 1(2):1-12. [In Indonesian].

Yusuf Y., Zuki Z., Amanda R. R., 2015 [The effect of various treatment for reduction formalin level on fish by spectrophotometry method]. Jurnal Riset Kimia 8(2):182– 188. [In Indonesian].

*** BPOM, Indonesian Food and Drug Authority, 2008 [Formaldehyde]. Badan POM RI, Jakarta, Indonesia. [In Indonesian].

Commented [S2]: I replace Harmita 2016 with Hayun et al., 2017

Commented [S3]: I replaced Fellows 2015 with Fellows 2000 Commented [S4]:

Commented [WU5]: Please specify what kind ogf thesis: PhD? MSc?

Received: 15 October 2020. Accepted: 2020. Published online: 2020. Authors:

Isny Maulidevi, Alma Ata University, Faculty of Health Sciences, Department of Nutrition, Jl. Brawijaya 99, 55183 Yogyakarta, Indonesia, e-mail: isnymaulidevi540@gmail.com Nurul Andriyani, Alma Ata University, Faculty of Health Sciences, Department of Nutrition, Jl. Brawijaya 99,

Nurul Andriyani, Alma Ata University, Faculty of Health Sciences, Department of Nutrition, Jl. Brawijaya 99, 55183 Yogyakarta, Indonesia, e-mail: nurulandriyani96@gmail.com Ryan Salfarino, Alma Ata University, Faculty of Health Sciences, Department of Nutrition, Jl. Brawijaya 99,

Sila Yogyakarta, Indonesia, e-mail: ryan.salfarino@gmail.com Veriani Aprilia, Alma Ata University, Faculty of Health Sciences, Department of Nutrition, Jl. Brawijaya 99, 55183 Yogyakarta, Indonesia, e-mail: verianiaprilia@almaata.ac.id

55183 Yogyakarta, Indonesia, e-mail: verianiaprilia@almaata.ac.id This is an open-access article distributed under the terms of the Creative Commons Attribution License, which

permits unrestricted use, distribution and reproduction in any medium, provided the original author and source are credited.

AACL Bioflux, 2020, Volume 13, Issue X. http://www.bioflux.com.ro/aacl

How to cite this article: Maulidevi I., Andriyani N., Salfarino R., Aprilia V., 2020 The residue of formalin in catfish (*Clarias gariepinus*) after processing and storage for short periods. AACL Bioflux 13(x):

AACL Bioflux, 2020, Volume 13, Issue X. http://www.bioflux.com.ro/aacl



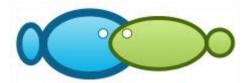
Penetapan revisi 2: 15 Desember 2020

Eni Kovacs <ek.bioflux@gmail.com> Kepada: verianiaprilia verianiaprilia <verianiaprilia@almaata.ac.id> 15 Desember 2020 pukul 02.35

Thank you for your email. Please address the issue from the reference list with track change.

[Kutipan teks disembunyikan]

Aprilia_Clarias gariepinus_final.doc



The residue of formalin in catfish (*Clarias* gariepinus) after processing and storage for short periods

İsny Maulidevi, Nurul Andriyani, Ryan Salfarino, Veriani Aprilia

Department of Nutrition, Faculty of Health Sciences, Alma Ata University, Yogyakarta, Indonesia. Corresponding author: V. Aprilia, verianiaprilia@almaata.ac.id

Abstract. Fresh fish is a highly perishable product due to its high protein content. A preservative may be needed to inhibit it. Nowadays, many fishes were reported to be exposed to formalin. This study aimed to determine the effect of short period processing (washing, frying, boiling in short-period) and storage (at the room and at freezing temperatures) on the residue of formalin in catfish, as a food model. Catfish, *Clarias gariepinus*, were divided into 8 groups, e.g. before treatment, washing in airflow for 1 minute, frying in 200°C for 8 minutes, boiling in 100°C for 7 minutes, storage at room temperature (29±1°C) for 24 and 48 hours, storage at freezing temperature (-3°C) for 24 and 48 hours. Before treatment, the catfish was soaked in 1,480 ppm of formalin solutions. The residue of formalin was measured at the end of each treatment by using a spectrophotometer. Data were then analyzed with analysis of variance with SPSS software. The results showed that there were significant effects of processing and storage treatment on the formalin residue of catfish. The formalin residue of all treatments was lower than before treatments. The frying processing and storage at freezing temperatures were the most effective treatments in reducing the formalin residue. **Key Words**: fresh fish, preservative, food model, treatments, formalin residue.

Introduction. Fresh fish is a highly perishable product due to its high protein content for microbial activity (Wijayanti & Lukitasari 2016). A preservative maybe needs to keep the freshness, inhibit the decay and prolong the shelf life (Antoni 2010). Nowadays, there are many kinds of food-grade preservatives available in the market. Unfortunately, formalin is still frequently found in food products, including fresh fish, being used among fishermen mainly because it is effective in preserving the product, cheaper and accessible (Male et al 2017).

Formalin is a forty percent solution of formaldehyde in water. This compound is detected in the animal body as an intermediary metabolite. It also uses for several purposes, such as in medicine for specimen preservation and the treatment of hemorrhage (Pandey et al 2000). The preservatives activity of formalin is due to its ability to react with protein generating methylene compounds, which are more difficult to be decayed by microbes (Ichya'uddin 2016). In low concentrations, formalin is not toxic for humans, but in higher concentrations, it had negative effects such as acute toxicity and death (Pandey et al 2000). It also decreases the levels of body antioxidants, resulting in liver damage (Yulisa et al 2014). Other studies also reported the cases of eczema, eye irritation, respiratory tract irritation, cancer, increased growth rate of malignant neoplastic diseases and denaturation of DNA (Suwanaruang 2018; Wilianto & Yudianto 2013). Therefore, its uses are forbidden.

Formalin solution is not stable during storage and exposure to high temperatures (Kaneko et al 1977; Laksmiani et al 2015). Those are caused by desolvation or evaporation of formalin during washing, boiling, or frying (Fellows 2000; Hayun et al 2017; Muntaha et al 2015; Sundari et al 2015) and enzymatic process in storage (Rachmawati et al 2007). Formalin that was in contact with fish decreased up to 43-53% after 5 times rinsing in 250 mL of distilled water and boiling for 30 minutes (Yusuf et al 2015). Other studies were also reported that soaking in warm water and frying decreased

the concentration of formalin up to 63.27% and 83.03%, respectively (Levita et al 2010). In daily practices, fish is usually washed in flowing water before processing, then fryed/boiled for a short time. However, there was no study explaining the influences of these processing and storage methods on the concentration of formalin.

The purpose of this study was to identify the effect of processing for a short time and storage (at room and freezing temperatures) on the residue of formalin. This study used catfish, *Clarias gariepinus*, as a food model.

Material and Method

Material. The study used fresh catfish, purchased from the traditional market in Yogyakarta, Indonesia. The other used ingredients were formalin solution and Schiff's reagent. Formalin solution was made from 4 mL of formalin 37% in 1,000 mL of distilled water (Rachmawati et al 2007).

Method. The study was divided into two sections, i.e. 1) analyzing the effect of processing and 2) analyzing the effect of storage treatment. Fresh catfish were slaughtered, eviscerated, cleaned, and soaked in 1,480 ppm formalin solution (Rachmawati et al 2007; Sugiarti et al 2014). For the first study, catfish were divided into four groups, i.e.: controls (before treatment), washing, frying, and boiling treatments. Washing was done by watering the samples with running water for 1 minute, while frying was done by frying the samples in oil at 200°C for 8 minutes. Boiling treatment was done by putting the samples into boiled water (100°C) for 7 minutes (Sugiarti et al 2014). For the second study, storage treatments, the catfish was also divided into four groups: room temperature storage (29°C) for 24 hours and 48 hours and freezing temperatures storage (-3°C) for 24 hours and 48 hours (Handayani et al 2010). At the end of treatments, the concentration of formalin in samples was analyzed using a spectrophotometer. As the preparation step, a 5 g sample was dissolved in 25 mL of distilled water. One mL of supernatant was mixed with 2 mL of Schiff's reagent, then heated in a water bath at 60°C for 5 minutes. It was incubated for 10 minutes before measuring the absorbance at 520 nm (Shita 2016; Mudaffar 2018).

Statistical analysis. Data were presented in mean±SD and analyzed with: (1) one-way analysis of variance (ANOVA) to determine the differences among all processing treatments and the effect of storage periods on the concentration of formalin; (2) paired sample T-test to determine the differences between the same samples before and after processing treatments; (3) independent sample t-test to know the effect of temperature at the same storage period, and (4) two-way ANOVA to determine the combination effect of storage temperature and period on the concentration of formalin. All data were analyzed with SPSS software (version 16.0 for Windows; SPSS Inc., Chicago, IL, USA) at the 5% significance level.

Results

Effect of processing on the formalin residue of C. gariepinus. Table 1 shows that all treatments had significant effects on the residue of formalin (p<0.05). It can be seen from the p-value between the same samples before and after treatments. Among all treatments, washing shows the highest residue of formalin (p<0.05). Both boiling and frying had an almost similar residue of formalin (p>0.05). The lower residue of formalin in boiling and frying may be due to heating exposure.

Table 1

Processing	Residue of fo	rmalin (ppm)	Δ residue of formalin
treatments	Before treatment	After treatment	<i>before and after treatment</i> (%)
Washing	1,420.17±66.94 ^{c1}	310.42±77.61 ^{b2}	78.14
Frying	1,420.17±66.94 ^{c1}	14.18 ± 6.69^{a2}	99.01
Boiling	1,420.17±66.94 ^{c1}	27.50±9.11 ^{a2}	98.10

Mean \pm SD in the same column with different alphabetical superscripts (a, b, or c) are significantly different (p<0.05) by using one-way ANOVA statistical analysis. Mean \pm SD in the same row with different number superscripts (1 or 2) are significantly different (p<0.05) by using paired sample T-test statistical analysis.

Effect of storage temperatures and periods on the formalin residue of C. gariepinus. Table 2 shows the effect of storage temperatures, of periods and of their combination on the concentration of formalin. Based on this data, the storage period influenced the residue of formalin decrease (p<0.05) during the first 24 hours, but after a storage period of 48 hours the decrease was not significantly different (p>0.05).

On the other hand, there were no differences in the residue of formalin stored at room and freezing temperature, suggesting that the temperature of storage did not affect the formalin residue (p>0.05). Although there were differences in the statistical results between the temperature effects and the storage periods effects, the combined effect of both variables showed no impact on the concentration of formalin (p>0.05).

Table 2

Effect of storage temperature and period on the formalin residue of *Clarias gariepinus*

Temperature	Periods (t) (hours)				p-value	9
<i>(T)</i>	0	24	48	Т	t	T*t
Room (29°C)	1,420.17±66.94 ^{a1}	52.18±28.78 ^{a2}	40.64±3.93 ^{a2}	0.20	0.00	0.76
Freezing (-3°C)	1,420.17±66.94 ^{a1}		13.65±4.23 ^{a2}	0.20	0.00	0.76

Mean \pm SD in the same column with different alphabetical superscripts (a, b, or c) are significantly different (p<0.05) and so are in the same row with different number superscript (1, 2, or 3). The p-value for T, t, and T*t was obtained from independent T-test, one-way ANOVA, and two-way ANOVA, respectively.

Discussion

The residue of formalin before treatment. The residue of formalin before and after treatment (either fish processing or storage) was significantly different (Table 1 and Table 2). In this study, the *C. gariepinus* specimens were submerged for 60 minutes in formalin solution of 1,480 ppm. The 60 minutes submersion period was chosen because the aldehyde compound may be readily bonded to the amino acids of fish in a small amount (Rahmadhani et al 2017; Baroroh & Kresnadipayana 2016). Formalin would attack lysine for the first time, followed by histidine and tyrosine, leading to the formation of a methylene compound (Ichya'uddin 2016). The protein-containing methylene compounds could not be digested by living things and microbes, especially pathogens, so that the shelf life would be longer (Purawisastra & Sahara 2011). The beneficial bacteria in the fish may also be killed by dehydration mechanism, as another effect of the existence of formalin in the body (Mudzkirah 2016).

The effect of processing on the formalin residue of C. gariepinus. The residue of formalin was reduced by 78.14% during washing treatment. The reduction was significant (p<0.05). The formalin was easily washed out because of its high solubility in the water, which is 4×10^5 mg L⁻¹ at 20°C (BPOM 2008; Sugiarti et al 2014). A study conducted by Yusuf et al (2015) showed the same trend, but the reduction was lower (with only 43%). The differences in the level of reduction can be affected by the duration

of formalin submersion. The longer the submersion time, the smaller the decrease, as more formalin was bonded with the fish protein.

In this study, frying also significantly reduced the residue of formalin (p<0.05). The reduction level was 99% that indicated most of the formalin disappeared during the frying process. Formalin was included in a volatile compound, especially above its boiling point (Joshi et al 2015). This study confirmed the results of Sugiarti et al (2014), showing a reduction level of formalin up to 99.28% after 1 hour of submersion followed by the frying process (no information about the period of frying process) in squid. Frying could denature the protein and also hydrolyze its bonding with formalin (Levita et al 2010). The high temperature in frying increased the kinetic energy and led to faster movement of the molecules that could damage the molecular bonds (Sugiarti et al 2014).

Heat processing by boiling reduced the residue of formalin with up to 98.1%, more than the washing process (p<0.05), but at the same degree as the frying process (p>0.05). The high reduction of formalin in the boiling process was due to the accelerated formalin dissolution rate at higher temperatures (Annisak 2019). This effect could be enhanced by boiling the sample in an open pan (Kamal et al 2017).

The effect of storage on the formalin residue of C. gariepinus. This experiment demonstrated that only the period of storage significantly influenced the residue of formalin (p<0.05), while the temperature of storage and the combination between temperature and period of storage did not (Table 2). The previous different studies showed that a storage at the room temperature or at the freezing temperature had similar effects on the formalin residue decrease (Jawahar et al 2017; Murtini et al 2014), possibly due to the same level of protein degradation by breaking the bonds between protein and formalin and releasing free compounds. At the room temperature, the protein degradation was essentially caused by the microbial activity, while at the freezing temperature it was caused mostly by physical destruction, leading to conformational and functional changes (Murtini et al 2014; Yeasmin et al 2010). Formalin in the free form is more reactive and can also be more easily degraded (Riyanto et al 2006; Rachmawati et al 2007).

Conclusions. This research concluded that short time processing and storage treatments decreased the concentration of formalin. Heat involvement determined a more effective decrease of concentration (the decrease level was >90%). In the storage study, the residue of formalin was significantly reduced by the period of storage, mainly during the first 24 hours, then the formalin concentration reduction was insignificant, up to 48 hours of storage.

Acknowledgements. The authors are grateful to the Indonesian Ministry of Research, Technology, and Higher Education for providing the financial support in pursuing this research project (Program Kreativitas Mahasiswa Kemeristekdikti 2019 Program).

Reference

- Annisak S. K., Indriyanti N. Y., Mulyani B., 2019 [Constructive controversy and incubation presented according to the representation of tetrahedral chemical learning according to critical thinking ability]. Jurnal Inovasi Pendidikan IPA 5(1):10–22. [In Indonesian].
- Antoni S., 2010 [Analysis of formaldehyde content on salty fish by spectrophotometry method in Tampan District Pekanbaru]. BSc Thesis, Universitas Islam Negeri Sultan Syarif Kasim Riau, Indonesia, 53 p. [In Indonesian].
- Baroroh R. D., Kresnadipayana D., 2016 [The effect of variety consentration of starfruit juice (*Averrhoa bilimbi* L.) on soaked tofu for formaldehyde reduction]. BSc Thesis, Universitas Setia Budi, Indonesia, 1-2 p. [In Indonesian]. <u>2 pages thesis?This is the third time we address this issue. Thank you.</u>
- Fellows P. J., 2000 Food processing technology: principle and practice. Woodhead Publishing Limited and CRC Press LLC, Boca Roton, USA, 356 p.

- Handayani A., Alimin, Rustiah W. O., 2010 [The effect of low temperature storage (freezer -3oc) on water and fat content in lemuru fish (*Sardinella longiceps*)]. Al-Kimia (3):64–75. [In Indonesian].
- Hayun H., Harmita K., Pramudita T. B., 2017 Determination of formaldehyde content in wet noodles by thin layer chromatography-densitometry after derivatization with Nash reagent. Oriental Journal of Chemistry 33(3):1400-1405.
- Ichya'uddin M., 2014 [Analysis of Formaldehyde content and organoleptic test on salty fish in several traditional markets in Tuban]. BSc Thesis, Universitas Islam Negeri Maulana Malik Ibrahim, Indonesia, 112 p. [In Indonesian].
- Jawahar L. S., John C., Shafeekh M., Anupama T. K., Sankar T. V., 2017 Retention of residual formaldehyde in treated Indian mackerel (*Rastrelliger kanagurta*) under iced storage and related food safety concern. Indian Journal of Fisheries 64(4):87–93.
- Joshi R., Bhatta R., Paudel P. N., Kafle B. K., 2015 Formaldehyde content of selected fish from the wet markets of Kathmandu valley. International Food Research Journal 22(4):1434-1437.
- Kamal A. S., Uddin M. M., Rahman R., Samira S., Rahman M., Nandy S., Khan M. S., 2017 Time and temperature effect on the residual concentration of formaldehyde in formalin treated samples of *Labeo rohita*. International Conference on Food Security and Nutrition, 6 p.
- Kaneko M., Wada Y., Fukui S., Kanno S., 1977 NII-Electronic library service. The Journal of Hygienic Chemistry 23(6):393–396.
- Laksmiani N., Widjaja I., Sonia, 2015 [Stability of formalin in different temperature and period of heating]. Jurnal Farmasi Udayana 4(2):76–81.
- Levita J., Musfiroh I., Indriyati W., Mustarichie R., 2010 The effect of soaking, washing and frying on the concentration of formaldehyde in Sange Belah salty fish. Jurnal Ilmu-ilmu Hayati dan Fisik 12(1):31–34.
- Male Y. T., Letsoin L. I., Siahaya N. A., 2017 [Analysis of formaldehyde content on noodle in several area in Ambon]. Jurnal Kementerian Perindustrian 13(2):5–10. [In Indonesian].
- Mudaffar R. A., 2018 [Qualitative and quantitative test of formaldehyde on apple, grape, and litchi for sale in Makassar]. Jurnal Perbal 6(3):59-65. [In Indonesian].
- Mudzkirah I., 2016 [Identification of borax and formalin on snack food in UIN Alaudin Makassar Canteen]. BSc Thesis, UIN Alauddin Makassar, Indonesia, 95 p. [In Indonesian].
- Muntaha A., Haitami, Hayati N., 2015 [Comparison of reduction formaldehyde level on boiled and hot water soaked tofu]. Medical Laboratory Technology Journal 1(2):84– 90. [In Indonesian].
- Murtini J. T., Riyanto R., Priyanto N., Hermana I., 2014 [Development of Formaldehyde on some kinds of seawater fish during storage in crushed ice]. JPB Perikanan 9(2):143–151. [In Indonesian].
- Pandey C. K., Agarwal A., Baronia A., Singh N., 2000 Toxicity of ingested formalin and its management. Human and Experiment Toxicology 360–366.
- Purawisastra S., Sahara E., 2011 [Absorption to formalin by certain foodstuff and disappearance through hot water soaked]. The Journal of Nutrition and Food Research 34(1):63–74. [In Indonesian].
- Rachmawati N., Riyanto R., Ariyani F., 2007 [Identification of formaldehyde on kerapu macan fish (*Ephinephelus fuscoguttatus*) during cold storage]. Jurnal Pascapanen dan Bioteknologi Kelautan dan Perikanan 2(2):137–145. [In Indonesian].
- Rahmadhani F., Safrida, Djufri, 2017 [The effect of soaking a various of solution for reduction formalin level on kembung salty fish (*Scomber canagorta*) in Lambaro traditional market Aceh]. Jurnal Ilmiah Mahasiswa Pendidikan Biologi 2(3):1-12. [In Indonesian].
- Riyanto R., Kusmarwati A., Dwiyitno, 2006 [Identification of formaldehyde foorming on kerapu fish (*Epinephelus fuscoguttatus*) during room temperature storage]. Jurnal Pascapanen dan Bioteknologi Kelautan dan Perikanan 1(2):111–116. [In Indonesian].

Shita A. E., 2016 [Selectivity of spectrophotometer analysis with Schiff's reagent]. BSc Thesis, Universitas Negeri Yogyakarta, Indonesia, 87 p. [In Indonesian].

- Sugiarti M., Anggo A. D., Riyadi P. H., 2014 [The effect of soaking in undercooking temperature and cooking method for reduction formalin level on cuttlefish]. Jurnal Pengolahan dan Bioteknologi Hasil Perikanan 3(2):90–98. [In Indonesian].
- Sundari D., Almasyhuri, Lamid A., 2015 [The effect of cooking process for nutrition composition of protein source]. Media Litbangkes 25(4):235–242. [In Indonesian].
- Suwanaruang T., 2018 Formalin contaminated in seafood and frozen meat at Somdet market, Kalasin Province. Journal of Environmental Protection 9:1286–1293.
- Wijayanti N. S., Lukitasari M., 2016 [Analysis of formalin level and organoleptic test in salty fish in Madiun Market]. Jurnal Florea 3(1):59–64. [In Indonesian].
- Wilianto W., Yudianto A., 2013 [The effect of formalin exposure on various level for DNA psoas major by PCR Lokus CSF1PO, D5S818, D13S317, D21S11 Analysis]. Jurnal Kedokteran Forensik Indonesia 15(1):11–16. [In Indonesian].
- Yeasmin T., Reza M. S., Shikha F. H., Khan M. N. A., Kamal M., 2010 Quality changes in formalin treated rohu fish (*Labeo rohita*, Hamilton) during ice storage condition. Asian Journal of Agricultural Science 2(4):158–163.
- Yulisa N., Asni E., Azrin M., 2014 [Formaldehyde test of gurame fish in Pekanbaru Traditional Market]. Jurnal Online Mahasiswa 1(2):1-12. [In Indonesian].
- Yusuf Y., Zuki Z., Amanda R. R., 2015 [The effect of various treatment for reduction formalin level on fish by spectrophotometry method]. Jurnal Riset Kimia 8(2):182–188. [In Indonesian].
- *** BPOM, Indonesian Food and Drug Authority, 2008 [Formaldehyde]. Badan POM RI, Jakarta, Indonesia. [In Indonesian].

Received: 15 October 2020. Accepted: 2020. Published online: 2020. Authors:

Isny Maulidevi, Alma Ata University, Faculty of Health Sciences, Department of Nutrition, Jl. Brawijaya 99, 55183 Yogyakarta, Indonesia, e-mail: isnymaulidevi540@gmail.com

Nurul Andriyani, Alma Ata University, Faculty of Health Sciences, Department of Nutrition, Jl. Brawijaya 99, 55183 Yogyakarta, Indonesia, e-mail: nurulandriyani96@gmail.com

How to cite this article:

Ryan Salfarino, Alma Ata University, Faculty of Health Sciences, Department of Nutrition, Jl. Brawijaya 99, 55183 Yogyakarta, Indonesia, e-mail: ryan.salfarino@gmail.com

Veriani Aprilia, Alma Ata University, Faculty of Health Sciences, Department of Nutrition, Jl. Brawijaya 99, 55183 Yogyakarta, Indonesia, e-mail: verianiaprilia@almaata.ac.id

This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution and reproduction in any medium, provided the original author and source are credited.

Maulidevi I., Andriyani N., Salfarino R., Aprilia V., 2020 The residue of formalin in catfish (*Clarias gariepinus*) after processing and storage for short periods. AACL Bioflux 13(x):

verianiaprilia verianiaprilia <verianiaprilia@almaata.ac.id>



Pengiriman Hasil Penetapan revisi 2: 15 Desember 2020

Kepada: Eni Kovacs <ek.bioflux@gmail.com>

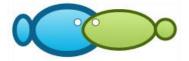
Dear Kovacs,

Thank you for your politeness. We decided to erase the reference from the list for a reason that we have written in the document.

Regards,

Veriani Aprilia [Kutipan teks disembunyikan]

Aprilia_Clarias gariepinus_final 5.doc 172K



The residue of formalin in catfish (*Clarias* gariepinus) after processing and storage for short periods

İsny Maulidevi, Nurul Andriyani, Ryan Salfarino, Veriani Aprilia

Department of Nutrition, Faculty of Health Sciences, Alma Ata University, Yogyakarta, Indonesia. Corresponding author: V. Aprilia, verianiaprilia@almaata.ac.id

Abstract. Fresh fish is a highly perishable product due to its high protein content. A preservative may be needed to inhibit it. Nowadays, many fishes were reported to be exposed to formalin. This study aimed to determine the effect of short period processing (washing, frying, boiling in short-period) and storage (at the room and at freezing temperatures) on the residue of formalin in catfish, as a food model. Catfish, *Clarias gariepinus*, were divided into 8 groups, e.g. before treatment, washing in airflow for 1 minute, frying in 200°C for 8 minutes, boiling in 100°C for 7 minutes, storage at room temperature ($29\pm1°C$) for 24 and 48 hours, storage at freezing temperature (-3°C) for 24 and 48 hours. Before treatment, the catfish was soaked in 1,480 ppm of formalin solutions. The residue of formalin was measured at the end of each treatment by using a spectrophotometer. Data were then analyzed with analysis of variance with SPS software. The results showed that there were significant effects of all treatments was lower than before treatments. The frying processing and storage at freezing temperatures of catfish. The formalin residue of all treatments was lower than before treatments in reducing the formalin residue. **Key Words**: fresh fish, preservative, food model, treatments, formalin residue.

Introduction. Fresh fish is a highly perishable product due to its high protein content for microbial activity (Wijayanti & Lukitasari 2016). A preservative maybe needs to keep the freshness, inhibit the decay and prolong the shelf life (Antoni 2010). Nowadays, there are many kinds of food-grade preservatives available in the market. Unfortunately, formalin is still frequently found in food products, including fresh fish, being used among fishermen mainly because it is effective in preserving the product, cheaper and accessible (Male et al 2017).

Formalin is a forty percent solution of formaldehyde in water. This compound is detected in the animal body as an intermediary metabolite. It also uses for several purposes, such as in medicine for specimen preservation and the treatment of hemorrhage (Pandey et al 2000). The preservatives activity of formalin is due to its ability to react with protein generating methylene compounds, which are more difficult to be decayed by microbes (Ichya'uddin 2016). In low concentrations, formalin is not toxic for humans, but in higher concentrations, it had negative effects such as acute toxicity and death (Pandey et al 2000). It also decreases the levels of body antioxidants, resulting in liver damage (Yulisa et al 2014). Other studies also reported the cases of eczema, eye irritation, respiratory tract irritation, cancer, increased growth rate of malignant neoplastic diseases and denaturation of DNA (Suwanaruang 2018; Wilianto & Yudianto 2013). Therefore, its uses are forbidden.

Formalin solution is not stable during storage and exposure to high temperatures (Kaneko et al 1977; Laksmiani et al 2015). Those are caused by desolvation or evaporation of formalin during washing, boiling, or frying (Fellows 2000; Hayun et al 2017; Muntaha et al 2015; Sundari et al 2015) and enzymatic process in storage (Rachmawati et al 2007). Formalin that was in contact with fish decreased up to 43-53% after 5 times rinsing in 250 mL of distilled water and boiling for 30 minutes (Yusuf et al 2015). Other studies were also reported that soaking in warm water and frying decreased

AACL Bioflux, 2020, Volume 13, Issue X. http://www.bioflux.com.ro/aacl

the concentration of formalin up to 63.27% and 83.03%, respectively (Levita et al 2010). In daily practices, fish is usually washed in flowing water before processing, then fryed/boiled for a short time. However, there was no study explaining the influences of these processing and storage methods on the concentration of formalin.

The purpose of this study was to identify the effect of processing for a short time and storage (at room and freezing temperatures) on the residue of formalin. This study used catfish, *Clarias gariepinus*, as a food model.

Material and Method

Material. The study used fresh catfish, purchased from the traditional market in Yogyakarta, Indonesia. The other used ingredients were formalin solution and Schiff's reagent. Formalin solution was made from 4 mL of formalin 37% in 1,000 mL of distilled water (Rachmawati et al 2007).

Method. The study was divided into two sections, i.e. 1) analyzing the effect of processing and 2) analyzing the effect of storage treatment. Fresh catfish were slaughtered, eviscerated, cleaned, and soaked in 1,480 ppm formalin solution (Rachmawati et al 2007; Sugiarti et al 2014). For the first study, catfish were divided into four groups, i.e.: controls (before treatment), washing, frying, and boiling treatments. Washing was done by watering the samples with running water for 1 minute, while frying was done by frying the samples in oil at 200°C for 8 minutes. Boiling treatment was done by putting the samples into boiled water (100°C) for 7 minutes (Sugiarti et al 2014). For the second study, storage treatments, the catfish was also divided into four groups: room temperature storage (29°C) for 24 hours and 48 hours and freezing temperatures storage (-3°C) for 24 hours and 48 hours (Handayani et al 2010). At the end of treatments, the concentration of formalin in samples was analyzed using a spectrophotometer. As the preparation step, a 5 g sample was dissolved in 25 mL of distilled water. One mL of supernatant was mixed with 2 mL of Schiff's reagent, then heated in a water bath at 60°C for 5 minutes. It was incubated for 10 minutes before measuring the absorbance at 520 nm (Shita 2016; Mudaffar 2018).

Statistical analysis. Data were presented in mean±SD and analyzed with: (1) one-way analysis of variance (ANOVA) to determine the differences among all processing treatments and the effect of storage periods on the concentration of formalin; (2) paired sample T-test to determine the differences between the same samples before and after processing treatments; (3) independent sample t-test to know the effect of temperature at the same storage period, and (4) two-way ANOVA to determine the combination effect of storage temperature and period on the concentration of formalin. All data were analyzed with SPSS software (version 16.0 for Windows; SPSS Inc., Chicago, IL, USA) at the 5% significance level.

Results

Effect of processing on the formalin residue of C. gariepinus. Table 1 shows that all treatments had significant effects on the residue of formalin (p<0.05). It can be seen from the p-value between the same samples before and after treatments. Among all treatments, washing shows the highest residue of formalin (p<0.05). Both boiling and frying had an almost similar residue of formalin (p>0.05). The lower residue of formalin in boiling and frying may be due to heating exposure.

Effect of processing on the formalin residue of Clarias gariepinus

Processing	Residue of formalin (ppm)		Δ residue of formalin		
treatments	Before treatment	After treatment	<i>before and after treatment</i> (%)		
Washing	1,420.17±66.94 ^{c1}	310.42±77.61 ^{b2}	78.14		
Frying	1,420.17±66.94 ^{c1}	14.18 ± 6.69^{a2}	99.01		
Boiling	1,420.17±66.94 ^{c1}	27.50 ± 9.11^{a2}	98.10		

Mean \pm SD in the same column with different alphabetical superscripts (a, b, or c) are significantly different (p<0.05) by using one-way ANOVA statistical analysis. Mean \pm SD in the same row with different number superscripts (1 or 2) are significantly different (p<0.05) by using paired sample T-test statistical analysis.

Effect of storage temperatures and periods on the formalin residue of C. gariepinus. Table 2 shows the effect of storage temperatures, of periods and of their combination on the concentration of formalin. Based on this data, the storage period influenced the residue of formalin decrease (p<0.05) during the first 24 hours, but after a storage period of 48 hours the decrease was not significantly different (p>0.05).

On the other hand, there were no differences in the residue of formalin stored at room and freezing temperature, suggesting that the temperature of storage did not affect the formalin residue (p>0.05). Although there were differences in the statistical results between the temperature effects and the storage periods effects, the combined effect of both variables showed no impact on the concentration of formalin (p>0.05).

Table 2

Effect of storage temperature and period on the formalin residue of *Clarias gariepinus*

Temperature	Per	riods (t) (hours)			p-value	9
(T)	0	24	48	Т	t	T*t
Room (29°C)	1,420.17±66.94 ^{a1}	52.18±28.78 ^{a2}	40.64±3.93 ^{a2}	0.20	0.00	0.76
Freezing (-3°C)	1,420.17±66.94 ^{a1}	39.04±7.07 ^{a2}	13.65±4.23 ^{a2}	0.20	0.00	0.76

Mean±SD in the same column with different alphabetical superscripts (a, b, or c) are significantly different (p<0.05) and so are in the same row with different number superscript (1, 2, or 3). The p-value for T, t, and T*t was obtained from independent T-test, one-way ANOVA, and two-way ANOVA, respectively.

Discussion

The residue of formalin before treatment. The residue of formalin before and after treatment (either fish processing or storage) was significantly different (Table 1 and Table 2). In this study, the *C. gariepinus* specimens were submerged for 60 minutes in formalin solution of 1,480 ppm. The 60 minutes submersion period was chosen because the aldehyde compound may be readily bonded to the amino acids of fish in a small amount (Rahmadhani et al 2017; Baroroh & Kresnadipayana 2016). Formalin would attack lysine for the first time, followed by histidine and tyrosine, leading to the formation of a methylene compound (Ichya'uddin 2016). The protein-containing methylene compounds could not be digested by living things and microbes, especially pathogens, so that the shelf life would be longer (Purawisastra & Sahara 2011). The beneficial bacteria in the fish may also be killed by dehydration mechanism, as another effect of the existence of formalin in the body (Mudzkirah 2016).

The effect of processing on the formalin residue of C. gariepinus. The residue of formalin was reduced by 78.14% during washing treatment. The reduction was significant (p<0.05). The formalin was easily washed out because of its high solubility in the water, which is 4 x 10⁵ mg L⁻¹ at 20°C (BPOM 2008; Sugiarti et al 2014). A study conducted by Yusuf et al (2015) showed the same trend, but the reduction was lower (with only 43%). The differences in the level of reduction can be affected by the duration

AACL Bioflux, 2020, Volume 13, Issue X. http://www.bioflux.com.ro/aacl 1092

Commented [S1]: We have only the resume of this research. We have tried to contacted the author, but still not succeed. Would I erase this reference? Our sentences here is still relevant if we cite from only 1 reference (Rahmadhani et al 2017).

Table 1

of formalin submersion. The longer the submersion time, the smaller the decrease, as more formalin was bonded with the fish protein.

In this study, frying also significantly reduced the residue of formalin (p<0.05). The reduction level was 99% that indicated most of the formalin disappeared during the frying process. Formalin was included in a volatile compound, especially above its boiling point (Joshi et al 2015). This study confirmed the results of Sugiarti et al (2014), showing a reduction level of formalin up to 99.28% after 1 hour of submersion followed by the frying process (no information about the period of frying process) in squid. Frying could denature the protein and also hydrolyze its bonding with formalin (Levita et al 2010). The high temperature in frying increased the kinetic energy and led to faster movement of the molecules that could damage the molecular bonds (Sugiarti et al 2014).

Heat processing by boiling reduced the residue of formalin with up to 98.1%, more than the washing process (p<0.05), but at the same degree as the frying process (p>0.05). The high reduction of formalin in the boiling process was due to the accelerated formalin dissolution rate at higher temperatures (Annisak 2019). This effect could be enhanced by boiling the sample in an open pan (Kamal et al 2017).

The effect of storage on the formalin residue of *C. gariepinus*. This experiment demonstrated that only the period of storage significantly influenced the residue of formalin (p<0.05), while the temperature of storage and the combination between temperature and period of storage did not (Table 2). The previous different studies showed that a storage at the room temperature or at the freezing temperature had similar effects on the formalin residue decrease (Jawahar et al 2017; Murtini et al 2014), possibly due to the same level of protein degradation by breaking the bonds between protein and formalin and releasing free compounds. At the room temperature, the protein degradation was essentially caused by the microbial activity, while at the freezing temperature it was caused mostly by physical destruction, leading to conformational and functional changes (Murtini et al 2014; Yeasmin et al 2010). Formalin in the free form is more reactive and can also be more easily degraded (Riyanto et al 2006; Rachmawati et al 2007).

Conclusions. This research concluded that short time processing and storage treatments decreased the concentration of formalin. Heat involvement determined a more effective decrease of concentration (the decrease level was >90%). In the storage study, the residue of formalin was significantly reduced by the period of storage, mainly during the first 24 hours, then the formalin concentration reduction was insignificant, up to 48 hours of storage.

Acknowledgements. The authors are grateful to the Indonesian Ministry of Research, Technology, and Higher Education for providing the financial support in pursuing this research project (Program Kreativitas Mahasiswa Kemeristekdikti 2019 Program).

Reference

- Annisak S. K., Indriyanti N. Y., Mulyani B., 2019 [Constructive controversy and incubation presented according to the representation of tetrahedral chemical learning according to critical thinking ability]. Jurnal Inovasi Pendidikan IPA 5(1):10–22. [In Indonesian].
- Antoni S., 2010 [Analysis of formaldehyde content on salty fish by spectrophotometry method in Tampan District Pekanbaru]. BSc Thesis, Universitas Islam Negeri Sultan Syarif Kasim Riau, Indonesia, 53 p. [In Indonesian].
- Baroroh R. D., Kresnadipayana D., 2016 [The effect of variety consentration of starfruit juice (*Averrhoa bilimbi* L.) on soaked tofu for formaldehyde reduction]. BSc Thesis, Universitas Setia Budi, Indonesia, 1-2 p. [In Indonesian]. 2 pages thesis?This is the third time we address this issue. Thank you. we have only the resume of this research. we also have tried to contact the author, but still not success. could i

1093

AACL Bioflux, 2020, Volume 13, Issue X. http://www.bioflux.com.ro/aacl erase this reference from the list? our statement is still relevant with only one source.

- Fellows P. J., 2000 Food processing technology: principle and practice. Woodhead Publishing Limited and CRC Press LLC, Boca Roton, USA, 356 p.
- Handayani A., Alimin, Rustiah W. O., 2010 [The effect of low temperature storage (freezer -3oc) on water and fat content in lemuru fish (*Sardinella longiceps*)]. Al-Kimia (3):64–75. [In Indonesian].
- Hayun H., Harmita K., Pramudita T. B., 2017 Determination of formaldehyde content in wet noodles by thin layer chromatography-densitometry after derivatization with Nash reagent. Oriental Journal of Chemistry 33(3):1400-1405.
- Ichya'uddin M., 2014 [Analysis of Formaldehyde content and organoleptic test on salty fish in several traditional markets in Tuban]. BSc Thesis, Universitas Islam Negeri Maulana Malik Ibrahim, Indonesia, 112 p. [In Indonesian].
- Jawahar L. S., John C., Shafeekh M., Anupama T. K., Sankar T. V., 2017 Retention of residual formaldehyde in treated Indian mackerel (*Rastrelliger kanagurta*) under iced storage and related food safety concern. Indian Journal of Fisheries 64(4):87– 93.
- Joshi R., Bhatta R., Paudel P. N., Kafle B. K., 2015 Formaldehyde content of selected fish from the wet markets of Kathmandu valley. International Food Research Journal 22(4):1434-1437.
- Kamal A. S., Uddin M. M., Rahman R., Samira S., Rahman M., Nandy S., Khan M. S., 2017 Time and temperature effect on the residual concentration of formaldehyde in formalin treated samples of *Labeo rohita*. International Conference on Food Security and Nutrition, 6 p.
- Kaneko M., Wada Y., Fukui S., Kanno S., 1977 NII-Electronic library service. The Journal of Hygienic Chemistry 23(6):393–396.
- Laksmiani N., Widjaja I., Sonia, 2015 [Stability of formalin in different temperature and period of heating]. Jurnal Farmasi Udayana 4(2):76–81.
- Levita J., Musfiroh I., Indriyati W., Mustarichie R., 2010 The effect of soaking, washing and frying on the concentration of formaldehyde in Sange Belah salty fish. Jurnal Ilmu-ilmu Hayati dan Fisik 12(1):31–34.
- Male Y. T., Letsoin L. I., Siahaya N. A., 2017 [Analysis of formaldehyde content on noodle in several area in Ambon]. Jurnal Kementerian Perindustrian 13(2):5–10. [In Indonesian].
- Mudaffar R. A., 2018 [Qualitative and quantitative test of formaldehyde on apple, grape, and litchi for sale in Makassar]. Jurnal Perbal 6(3):59-65. [In Indonesian].
- Mudzkirah I., 2016 [Identification of borax and formalin on snack food in UIN Alaudin Makassar Canteen]. BSc Thesis, UIN Alauddin Makassar, Indonesia, 95 p. [In Indonesian].
- Muntaha A., Haitami, Hayati N., 2015 [Comparison of reduction formaldehyde level on boiled and hot water soaked tofu]. Medical Laboratory Technology Journal 1(2):84– 90. [In Indonesian].
- Murtini J. T., Riyanto R., Priyanto N., Hermana I., 2014 [Development of Formaldehyde on some kinds of seawater fish during storage in crushed ice]. JPB Perikanan 9(2):143–151. [In Indonesian].
- Pandey C. K., Agarwal A., Baronia A., Singh N., 2000 Toxicity of ingested formalin and its management. Human and Experiment Toxicology 360–366.
- Purawisastra S., Sahara E., 2011 [Absorption to formalin by certain foodstuff and disappearance through hot water soaked]. The Journal of Nutrition and Food Research 34(1):63–74. [In Indonesian].
- Rachmawati N., Riyanto R., Āriyani F., 2007 [Identification of formaldehyde on kerapu macan fish (*Ephinephelus fuscoguttatus*) during cold storage]. Jurnal Pascapanen dan Bioteknologi Kelautan dan Perikanan 2(2):137–145. [In Indonesian].
- Rahmadhani F., Safrida, Djufri, 2017 [The effect of soaking a various of solution for reduction formalin level on kembung salty fish (*Scomber canagorta*) in Lambaro traditional market Aceh]. Jurnal Ilmiah Mahasiswa Pendidikan Biologi 2(3):1-12. [In Indonesian].

AACL Bioflux, 2020, Volume 13, Issue X. http://www.bioflux.com.ro/aacl

1094

- Riyanto R., Kusmarwati A., Dwiyitno, 2006 [Identification of formaldehyde foorming on kerapu fish (*Epinephelus fuscoguttatus*) during room temperature storage]. Jurnal Pascapanen dan Bioteknologi Kelautan dan Perikanan 1(2):111–116. [In Indonesian].
- Shita A. E., 2016 [Selectivity of spectrophotometer analysis with Schiff's reagent]. BSc Thesis, Universitas Negeri Yogyakarta, Indonesia, 87 p. [In Indonesian].
- Sugiarti M., Anggo A. D., Riyadi P. H., 2014 [The effect of soaking in undercooking temperature and cooking method for reduction formalin level on cuttlefish]. Jurnal Pengolahan dan Bioteknologi Hasil Perikanan 3(2):90–98. [In Indonesian].
- Sundari D., Almasyhuri, Lamid A., 2015 [The effect of cooking process for nutrition composition of protein source]. Media Litbangkes 25(4):235–242. [In Indonesian].
- Suwanaruang T., 2018 Formalin contaminated in seafood and frozen meat at Somdet market, Kalasin Province. Journal of Environmental Protection 9:1286–1293.
- Wijayanti N. S., Lukitasari M., 2016 [Analysis of formalin level and organoleptic test in salty fish in Madiun Market]. Jurnal Florea 3(1):59–64. [In Indonesian].
- Wilianto W., Yudianto A., 2013 [The effect of formalin exposure on various level for DNA psoas major by PCR Lokus CSF1PO, D5S818, D13S317, D21S11 Analysis]. Jurnal Kedokteran Forensik Indonesia 15(1):11–16. [In Indonesian].
- Yeasmin T., Reza M. S., Shikha F. H., Khan M. N. A., Kamal M., 2010 Quality changes in formalin treated rohu fish (*Labeo rohita*, Hamilton) during ice storage condition. Asian Journal of Agricultural Science 2(4):158–163.
- Yulisa N., Asni E., Azrin M., 2014 [Formaldehyde test of gurame fish in Pekanbaru Traditional Market]. Jurnal Online Mahasiswa 1(2):1-12. [In Indonesian].
- Yusuf Y., Zuki Z., Amanda R. R., 2015 [The effect of various treatment for reduction formalin level on fish by spectrophotometry method]. Jurnal Riset Kimia 8(2):182–188. [In Indonesian].
- *** BPOM, Indonesian Food and Drug Authority, 2008 [Formaldehyde]. Badan POM RI, Jakarta, Indonesia. [In Indonesian].

Received: 15 October 2020. Accepted: 2020. Published online: 2020. Authors:

Isny Maulidevi, Alma Ata University, Faculty of Health Sciences, Department of Nutrition, Jl. Brawijaya 99, 55183 Yogyakarta, Indonesia, e-mail: isnymaulidevi540@gmail.com Nurul Andriyani, Alma Ata University, Faculty of Health Sciences, Department of Nutrition, Jl. Brawijaya 99,

Nurul Andriyani, Alma Ata University, Faculty of Health Sciences, Department of Nutrition, Jl. Brawijaya 99, 55183 Yogyakarta, Indonesia, e-mail: nurulandriyani96@gmail.com

Ryan Salfarino, Alma Ata University, Faculty of Health Sciences, Department of Nutrition, Jl. Brawijaya 99, 55183 Yogyakarta, Indonesia, e-mail: ryan.salfarino@gmail.com Veriani Aprilia, Alma Ata University, Faculty of Health Sciences, Department of Nutrition, Jl. Brawijaya 99,

55183 Yogvakarta, Indonesia, e-mail: verianiaprilia@almaata.ac.id

This is an open-access article distributed under the terms of the Creative Commons Attribution License, which

AACL Bioflux, 2020, Volume 13, Issue X. http://www.bioflux.com.ro/aacl permits unrestricted use, distribution and reproduction in any medium, provided the original author and source are credited. How to cite this article: Maulidevi I., Andriyani N., Salfarino R., Aprilia V., 2020 The residue of formalin in catfish (*Clarias gariepinus*) after processing and storage for short periods. AACL Bioflux 13(x):



verianiaprilia verianiaprilia <verianiaprilia@almaata.ac.id>

Pengiriman Bukti Pembayaran: 16 Desember 2020

your invoice, paid. Thank you! In attachment

1 pesan

Mag Ioan Valentin <zoobiomag2004@yahoo.com> Balas Ke: Mag Ioan Valentin <zoobiomag2004@yahoo.com> Kepada: Verianiaprilia Verianiaprilia <verianiaprilia@almaata.ac.id> 16 Desember 2020 pukul 23.28

your invoice, paid. Thank you! In attachment

Petrescu-Mag I. V., PhD, Hab. Associate Professor & Senior Researcher (CSI) USAMV Cluj & Bioflux SRL & University of Oradea, Tel: 0040 744470794 ; e-mail zoobiomag2004@yahoo.com Visit our journals: Aquaculture, Aquarium, Conservation & Legislation http://www. bioflux.com.ro/aacl Advances in Environmental Sciences www.aes.bioflux.com.ro Human & Veterinary Medicine www.hvm.bioflux.com.ro Advances in Agriculture & Botanics www.aab.bioflux.com.ro Animal Biology & Animal Husbandry www.abah.bioflux.com.ro Extreme Life, Biospeology & Astrobiology www.elba.bioflux.com.ro Porcine Research www.porc.bioflux.com.ro Rabbit Genetics www.rg.bioflux.com.ro ProEnvironment Promediu http://journals.usamvcluj.ro/index.php/promediu Poeciliid Research www.pr.bioflux.com.ro

aprilia.jpg 124K





verianiaprilia verianiaprilia <verianiaprilia@almaata.ac.id>

Pengiriman Draft Final: 20 Desember 2020

Final draft

4 pesan

Eni Kovacs <ek.bioflux@gmail.com> Kepada: verianiaprilia verianiaprilia <verianiaprilia@almaata.ac.id> 20 Desember 2020 pukul 03.34

Dear Veriani Aprilia,

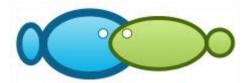
Could you please check the final form of the article attached, some adjustments were operated.

If you have any final observations and instructions prior to the article publishing, please inform us, any changes should be clearly highlighted. Change requests cannot be considered after publishing the material.

If all the authors agree with the final form of the article, could you please send us your acceptance for publishing, signed by all the authors.

Kind regards, Editor AACL Bioflux Eniko Kovacs

Aprilia_Clarias gariepinus_final.doc 168K



The residue of formalin in catfish (*Clarias* gariepinus) after processing and storage for short periods

İsny Maulidevi, Nurul Andriyani, Ryan Salfarino, Veriani Aprilia

Department of Nutrition, Faculty of Health Sciences, Alma Ata University, Yogyakarta, Indonesia. Corresponding author: V. Aprilia, verianiaprilia@almaata.ac.id

Abstract. Fresh fish is a highly perishable product due to its high protein content. A preservative may be needed to inhibit it. Nowadays, many fishes were reported to be exposed to formalin. This study aimed to determine the effect of short period processing (washing, frying, boiling in short-period) and storage (at the room and at freezing temperatures) on the residue of formalin in catfish, as a food model. Catfish, *Clarias gariepinus*, were divided into 8 groups, e.g. before treatment, washing in airflow for 1 minute, frying in 200°C for 8 minutes, boiling in 100°C for 7 minutes, storage at room temperature (29±1°C) for 24 and 48 hours, storage at freezing temperature (-3°C) for 24 and 48 hours. Before treatment, the catfish was soaked in 1,480 ppm of formalin solutions. The residue of formalin was measured at the end of each treatment by using a spectrophotometer. Data were then analyzed with analysis of variance with SPSS software. The results showed that there were significant effects of processing and storage treatment on the formalin residue of catfish. The formalin residue of all treatments was lower than before treatments. The frying processing and storage at freezing temperatures were the most effective treatments in reducing the formalin residue. **Key Words**: fresh fish, preservative, food model, treatments, formalin residue.

Introduction. Fresh fish is a highly perishable product due to its high protein content for microbial activity (Wijayanti & Lukitasari 2016). A preservative maybe needs to keep the freshness, inhibit the decay and prolong the shelf life (Antoni 2010). Nowadays, there are many kinds of food-grade preservatives available in the market. Unfortunately, formalin is still frequently found in food products, including fresh fish, being used among fishermen mainly because it is effective in preserving the product, cheaper and accessible (Male et al 2017).

Formalin is a forty percent solution of formaldehyde in water. This compound is detected in the animal body as an intermediary metabolite. It also uses for several purposes, such as in medicine for specimen preservation and the treatment of hemorrhage (Pandey et al 2000). The preservatives activity of formalin is due to its ability to react with protein generating methylene compounds, which are more difficult to be decayed by microbes (Ichya'uddin 2016). In low concentrations, formalin is not toxic for humans, but in higher concentrations, it had negative effects such as acute toxicity and death (Pandey et al 2000). It also decreases the levels of body antioxidants, resulting in liver damage (Yulisa et al 2014). Other studies also reported the cases of eczema, eye irritation, respiratory tract irritation, cancer, increased growth rate of malignant neoplastic diseases and denaturation of DNA (Suwanaruang 2018; Wilianto & Yudianto 2013). Therefore, its uses are forbidden.

Formalin solution is not stable during storage and exposure to high temperatures (Kaneko et al 1977; Laksmiani et al 2015). Those are caused by desolvation or evaporation of formalin during washing, boiling, or frying (Fellows 2000; Hayun et al 2017; Muntaha et al 2015; Sundari et al 2015) and enzymatic process in storage (Rachmawati et al 2007). Formalin that was in contact with fish decreased up to 43-53% after 5 times rinsing in 250 mL of distilled water and boiling for 30 minutes (Yusuf et al 2015). Other studies were also reported that soaking in warm water and frying decreased

the concentration of formalin up to 63.27% and 83.03%, respectively (Levita et al 2010). In daily practices, fish is usually washed in flowing water before processing, then fryed/boiled for a short time. However, there was no study explaining the influences of these processing and storage methods on the concentration of formalin.

The purpose of this study was to identify the effect of processing for a short time and storage (at room and freezing temperatures) on the residue of formalin. This study used catfish, *Clarias gariepinus*, as a food model.

Material and Method

Material. The study used fresh catfish, purchased from the traditional market in Yogyakarta, Indonesia. The other used ingredients were formalin solution and Schiff's reagent. Formalin solution was made from 4 mL of formalin 37% in 1,000 mL of distilled water (Rachmawati et al 2007).

Method. The study was divided into two sections, i.e. 1) analyzing the effect of processing and 2) analyzing the effect of storage treatment. Fresh catfish were slaughtered, eviscerated, cleaned, and soaked in 1,480 ppm formalin solution (Rachmawati et al 2007; Sugiarti et al 2014). For the first study, catfish were divided into four groups, i.e.: controls (before treatment), washing, frying, and boiling treatments. Washing was done by watering the samples with running water for 1 minute, while frying was done by frying the samples in oil at 200°C for 8 minutes. Boiling treatment was done by putting the samples into boiled water (100°C) for 7 minutes (Sugiarti et al 2014). For the second study, storage treatments, the catfish was also divided into four groups: room temperature storage (29°C) for 24 hours and 48 hours and freezing temperatures storage (-3°C) for 24 hours and 48 hours (Handayani et al 2010). At the end of treatments, the concentration of formalin in samples was analyzed using a spectrophotometer. As the preparation step, a 5 g sample was dissolved in 25 mL of distilled water. One mL of supernatant was mixed with 2 mL of Schiff's reagent, then heated in a water bath at 60°C for 5 minutes. It was incubated for 10 minutes before measuring the absorbance at 520 nm (Shita 2016; Mudaffar 2018).

Statistical analysis. Data were presented in mean±SD and analyzed with: (1) one-way analysis of variance (ANOVA) to determine the differences among all processing treatments and the effect of storage periods on the concentration of formalin; (2) paired sample T-test to determine the differences between the same samples before and after processing treatments; (3) independent sample t-test to know the effect of temperature at the same storage period, and (4) two-way ANOVA to determine the combination effect of storage temperature and period on the concentration of formalin. All data were analyzed with SPSS software (version 16.0 for Windows; SPSS Inc., Chicago, IL, USA) at the 5% significance level.

Results

Effect of processing on the formalin residue of C. gariepinus. Table 1 shows that all treatments had significant effects on the residue of formalin (p<0.05). It can be seen from the p-value between the same samples before and after treatments. Among all treatments, washing shows the highest residue of formalin (p<0.05). Both boiling and frying had an almost similar residue of formalin (p>0.05). The lower residue of formalin in boiling and frying may be due to heating exposure.

Table 1

Processing	Residue of formalin (ppm)		Δ residue of formalin
treatments	Before treatment	After treatment	<i>before and after treatment</i> (%)
Washing	1,420.17±66.94 ^{c1}	310.42±77.61 ^{b2}	78.14
Frying	1,420.17±66.94 ^{c1}	14.18 ± 6.69^{a2}	99.01
Boiling	1,420.17±66.94 ^{c1}	27.50±9.11 ^{a2}	98.10

Mean \pm SD in the same column with different alphabetical superscripts (a, b, or c) are significantly different (p<0.05) by using one-way ANOVA statistical analysis. Mean \pm SD in the same row with different number superscripts (1 or 2) are significantly different (p<0.05) by using paired sample T-test statistical analysis.

Effect of storage temperatures and periods on the formalin residue of C. gariepinus. Table 2 shows the effect of storage temperatures, of periods and of their combination on the concentration of formalin. Based on this data, the storage period influenced the residue of formalin decrease (p<0.05) during the first 24 hours, but after a storage period of 48 hours the decrease was not significantly different (p>0.05).

On the other hand, there were no differences in the residue of formalin stored at room and freezing temperature, suggesting that the temperature of storage did not affect the formalin residue (p>0.05). Although there were differences in the statistical results between the temperature effects and the storage periods effects, the combined effect of both variables showed no impact on the concentration of formalin (p>0.05).

Table 2

Effect of storage temperature and period on the formalin residue of *Clarias gariepinus*

Temperature	Periods (t) (hours)				p-value	9
<i>(T)</i>	0	24	48	Т	t	T*t
Room (29°C)	1,420.17±66.94 ^{a1}	52.18±28.78 ^{a2}	40.64±3.93 ^{a2}	0.20	0.00	0.76
Freezing (-3°C)	1,420.17±66.94 ^{a1}		13.65±4.23 ^{a2}	0.20	0.00	0.76

Mean \pm SD in the same column with different alphabetical superscripts (a, b, or c) are significantly different (p<0.05) and so are in the same row with different number superscript (1, 2, or 3). The p-value for T, t, and T*t was obtained from independent T-test, one-way ANOVA, and two-way ANOVA, respectively.

Discussion

The residue of formalin before treatment. The residue of formalin before and after treatment (either fish processing or storage) was significantly different (Table 1 and Table 2). In this study, the *C. gariepinus* specimens were submerged for 60 minutes in formalin solution of 1,480 ppm. The 60 minutes submersion period was chosen because the aldehyde compound may be readily bonded to the amino acids of fish in a small amount (Rahmadhani et al 2017). Formalin would attack lysine for the first time, followed by histidine and tyrosine, leading to the formation of a methylene compound (Ichya'uddin 2016). The protein-containing methylene compounds could not be digested by living things and microbes, especially pathogens, so that the shelf life would be longer (Purawisastra & Sahara 2011). The beneficial bacteria in the fish may also be killed by dehydration mechanism, as another effect of the existence of formalin in the body (Mudzkirah 2016).

The effect of processing on the formalin residue of C. gariepinus. The residue of formalin was reduced by 78.14% during washing treatment. The reduction was significant (p<0.05). The formalin was easily washed out because of its high solubility in the water, which is 4×10^5 mg L⁻¹ at 20°C (BPOM 2008; Sugiarti et al 2014). A study conducted by Yusuf et al (2015) showed the same trend, but the reduction was lower (with only 43%). The differences in the level of reduction can be affected by the duration

of formalin submersion. The longer the submersion time, the smaller the decrease, as more formalin was bonded with the fish protein.

In this study, frying also significantly reduced the residue of formalin (p<0.05). The reduction level was 99% that indicated most of the formalin disappeared during the frying process. Formalin was included in a volatile compound, especially above its boiling point (Joshi et al 2015). This study confirmed the results of Sugiarti et al (2014), showing a reduction level of formalin up to 99.28% after 1 hour of submersion followed by the frying process (no information about the period of frying process) in squid. Frying could denature the protein and also hydrolyze its bonding with formalin (Levita et al 2010). The high temperature in frying increased the kinetic energy and led to faster movement of the molecules that could damage the molecular bonds (Sugiarti et al 2014).

Heat processing by boiling reduced the residue of formalin with up to 98.1%, more than the washing process (p<0.05), but at the same degree as the frying process (p>0.05). The high reduction of formalin in the boiling process was due to the accelerated formalin dissolution rate at higher temperatures (Annisak 2019). This effect could be enhanced by boiling the sample in an open pan (Kamal et al 2017).

The effect of storage on the formalin residue of C. gariepinus. This experiment demonstrated that only the period of storage significantly influenced the residue of formalin (p<0.05), while the temperature of storage and the combination between temperature and period of storage did not (Table 2). The previous different studies showed that a storage at the room temperature or at the freezing temperature had similar effects on the formalin residue decrease (Jawahar et al 2017; Murtini et al 2014), possibly due to the same level of protein degradation by breaking the bonds between protein and formalin and releasing free compounds. At the room temperature, the protein degradation was essentially caused by the microbial activity, while at the freezing temperature it was caused mostly by physical destruction, leading to conformational and functional changes (Murtini et al 2014; Yeasmin et al 2010). Formalin in the free form is more reactive and can also be more easily degraded (Riyanto et al 2006; Rachmawati et al 2007).

Conclusions. This research concluded that short time processing and storage treatments decreased the concentration of formalin. Heat involvement determined a more effective decrease of concentration (the decrease level was >90%). In the storage study, the residue of formalin was significantly reduced by the period of storage, mainly during the first 24 hours, then the formalin concentration reduction was insignificant, up to 48 hours of storage.

Acknowledgements. The authors are grateful to the Indonesian Ministry of Research, Technology, and Higher Education for providing the financial support in pursuing this research project (Program Kreativitas Mahasiswa Kemeristekdikti 2019 Program).

Reference

- Annisak S. K., Indriyanti N. Y., Mulyani B., 2019 [Constructive controversy and incubation presented according to the representation of tetrahedral chemical learning according to critical thinking ability]. Jurnal Inovasi Pendidikan IPA 5(1):10–22. [In Indonesian].
- Antoni S., 2010 [Analysis of formaldehyde content on salty fish by spectrophotometry method in Tampan District Pekanbaru]. BSc Thesis, Universitas Islam Negeri Sultan Syarif Kasim Riau, Indonesia, 53 p. [In Indonesian].
- Fellows P. J., 2000 Food processing technology: principle and practice. Woodhead Publishing Limited and CRC Press LLC, Boca Roton, USA, 356 p.
- Handayani A., Alimin, Rustiah W. O., 2010 [The effect of low temperature storage (freezer -3oc) on water and fat content in lemuru fish (*Sardinella longiceps*)]. Al-Kimia (3):64–75. [In Indonesian].

- Hayun H., Harmita K., Pramudita T. B., 2017 Determination of formaldehyde content in wet noodles by thin layer chromatography-densitometry after derivatization with Nash reagent. Oriental Journal of Chemistry 33(3):1400-1405.
- Ichya'uddin M., 2014 [Analysis of Formaldehyde content and organoleptic test on salty fish in several traditional markets in Tuban]. BSc Thesis, Universitas Islam Negeri Maulana Malik Ibrahim, Indonesia, 112 p. [In Indonesian].
- Jawahar L. S., John C., Shafeekh M., Anupama T. K., Sankar T. V., 2017 Retention of residual formaldehyde in treated Indian mackerel (*Rastrelliger kanagurta*) under iced storage and related food safety concern. Indian Journal of Fisheries 64(4):87–93.
- Joshi R., Bhatta R., Paudel P. N., Kafle B. K., 2015 Formaldehyde content of selected fish from the wet markets of Kathmandu valley. International Food Research Journal 22(4):1434-1437.
- Kamal A. S., Uddin M. M., Rahman R., Samira S., Rahman M., Nandy S., Khan M. S., 2017 Time and temperature effect on the residual concentration of formaldehyde in formalin treated samples of *Labeo rohita*. International Conference on Food Security and Nutrition, 6 p.
- Kaneko M., Wada Y., Fukui S., Kanno S., 1977 NII-Electronic library service. The Journal of Hygienic Chemistry 23(6):393–396.
- Laksmiani N., Widjaja I., Sonia, 2015 [Stability of formalin in different temperature and period of heating]. Jurnal Farmasi Udayana 4(2):76–81.
- Levita J., Musfiroh I., Indriyati W., Mustarichie R., 2010 The effect of soaking, washing and frying on the concentration of formaldehyde in Sange Belah salty fish. Jurnal Ilmu-ilmu Hayati dan Fisik 12(1):31–34.
- Male Y. T., Letsoin L. I., Siahaya N. A., 2017 [Analysis of formaldehyde content on noodle in several area in Ambon]. Jurnal Kementerian Perindustrian 13(2):5–10. [In Indonesian].
- Mudaffar R. A., 2018 [Qualitative and quantitative test of formaldehyde on apple, grape, and litchi for sale in Makassar]. Jurnal Perbal 6(3):59-65. [In Indonesian].
- Mudzkirah I., 2016 [Identification of borax and formalin on snack food in UIN Alaudin Makassar Canteen]. BSc Thesis, UIN Alauddin Makassar, Indonesia, 95 p. [In Indonesian].
- Muntaha A., Haitami, Hayati N., 2015 [Comparison of reduction formaldehyde level on boiled and hot water soaked tofu]. Medical Laboratory Technology Journal 1(2):84– 90. [In Indonesian].
- Murtini J. T., Riyanto R., Priyanto N., Hermana I., 2014 [Development of Formaldehyde on some kinds of seawater fish during storage in crushed ice]. JPB Perikanan 9(2):143–151. [In Indonesian].
- Pandey C. K., Agarwal A., Baronia A., Singh N., 2000 Toxicity of ingested formalin and its management. Human and Experiment Toxicology 360–366.
- Purawisastra S., Sahara E., 2011 [Absorption to formalin by certain foodstuff and disappearance through hot water soaked]. The Journal of Nutrition and Food Research 34(1):63–74. [In Indonesian].
- Rachmawati N., Riyanto R., Ariyani F., 2007 [Identification of formaldehyde on kerapu macan fish (*Ephinephelus fuscoguttatus*) during cold storage]. Jurnal Pascapanen dan Bioteknologi Kelautan dan Perikanan 2(2):137–145. [In Indonesian].
- Rahmadhani F., Safrida, Djufri, 2017 [The effect of soaking a various of solution for reduction formalin level on kembung salty fish (*Scomber canagorta*) in Lambaro traditional market Aceh]. Jurnal Ilmiah Mahasiswa Pendidikan Biologi 2(3):1-12. [In Indonesian].
- Riyanto R., Kusmarwati A., Dwiyitno, 2006 [Identification of formaldehyde foorming on kerapu fish (*Epinephelus fuscoguttatus*) during room temperature storage]. Jurnal Pascapanen dan Bioteknologi Kelautan dan Perikanan 1(2):111–116. [In Indonesian].
- Shita A. E., 2016 [Selectivity of spectrophotometer analysis with Schiff's reagent]. BSc Thesis, Universitas Negeri Yogyakarta, Indonesia, 87 p. [In Indonesian].

Sugiarti M., Anggo A. D., Riyadi P. H., 2014 [The effect of soaking in undercooking temperature and cooking method for reduction formalin level on cuttlefish]. Jurnal Pengolahan dan Bioteknologi Hasil Perikanan 3(2):90–98. [In Indonesian].

- Sundari D., Almasyhuri, Lamid A., 2015 [The effect of cooking process for nutrition composition of protein source]. Media Litbangkes 25(4):235–242. [In Indonesian].
- Suwanaruang T., 2018 Formalin contaminated in seafood and frozen meat at Somdet market, Kalasin Province. Journal of Environmental Protection 9:1286–1293.
- Wijayanti N. S., Lukitasari M., 2016 [Analysis of formalin level and organoleptic test in salty fish in Madiun Market]. Jurnal Florea 3(1):59–64. [In Indonesian].
- Wilianto W., Yudianto A., 2013 [The effect of formalin exposure on various level for DNA psoas major by PCR Lokus CSF1PO, D5S818, D13S317, D21S11 Analysis]. Jurnal Kedokteran Forensik Indonesia 15(1):11–16. [In Indonesian].
- Yeasmin T., Reza M. S., Shikha F. H., Khan M. N. A., Kamal M., 2010 Quality changes in formalin treated rohu fish (*Labeo rohita*, Hamilton) during ice storage condition. Asian Journal of Agricultural Science 2(4):158–163.
- Yulisa N., Asni E., Azrin M., 2014 [Formaldehyde test of gurame fish in Pekanbaru Traditional Market]. Jurnal Online Mahasiswa 1(2):1-12. [In Indonesian].
- Yusuf Y., Zuki Z., Amanda R. R., 2015 [The effect of various treatment for reduction formalin level on fish by spectrophotometry method]. Jurnal Riset Kimia 8(2):182– 188. [In Indonesian].
- *** BPOM, Indonesian Food and Drug Authority, 2008 [Formaldehyde]. Badan POM RI, Jakarta, Indonesia. [In Indonesian].

Received: 15 October 2020. Accepted: 2020. Published online: 2020. Authors:

Isny Maulidevi, Alma Ata University, Faculty of Health Sciences, Department of Nutrition, Jl. Brawijaya 99, 55183 Yogyakarta, Indonesia, e-mail: isnymaulidevi540@gmail.com

Nurul Andriyani, Alma Ata University, Faculty of Health Sciences, Department of Nutrition, Jl. Brawijaya 99, 55183 Yogyakarta, Indonesia, e-mail: nurulandriyani96@gmail.com

Ryan Salfarino, Alma Ata University, Faculty of Health Sciences, Department of Nutrition, Jl. Brawijaya 99, 55183 Yogyakarta, Indonesia, e-mail: ryan.salfarino@gmail.com

Veriani Aprilia, Alma Ata University, Faculty of Health Sciences, Department of Nutrition, Jl. Brawijaya 99, 55183 Yogyakarta, Indonesia, e-mail: verianiaprilia@almaata.ac.id

How to cite this article:

Maulidevi I., Andriyani N., Salfarino R., Aprilia V., 2020 The residue of formalin in catfish (*Clarias gariepinus*) after processing and storage for short periods. AACL Bioflux 13(6):

This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution and reproduction in any medium, provided the original author and source are credited.



verianiaprilia verianiaprilia <verianiaprilia@almaata.ac.id>

23 Desember 2020 pukul 00.46

Revisi Draft Final: 23 Desember 2020

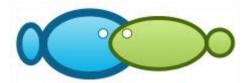
Kepada: Eni Kovacs <ek.bioflux@gmail.com>

Dear Kovacs

This is our revision. Several grammatical errors were found after using the Grammarly software. thank you

Regards Veriani Aprilia [Kutipan teks disembunyikan]

Aprilia_Clarias gariepinus_final6.doc



The residue of formalin in catfish (*Clarias* gariepinus) after processing and storage for short periods

Isny Maulidevi, Nurul Andriyani, Ryan Salfarino, Veriani Aprilia

Department of Nutrition, Faculty of Health Sciences, Universitas Alma Ata, Yogyakarta, Indonesia. Corresponding author: V. Aprilia, verianiaprilia@almaata.ac.id

Abstract. Fresh fish is a highly perishable product due to its high protein content. A preservative may be needed to inhibit it. Nowadays, many fishes were reported to be exposed to formalin. This study aimed to determine the effect of short period processing (washing, frying, boiling in short-period) and storage (at the room and at freezing temperatures) on the residue of formalin in catfish, as a food model. Catfish, *Clarias gariepinus*, were divided into 8 groups, e.g. before treatment, washing in airflow for 1 minute, frying in 200°C for 8 minutes, boiling in 100°C for 7 minutes, storage at room temperature (29±1°C) for 24 and 48 hours, storage at freezing temperature (-3°C) for 24 and 48 hours. Before treatment, the catfish was soaked in 1,480 ppm of formalin solutions. The residue of formalin was measured at the end of each treatment by using a spectrophotometer. Data were then analyzed with analysis of variance with SPSS software. The results showed that there were significant effects of processing and storage treatment on the formalin residue of catfish. The formalin residue of all treatments was lower than before treatments. The frying processing and storage at freezing temperatures were the most effective treatments in reducing the formalin residue. **Key Words**: fresh fish, preservative, food model, treatments, formalin residue.

Introduction. Fresh fish is a highly perishable product due to its high protein content for microbial activity (Wijayanti & Lukitasari 2016). A preservative maybe needs to keep the freshness, inhibit the decay and prolong the shelf life (Antoni 2010). Nowadays, there are many kinds of food-grade preservatives available in the market. Unfortunately, formalin is still frequently found in food products, including fresh fish, being used among fishermen mainly because it is effective in preserving the product, cheaper and accessible (Male et al 2017).

Formalin is a forty percent solution of formaldehyde in water. This compound is detected in the animal body as an intermediary metabolite. It also uses for several purposes, such as in medicine for specimen preservation and the treatment of hemorrhage (Pandey et al 2000). The preservative activity of formalin is due to its ability to react with protein generating methylene compounds, which are more difficult to be decayed by microbes (Ichya'uddin 2016). In low concentrations, formalin is not toxic for humans, but in higher concentrations, it had negative effects such as acute toxicity and death (Pandey et al 2000). It also decreases the levels of body antioxidants, resulting in liver damage (Yulisa et al 2014). Other studies also reported the cases of eczema, eye irritation, respiratory tract irritation, cancer, increased growth rate of malignant neoplastic diseases and denaturation of DNA (Suwanaruang 2018; Wilianto & Yudianto 2013). Therefore, its uses are forbidden.

Formalin solution is not stable during storage and exposure to high temperatures (Kaneko et al 1977; Laksmiani et al 2015). Those are caused by desolvation or evaporation of formalin during washing, boiling, or frying (Fellows 2000; Hayun et al 2017; Muntaha et al 2015; Sundari et al 2015) and enzymatic process in storage (Rachmawati et al 2007). Formalin that was in contact with fish decreased up to 43-53% after 5 times rinsing in 250 mL of distilled water and boiling for 30 minutes (Yusuf et al 2015). Other studies were also reported that soaking in warm water and frying decreased

the concentration of formalin up to 63.27% and 83.03%, respectively (Levita et al 2010). In daily practices, fish is usually washed in flowing water before processing, then fried/boiled for a short time. However, there was no study explaining the influences of these processing and storage methods on the concentration of formalin.

The purpose of this study was to identify the effect of processing for a short time and storage (at room and freezing temperatures) on the residue of formalin. This study used catfish, *Clarias gariepinus*, as a food model.

Material and Method

Material. The study used fresh catfish, purchased from the traditional market in Yogyakarta, Indonesia. The other used ingredients were formalin solution and Schiff's reagent. Formalin solution was made from 4 mL of formalin 37% in 1,000 mL of distilled water (Rachmawati et al 2007).

Method. The study was divided into two sections, i.e. 1) analyzing the effect of processing and 2) analyzing the effect of storage treatment. Fresh catfish were slaughtered, eviscerated, cleaned, and soaked in 1,480 ppm formalin solution (Rachmawati et al 2007; Sugiarti et al 2014). For the first study, catfish were divided into four groups, i.e.: controls (before treatment), washing, frying, and boiling treatments. Washing was done by watering the samples with running water for 1 minute, while frying was done by frying the samples in oil at 200°C for 8 minutes. Boiling treatment was done by putting the samples into boiled water (100°C) for 7 minutes (Sugiarti et al 2014). For the second study, storage treatments, the catfish was also divided into four groups: room temperature storage (29°C) for 24 hours and 48 hours and freezing temperatures storage (-3°C) for 24 hours and 48 hours (Handayani et al 2010). At the end of treatments, the concentration of formalin in samples was analyzed using a spectrophotometer. As the preparation step, a 5 g sample was dissolved in 25 mL of distilled water. One mL of supernatant was mixed with 2 mL of Schiff's reagent, then heated in a water bath at 60°C for 5 minutes. It was incubated for 10 minutes before measuring the absorbance at 520 nm (Shita 2016; Mudaffar 2018).

Statistical analysis. Data were presented in mean±SD and analyzed with: (1) one-way analysis of variance (ANOVA) to determine the differences among all processing treatments and the effect of storage periods on the concentration of formalin; (2) paired sample T-test to determine the differences between the same samples before and after processing treatments; (3) independent sample t-test to know the effect of temperature at the same storage period, and (4) two-way ANOVA to determine the **combined** effect of storage temperature and period on the concentration of formalin. All data were analyzed with SPSS software (version 16.0 for Windows; SPSS Inc., Chicago, IL, USA) at the 5% significance level.

Results

Effect of processing on the formalin residue of C. gariepinus. Table 1 shows that all treatments had significant effects on the residue of formalin (p<0.05). It can be seen from the p-value between the same samples before and after treatments. Among all treatments, washing shows the highest residue of formalin (p<0.05). Both boiling and frying had an almost similar residue of formalin (p>0.05). The lower residue of formalin in boiling and frying may be due to heating exposure.

Table 1

Processing	Residue of formalin (ppm)		Δ residue of formalin
treatments	Before treatment	After treatment	<i>before and after treatment</i> (%)
Washing	1,420.17±66.94 ^{c1}	310.42±77.61 ^{b2}	78.14
Frying	1,420.17±66.94 ^{c1}	14.18 ± 6.69^{a2}	99.01
Boiling	1,420.17±66.94 ^{c1}	27.50±9.11 ^{a2}	98.10

Mean \pm SD in the same column with different alphabetical superscripts (a, b, or c) are significantly different (p<0.05) by using one-way ANOVA statistical analysis. Mean \pm SD in the same row with different number superscripts (1 or 2) are significantly different (p<0.05) by using paired sample T-test statistical analysis.

Effect of storage temperatures and periods on the formalin residue of C. gariepinus. Table 2 shows the effect of storage temperatures, of periods and of their combination on the concentration of formalin. Based on this data, the storage period influenced the residue of formalin decrease (p<0.05) during the first 24 hours, but after a storage period of 48 hours the decrease was not significantly different (p>0.05).

On the other hand, there were no differences in the residue of formalin stored at room and freezing temperature, suggesting that the temperature of storage did not affect the formalin residue (p>0.05). Although there were differences in the statistical results between the temperature effects and the storage periods effects, the combined effect of both variables showed no impact on the concentration of formalin (p>0.05).

Table 2

Effect of storage temperature and period on the formalin residue of *Clarias gariepinus*

Temperature	Periods (t) (hours)			p-value		
<i>(T)</i>	0	24	48	Т	t	T*t
Room (29°C)	1,420.17±66.94 ^{a1}	52.18±28.78 ^{a2}	40.64±3.93 ^{a2}	0.20	0.00	0.76
Freezing (-3°C)	1,420.17±66.94 ^{a1}		13.65±4.23 ^{a2}	0.20	0.00	0.76

Mean \pm SD in the same column with different alphabetical superscripts (a, b, or c) are significantly different (p<0.05) and so are in the same row with different number superscript (1, 2, or 3). The p-value for T, t, and T*t was obtained from independent T-test, one-way ANOVA, and two-way ANOVA, respectively.

Discussion

The residue of formalin before treatment. The residue of formalin before and after treatment (either fish processing or storage) was significantly different (Table 1 and Table 2). In this study, the *C. gariepinus* specimens were submerged for 60 minutes in formalin solution of 1,480 ppm. The 60 minutes submersion period was chosen because the aldehyde compound may be readily bonded to the amino acids of fish in a small amount (Rahmadhani et al 2017). Formalin would attack lysine for the first time, followed by histidine and tyrosine, leading to the formation of a methylene compound (Ichya'uddin 2016). The protein-containing methylene compounds could not be digested by living things and microbes, especially pathogens, so that the shelf life would be longer (Purawisastra & Sahara 2011). The beneficial bacteria in the fish may also be killed by dehydration mechanism, as another effect of the existence of formalin in the body (Mudzkirah 2016).

The effect of processing on the formalin residue of C. gariepinus. The residue of formalin was reduced by 78.14% during washing treatment. The reduction was significant (p<0.05). The formalin was easily washed out because of its high solubility in the water, which is 4×10^5 mg L⁻¹ at 20°C (BPOM 2008; Sugiarti et al 2014). A study conducted by Yusuf et al (2015) showed the same trend, but the reduction was lower (with only 43%). The differences in the level of reduction can be affected by the duration

- Hayun H., Harmita K., Pramudita T. B., 2017 Determination of formaldehyde content in wet noodles by thin layer chromatography-densitometry after derivatization with Nash reagent. Oriental Journal of Chemistry 33(3):1400-1405.
- Ichya'uddin M., 2014 [Analysis of Formaldehyde content and organoleptic test on salty fish in several traditional markets in Tuban]. BSc Thesis, Universitas Islam Negeri Maulana Malik Ibrahim, Indonesia, 112 p. [In Indonesian].
- Jawahar L. S., John C., Shafeekh M., Anupama T. K., Sankar T. V., 2017 Retention of residual formaldehyde in treated Indian mackerel (*Rastrelliger kanagurta*) under iced storage and related food safety concern. Indian Journal of Fisheries 64(4):87–93.
- Joshi R., Bhatta R., Paudel P. N., Kafle B. K., 2015 Formaldehyde content of selected fish from the wet markets of Kathmandu valley. International Food Research Journal 22(4):1434-1437.
- Kamal A. S., Uddin M. M., Rahman R., Samira S., Rahman M., Nandy S., Khan M. S., 2017 Time and temperature effect on the residual concentration of formaldehyde in formalin treated samples of *Labeo rohita*. International Conference on Food Security and Nutrition, 6 p.
- Kaneko M., Wada Y., Fukui S., Kanno S., 1977 NII-Electronic library service. The Journal of Hygienic Chemistry 23(6):393–396.
- Laksmiani N., Widjaja I., Sonia, 2015 [Stability of formalin in different temperature and period of heating]. Jurnal Farmasi Udayana 4(2):76–81.
- Levita J., Musfiroh I., Indriyati W., Mustarichie R., 2010 The effect of soaking, washing and frying on the concentration of formaldehyde in Sange Belah salty fish. Jurnal Ilmu-ilmu Hayati dan Fisik 12(1):31–34.
- Male Y. T., Letsoin L. I., Siahaya N. A., 2017 [Analysis of formaldehyde content on noodle in several areas in Ambon]. Jurnal Kementerian Perindustrian 13(2):5–10. [In Indonesian].
- Mudaffar R. A., 2018 [Qualitative and quantitative test of formaldehyde on apple, grape, and litchi for sale in Makassar]. Jurnal Perbal 6(3):59-65. [In Indonesian].
- Mudzkirah I., 2016 [Identification of borax and formalin on snack food in UIN Alaudin Makassar Canteen]. BSc Thesis, UIN Alauddin Makassar, Indonesia, 95 p. [In Indonesian].
- Muntaha A., Haitami, Hayati N., 2015 [Comparison of reduction formaldehyde level on boiled and hot water soaked tofu]. Medical Laboratory Technology Journal 1(2):84– 90. [In Indonesian].
- Murtini J. T., Riyanto R., Priyanto N., Hermana I., 2014 [Development of Formaldehyde on some kinds of seawater fish during storage in crushed ice]. JPB Perikanan 9(2):143–151. [In Indonesian].
- Pandey C. K., Agarwal A., Baronia A., Singh N., 2000 Toxicity of ingested formalin and its management. Human and Experiment Toxicology 360–366.
- Purawisastra S., Sahara E., 2011 [Absorption to formalin by certain foodstuff and disappearance through hot water soaked]. The Journal of Nutrition and Food Research 34(1):63–74. [In Indonesian].
- Rachmawati N., Riyanto R., Ariyani F., 2007 [Identification of formaldehyde on kerapu macan fish (*Ephinephelus fuscoguttatus*) during cold storage]. Jurnal Pascapanen dan Bioteknologi Kelautan dan Perikanan 2(2):137–145. [In Indonesian].
- Rahmadhani F., Safrida, Djufri, 2017 [The effect of soaking a various of solution for reduction formalin level on kembung salty fish (*Scomber canagorta*) in Lambaro traditional market Aceh]. Jurnal Ilmiah Mahasiswa Pendidikan Biologi 2(3):1-12. [In Indonesian].
- Riyanto R., Kusmarwati A., Dwiyitno, 2006 [Identification of formaldehyde forming on kerapu fish (*Epinephelus fuscoguttatus*) during room temperature storage]. Jurnal Pascapanen dan Bioteknologi Kelautan dan Perikanan 1(2):111–116. [In Indonesian].
- Shita A. E., 2016 [Selectivity of spectrophotometer analysis with Schiff's reagent]. BSc Thesis, Universitas Negeri Yogyakarta, Indonesia, 87 p. [In Indonesian].

of formalin submersion. The longer the submersion time, the smaller the decrease, as more formalin was bonded with the fish protein.

In this study, frying also significantly reduced the residue of formalin (p<0.05). The reduction level was 99% that indicated most of the formalin disappeared during the frying process. Formalin was included in a volatile compound, especially above its boiling point (Joshi et al 2015). This study confirmed the results of Sugiarti et al (2014), showing a reduction level of formalin up to 99.28% after 1 hour of submersion followed by the frying process (no information about the period of frying process) in squid. Frying could denature the protein and also hydrolyze its bonding with formalin (Levita et al 2010). The high temperature in frying increased the kinetic energy and led to faster movement of the molecules that could damage the molecular bonds (Sugiarti et al 2014).

Heat processing by boiling reduced the residue of formalin by up to 98.1%, more than the washing process (p<0.05), but at the same degree as the frying process (p>0.05). The high reduction of formalin in the boiling process was due to the accelerated formalin dissolution rate at higher temperatures (Annisak 2019). This effect could be enhanced by boiling the sample in an open pan (Kamal et al 2017).

The effect of storage on the formalin residue of C. gariepinus. This experiment demonstrated that only the period of storage significantly influenced the residue of formalin (p<0.05), while the temperature of storage and the combination between temperature and period of storage did not (Table 2). The previous different studies showed that a storage at the room temperature or at the freezing temperature had similar effects on the formalin residue decrease (Jawahar et al 2017; Murtini et al 2014), possibly due to the same level of protein degradation by breaking the bonds between protein and formalin and releasing free compounds. At the room temperature, the protein degradation was essentially caused by the microbial activity, while at the freezing temperature it was caused mostly by physical destruction, leading to conformational and functional changes (Murtini et al 2014; Yeasmin et al 2010). Formalin in the free form is more reactive and can also be more easily degraded (Riyanto et al 2006; Rachmawati et al 2007).

Conclusions. This research concluded that short-time processing and storage treatments decreased the concentration of formalin. Heat involvement determined a more effective decrease in concentration (the decrease level was >90%). In the storage study, the residue of formalin was significantly reduced by the period of storage, mainly during the first 24 hours, then the formalin concentration reduction was insignificant, up to 48 hours of storage.

Acknowledgements. The authors are grateful to the Indonesian Ministry of Research, Technology, and Higher Education for providing the financial support in pursuing this research project (Program Kreativitas Mahasiswa Kemeristekdikti 2019 Program).

Reference

- Annisak S. K., Indriyanti N. Y., Mulyani B., 2019 [Constructive controversy and incubation presented according to the representation of tetrahedral chemical learning according to critical thinking ability]. Jurnal Inovasi Pendidikan IPA 5(1):10–22. [In Indonesian].
- Antoni S., 2010 [Analysis of formaldehyde content on salty fish by spectrophotometry method in Tampan District Pekanbaru]. BSc Thesis, Universitas Islam Negeri Sultan Syarif Kasim Riau, Indonesia, 53 p. [In Indonesian].
- Fellows P. J., 2000 Food processing technology: principle and practice. Woodhead Publishing Limited and CRC Press LLC, Boca Roton, USA, 356 p.
- Handayani A., Alimin, Rustiah W. O., 2010 [The effect of low-temperature storage (freezer -3oc) on water and fat content in lemuru fish (*Sardinella longiceps*)]. Al-Kimia (3):64–75. [In Indonesian].

Sugiarti M., Anggo A. D., Riyadi P. H., 2014 [The effect of soaking in undercooking temperature and cooking method for reduction formalin level on cuttlefish]. Jurnal Pengolahan dan Bioteknologi Hasil Perikanan 3(2):90–98. [In Indonesian].

- Sundari D., Almasyhuri, Lamid A., 2015 [The effect of cooking process for nutrition composition of protein source]. Media Litbangkes 25(4):235–242. [In Indonesian].
- Suwanaruang T., 2018 Formalin contaminated in seafood and frozen meat at Somdet market, Kalasin Province. Journal of Environmental Protection 9:1286–1293.
- Wijayanti N. S., Lukitasari M., 2016 [Analysis of formalin level and organoleptic test in salty fish in Madiun Market]. Jurnal Florea 3(1):59–64. [In Indonesian].
- Wilianto W., Yudianto A., 2013 [The effect of formalin exposure on various levels for DNA psoas major by PCR Lokus CSF1PO, D5S818, D13S317, D21S11 Analysis]. Jurnal Kedokteran Forensik Indonesia 15(1):11–16. [In Indonesian].
- Yeasmin T., Reza M. S., Shikha F. H., Khan M. N. A., Kamal M., 2010 Quality changes in formalin treated rohu fish (*Labeo rohita*, Hamilton) during ice storage condition. Asian Journal of Agricultural Science 2(4):158–163.
- Yulisa N., Asni E., Azrin M., 2014 [Formaldehyde test of gurame fish in Pekanbaru Traditional Market]. Jurnal Online Mahasiswa 1(2):1-12. [In Indonesian].
- Yusuf Y., Zuki Z., Amanda R. R., 2015 [The effect of various treatments for reduction formalin level on fish by spectrophotometry method]. Jurnal Riset Kimia 8(2):182–188. [In Indonesian].
- *** BPOM, Indonesian Food and Drug Authority, 2008 [Formaldehyde]. Badan POM RI, Jakarta, Indonesia. [In Indonesian].

Received: 15 October 2020. Accepted: 2020. Published online: 2020. Authors:

Isny Maulidevi, Universitas Alma Ata, Faculty of Health Sciences, Department of Nutrition, Jl. Brawijaya 99, 55183 Yogyakarta, Indonesia, e-mail: isnymaulidevi540@gmail.com

Nurul Andriyani, Universitas Alma Ata, Faculty of Health Sciences, Department of Nutrition, Jl. Brawijaya 99, 55183 Yogyakarta, Indonesia, e-mail: nurulandriyani96@gmail.com

Ryan Salfarino, Universitas Alma Ata, Faculty of Health Sciences, Department of Nutrition, Jl. Brawijaya 99, 55183 Yogyakarta, Indonesia, e-mail: ryan.salfarino@gmail.com

Veriani Aprilia, Universitas Alma Ata, Faculty of Health Sciences, Department of Nutrition, Jl. Brawijaya 99, 55183 Yogyakarta, Indonesia, e-mail: verianiaprilia@almaata.ac.id

How to cite this article:

This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution and reproduction in any medium, provided the original author and source are credited.

Maulidevi I., Andriyani N., Salfarino R., Aprilia V., 2020 The residue of formalin in catfish (*Clarias gariepinus*) after processing and storage for short periods. AACL Bioflux 13(6):



Balasan Revisi Draft Final: 27 Desember 2020

Eni Kovacs <ek.bioflux@gmail.com> Kepada: verianiaprilia verianiaprilia <verianiaprilia@almaata.ac.id> 27 Desember 2020 pukul 20.32

Dear Veriani Aprilia,

As mentioned in our previous e-mail:

" If all the authors agree with the final form of the article, could you please send us your acceptance for publishing, signed by all the authors."

As the manuscript is in English, we will use Alma Ata University.

Thank you for your cooperation. [Kutipan teks disembunyikan]



Persetujuan Publikasi: 28 Desember 2020

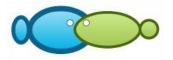
verianiaprilia verianiaprilia <verianiaprilia@almaata.ac.id> Kepada: Eni Kovacs <ek.bioflux@gmail.com>

Dear Kovacs,

Here are letter of acceptance for publishing of the articles. Thank you for your great assistance

Regards Veriani Aprilia [Kutipan teks disembunyikan]

acceptance for publishing_veriani aprilia.pdf 103K 28 Desember 2020 pukul 15.09



Acceptance for Publishing-Letter

Article title: The residue of formalin in catfish (Clarias gariepinus) after a short period processing and storage

Name of the authors: Isny Maulidevi, Nurul Andrivani, Ryan Salfarino, Veriani Aprilia

Hereby I would like to stated that we accept the final form of the manuscript entitled "The residue of formalin in catfish (Clarias gariepinus) after a short period processing and storage" to Aquaculture, Aquarium, Conservation & Legislation - International Journal of the Bioflux Society.

Authors

Isny Maulidevi



Ryan Salfarino Veriani Aprilia

h.

Date: December 26th, 2020



Pemberitahuan Publikasi Manuskrip: 29 Desember 2020

verianiaprilia verianiaprilia <verianiaprilia@almaata.ac.id>

Published manuscript

1 pesan

Eni Kovacs <ek.bioflux@gmail.com> Kepada: verianiaprilia verianiaprilia <verianiaprilia@almaata.ac.id> 29 Desember 2020 pukul 15.24

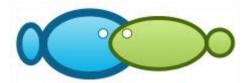
Dear Veriani Aprilia,

We would like to inform you that your manuscript has been published:

http://www.bioflux.com.ro/docs/2020.3441-3446.pdf

Thank you for publishing with us and best of luck in your future research.

Kind regards, Editor AACL Bioflux Eniko Kovacs



The residue of formalin in catfish (*Clarias* gariepinus) after processing and storage for short periods

Isny Maulidevi, Nurul Andriyani, Ryan Salfarino, Veriani Aprilia

Department of Nutrition, Faculty of Health Sciences, Alma Ata University, Yogyakarta, Indonesia. Corresponding author: V. Aprilia, verianiaprilia@almaata.ac.id

Abstract. Fresh fish is a highly perishable product due to its high protein content. A preservative may be needed to inhibit it. Nowadays, many fishes were reported to be exposed to formalin. This study aimed to determine the effect of short period processing (washing, frying, boiling in short-period) and storage (at the room and at freezing temperatures) on the residue of formalin in catfish, as a food model. Catfish, *Clarias gariepinus*, were divided into 8 groups, e.g. before treatment, washing in airflow for 1 minute, frying in 200°C for 8 minutes, boiling in 100°C for 7 minutes, storage at room temperature (29±1°C) for 24 and 48 hours, storage at freezing temperature (-3°C) for 24 and 48 hours. Before treatment, *C. gariepinus* was soaked in 1,480 ppm of formalin solutions. The residue of formalin was measured at the end of each treatment by using a spectrophotometer. Data were then analyzed with analysis of variance with SPSS software. The results showed that there were significant effects of processing and storage treatment on the formalin residue of *C. gariepinus*. The formalin residue of all treatments was lower than before treatments. The frying processing and storage at freezing temperatures in reducing the formalin residue. **Key Words**: fresh fish, preservative, food model, treatments, formalin residue.

Introduction. Fresh fish is a highly perishable product due to its high protein content for microbial activity (Wijayanti & Lukitasari 2016). A preservative maybe needs to keep the freshness, inhibit the decay and prolong the shelf life (Antoni 2010). Nowadays, there are many kinds of food-grade preservatives available in the market. Unfortunately, formalin is still frequently found in food products, including fresh fish, being used among fishermen mainly because it is effective in preserving the product, cheaper and accessible (Male et al 2017).

Formalin is a forty percent solution of formaldehyde in water. This compound is detected in the animal body as an intermediary metabolite. It also uses for several purposes, such as in medicine for specimen preservation and the treatment of hemorrhage (Pandey et al 2000). The preservative activity of formalin is due to its ability to react with protein generating methylene compounds, which are more difficult to be decayed by microbes (Ichya'uddin 2016). In low concentrations, formalin is not toxic for humans, but in higher concentrations, it had negative effects such as acute toxicity and death (Pandey et al 2000). It also decreases the levels of body antioxidants, resulting in liver damage (Yulisa et al 2014). Other studies also reported the cases of eczema, eye irritation, respiratory tract irritation, cancer, increased growth rate of malignant neoplastic diseases and denaturation of DNA (Suwanaruang 2018; Wilianto & Yudianto 2013). Therefore, its uses are forbidden.

Formalin solution is not stable during storage and exposure to high temperatures (Kaneko et al 1977; Laksmiani et al 2015). Those are caused by desolvation or evaporation of formalin during washing, boiling, or frying (Fellows 2000; Hayun et al 2017; Muntaha et al 2015; Sundari et al 2015) and enzymatic process in storage (Rachmawati et al 2007). Formalin that was in contact with fish decreased up to 43-53% after 5 times rinsing in 250 mL of distilled water and boiling for 30 minutes (Yusuf et al 2015). Other studies were also reported that soaking in warm water and frying decreased

the concentration of formalin up to 63.27% and 83.03%, respectively (Levita et al 2010). In daily practices, fish is usually washed in flowing water before processing, then fried/boiled for a short time. However, there was no study explaining the influences of these processing and storage methods on the concentration of formalin.

The purpose of this study was to identify the effect of processing for a short time and storage (at room and freezing temperatures) on the residue of formalin. This study used catfish, *Clarias gariepinus*, as a food model.

Material and Method

Material. The study used fresh *C. gariepinus*, purchased from the traditional market in Yogyakarta, Indonesia. The other used ingredients were formalin solution and Schiff's reagent. Formalin solution was made from 4 mL of formalin 37% in 1,000 mL of distilled water (Rachmawati et al 2007).

Method. The study was divided into two sections: 1) analyzing the effect of processing and 2) analyzing the effect of storage treatment. Fresh *C. gariepinus* were slaughtered, eviscerated, cleaned, and soaked in 1,480 ppm formalin solution (Rachmawati et al 2007; Sugiarti et al 2014).

For the first study, *C. gariepinus* were divided into four groups: controls (before treatment), washing, frying, and boiling treatments. Washing was done by watering the samples with running water for 1 minute, while frying was done by frying the samples in oil at 200°C for 8 minutes. Boiling treatment was done by putting the samples into boiled water (100°C) for 7 minutes (Sugiarti et al 2014).

For the second study, storage treatments, *C. gariepinus* was also divided into four groups: room temperature storage (29°C) for 24 hours and 48 hours and freezing temperatures storage (-3°C) for 24 hours and 48 hours (Handayani et al 2010). At the end of treatments, the concentration of formalin in samples was analyzed using a spectrophotometer. As the preparation step, a 5 g sample was dissolved in 25 mL of distilled water. 1 mL of supernatant was mixed with 2 mL of Schiff's reagent, and then heated in a water bath at 60°C for 5 minutes. It was incubated for 10 minutes before measuring the absorbance at 520 nm (Shita 2016; Mudaffar 2018).

Statistical analysis. Data were presented in mean±SD and analyzed with: (1) one-way analysis of variance (ANOVA) to determine the differences among all processing treatments and the effect of storage periods on the concentration of formalin; (2) paired sample T-test to determine the differences between the same samples before and after processing treatments; (3) independent sample t-test to know the effect of temperature at the same storage period, and (4) two-way ANOVA to determine the combined effect of storage temperature and period on the concentration of formalin. All data were analyzed with SPSS software (version 16.0 for Windows; SPSS Inc., Chicago, IL, USA) at 5% significance level.

Results

Effect of processing on the formalin residue of C. gariepinus. Table 1 shows that all treatments had significant effects on the residue of formalin (p<0.05). It can be seen from the p-value between the same samples before and after treatments. Among all treatments, washing shows the highest residue of formalin (p<0.05). Both boiling and frying had an almost similar residue of formalin (p>0.05). The lower residue of formalin in boiling and frying may be due to heating exposure.

Table 1

Processing	Residue of formalin (ppm)		Δ residue of formalin before
treatments	Before treatment	After treatment	and after treatment (%)
Washing	1,420.17±66.94 ^{c1}	310.42±77.61 ^{b2}	78.14
Frying	1,420.17±66.94 ^{c1}	14.18 ± 6.69^{a2}	99.01
Boiling	1,420.17±66.94 ^{c1}	27.50±9.11 ^{a2}	98.10

Mean \pm SD in the same column with different alphabetical superscripts (a, b, or c) are significantly different (p<0.05) by using one-way ANOVA statistical analysis. Mean \pm SD in the same row with different number superscripts (1 or 2) are significantly different (p<0.05) by using paired sample T-test statistical analysis.

Effect of storage temperatures and periods on the formalin residue of C. gariepinus. Table 2 shows the effect of storage temperatures, of periods and of their combination on the concentration of formalin. Based on this data, the storage period influenced the residue of formalin decrease (p<0.05) during the first 24 hours, but after a storage period of 48 hours the decrease was not significantly different (p>0.05).

On the other hand, there were no differences in the residue of formalin stored at room and freezing temperature, suggesting that the temperature of storage did not affect the formalin residue (p>0.05). Although there were differences in the statistical results between the temperature effects and the storage periods effects, the combined effect of both variables showed no impact on the concentration of formalin (p>0.05).

Table 2

Effect of storage temperature and period on the formalin residue of *Clarias gariepinus*

Temperature	Per	riods (t) (hours)			p-value	9
<i>(T)</i>	0	24	48	Т	t	T*t
Room (29°C)	1,420.17±66.94 ^{a1}	52.18±28.78 ^{a2}	40.64±3.93 ^{a2}	0.20	0.00	0.76
Freezing (-3°C)	1,420.17±66.94 ^{a1}	39.04±7.07 ^{a2}	40.64±3.93 ^{az} 13.65±4.23 ^{a2}	0.20	0.00	0.70

Mean±SD in the same column with different alphabetical superscripts (a, b, or c) are significantly different (p<0.05) and so are in the same row with different number superscript (1, 2, or 3). The p-value for T, t, and T*t was obtained from independent T-test, one-way ANOVA, and two-way ANOVA, respectively.

Discussion

The residue of formalin before treatment. The residue of formalin before and after treatment (either fish processing or storage) was significantly different (Table 1 and Table 2). In this study, the *C. gariepinus* specimens were submerged for 60 minutes in formalin solution of 1,480 ppm. The 60 minutes submersion period was chosen because the aldehyde compound may be readily bonded to the amino acids of fish in a small amount (Rahmadhani et al 2017). Formalin would attack lysine for the first time, followed by histidine and tyrosine, leading to the formation of a methylene compound (Ichya'uddin 2016). The protein-containing methylene compounds could not be digested by living things and microbes, especially pathogens, so that the shelf life would be longer (Purawisastra & Sahara 2011). The beneficial bacteria in the fish may also be killed by dehydration mechanism, as another effect of the existence of formalin in the body (Mudzkirah 2016).

The effect of processing on the formalin residue of C. gariepinus. The residue of formalin was reduced by 78.14% during washing treatment. The reduction was significant (p<0.05). The formalin was easily washed out because of its high solubility in the water, which is 4×10^5 mg L⁻¹ at 20°C (BPOM 2008; Sugiarti et al 2014). A study conducted by Yusuf et al (2015) showed the same trend, but the reduction was lower (with only 43%). The differences in the level of reduction can be affected by the duration of formalin submersion. The longer the submersion time, the smaller the decrease, as more formalin was bonded with the fish protein.

In the present study, frying also significantly reduced the residue of formalin (p<0.05). The reduction level was 99% indicating that most of the formalin disappeared during the frying process. Formalin was included in a volatile compound, especially above its boiling point (Joshi et al 2015). This study confirmed the results of Sugiarti et al (2014), showing a reduction level of formalin up to 99.28% after 1 hour of submersion followed by the frying process (no information about the period of frying process) in squid. Frying could denature the protein and also hydrolyze its bonding with formalin (Levita et al 2010). The high temperature in frying increased the kinetic energy and led to faster movement of the molecules that could damage the molecular bonds (Sugiarti et al 2014).

Heat processing by boiling reduced the residue of formalin by up to 98.1%, more than the washing process (p<0.05), but at the same degree as the frying process (p>0.05). The high reduction of formalin in the boiling process was due to the accelerated formalin dissolution rate at higher temperatures (Annisak 2019). This effect could be enhanced by boiling the sample in an open pan (Kamal et al 2017).

The effect of storage on the formalin residue of C. gariepinus. This experiment demonstrated that only the period of storage significantly influenced the residue of formalin (p<0.05), while the temperature of storage and the combination between temperature and period of storage did not (Table 2). The previous different studies showed that a storage at the room temperature or at the freezing temperature had similar effects on the formalin residue decrease (Jawahar et al 2017; Murtini et al 2014), possibly due to the same level of protein degradation by breaking the bonds between protein and formalin and releasing free compounds. At the room temperature, the protein degradation was essentially caused by the microbial activity, while at the freezing temperature it was caused mostly by physical destruction, leading to conformational and functional changes (Murtini et al 2014; Yeasmin et al 2010). Formalin in the free form is more reactive and can also be more easily degraded (Riyanto et al 2006; Rachmawati et al 2007).

Conclusions. The present research concluded that short-time processing and storage treatments decreased the concentration of formalin. Heat involvement determined a more effective decrease in concentration (the decrease level was >90%). In the storage study, the residue of formalin was significantly reduced by the period of storage, mainly during the first 24 hours, and then the formalin concentration reduction was insignificant, up to 48 hours of storage.

Acknowledgements. The authors are grateful to the Indonesian Ministry of Research, Technology, and Higher Education for providing the financial support in pursuing this research project (Program Kreativitas Mahasiswa Kemeristekdikti 2019 Program).

Reference

- Annisak S. K., Indriyanti N. Y., Mulyani B., 2019 [Constructive controversy and incubation presented according to the representation of tetrahedral chemical learning according to critical thinking ability]. Jurnal Inovasi Pendidikan IPA 5(1):10–22. [In Indonesian].
- Antoni S., 2010 [Analysis of formaldehyde content on salty fish by spectrophotometry method in Tampan District Pekanbaru]. BSc Thesis, Universitas Islam Negeri Sultan Syarif Kasim Riau, Indonesia, 53 p. [In Indonesian].
- Fellows P. J., 2000 Food processing technology: principle and practice. Woodhead Publishing Limited and CRC Press LLC, Boca Roton, USA, 356 p.
- Handayani A., Alimin, Rustiah W. O., 2010 [The effect of low temperature storage (freezer -3oc) on water and fat content in lemuru fish (*Sardinella longiceps*)]. Al-Kimia (3):64–75. [In Indonesian].

- Hayun H., Harmita K., Pramudita T. B., 2017 Determination of formaldehyde content in wet noodles by thin layer chromatography-densitometry after derivatization with Nash reagent. Oriental Journal of Chemistry 33(3):1400-1405.
- Ichya'uddin M., 2014 [Analysis of Formaldehyde content and organoleptic test on salty fish in several traditional markets in Tuban]. BSc Thesis, Universitas Islam Negeri Maulana Malik Ibrahim, Indonesia, 112 p. [In Indonesian].
- Jawahar L. S., John C., Shafeekh M., Anupama T. K., Sankar T. V., 2017 Retention of residual formaldehyde in treated Indian mackerel (*Rastrelliger kanagurta*) under iced storage and related food safety concern. Indian Journal of Fisheries 64(4):87–93.
- Joshi R., Bhatta R., Paudel P. N., Kafle B. K., 2015 Formaldehyde content of selected fish from the wet markets of Kathmandu valley. International Food Research Journal 22(4):1434-1437.
- Kamal A. S., Uddin M. M., Rahman R., Samira S., Rahman M., Nandy S., Khan M. S., 2017 Time and temperature effect on the residual concentration of formaldehyde in formalin treated samples of *Labeo rohita*. International Conference on Food Security and Nutrition, 6 p.
- Kaneko M., Wada Y., Fukui S., Kanno S., 1977 NII-Electronic library service. The Journal of Hygienic Chemistry 23(6):393–396.
- Laksmiani N., Widjaja I., Sonia, 2015 [Stability of formalin in different temperature and period of heating]. Jurnal Farmasi Udayana 4(2):76–81. [In Indonesian].
- Levita J., Musfiroh I., Indriyati W., Mustarichie R., 2010 The effect of soaking, washing and frying on the concentration of formaldehyde in Sange Belah salty fish. Jurnal Ilmu-ilmu Hayati dan Fisik 12(1):31–34.
- Male Y. T., Letsoin L. I., Siahaya N. A., 2017 [Analysis of formaldehyde content on noodle in several areas in Ambon]. Jurnal Kementerian Perindustrian 13(2):5–10. [In Indonesian].
- Mudaffar R. A., 2018 [Qualitative and quantitative test of formaldehyde on apple, grape, and litchi for sale in Makassar]. Jurnal Perbal 6(3):59-65. [In Indonesian].
- Mudzkirah I., 2016 [Identification of borax and formalin on snack food in UIN Alaudin Makassar Canteen]. BSc Thesis, UIN Alauddin Makassar, Indonesia, 95 p. [In Indonesian].
- Muntaha A., Haitami, Hayati N., 2015 [Comparison of reduction formaldehyde level on boiled and hot water soaked tofu]. Medical Laboratory Technology Journal 1(2):84– 90. [In Indonesian].
- Murtini J. T., Riyanto R., Priyanto N., Hermana I., 2014 [Development of Formaldehyde on some kinds of seawater fish during storage in crushed ice]. JPB Perikanan 9(2):143–151. [In Indonesian].
- Pandey C. K., Agarwal A., Baronia A., Singh N., 2000 Toxicity of ingested formalin and its management. Human and Experiment Toxicology 360–366.
- Purawisastra S., Sahara E., 2011 [Absorption to formalin by certain foodstuff and disappearance through hot water soaked]. The Journal of Nutrition and Food Research 34(1):63–74. [In Indonesian].
- Rachmawati N., Riyanto R., Ariyani F., 2007 [Identification of formaldehyde on kerapu macan fish (*Ephinephelus fuscoguttatus*) during cold storage]. Jurnal Pascapanen dan Bioteknologi Kelautan dan Perikanan 2(2):137–145. [In Indonesian].
- Rahmadhani F., Safrida, Djufri, 2017 [The effect of soaking a various of solution for reduction formalin level on kembung salty fish (*Scomber canagorta*) in Lambaro traditional market Aceh]. Jurnal Ilmiah Mahasiswa Pendidikan Biologi 2(3):1-12. [In Indonesian].
- Riyanto R., Kusmarwati A., Dwiyitno, 2006 [Identification of formaldehyde forming on kerapu fish (*Epinephelus fuscoguttatus*) during room temperature storage]. Jurnal Pascapanen dan Bioteknologi Kelautan dan Perikanan 1(2):111–116. [In Indonesian].
- Shita A. E., 2016 [Selectivity of spectrophotometer analysis with Schiff's reagent]. BSc Thesis, Universitas Negeri Yogyakarta, Indonesia, 87 p. [In Indonesian].

Sugiarti M., Anggo A. D., Riyadi P. H., 2014 [The effect of soaking in undercooking temperature and cooking method for reduction formalin level on cuttlefish]. Jurnal Pengolahan dan Bioteknologi Hasil Perikanan 3(2):90–98. [In Indonesian].

- Sundari D., Almasyhuri, Lamid A., 2015 [The effect of cooking process for nutrition composition of protein source]. Media Litbangkes 25(4):235–242. [In Indonesian].
- Suwanaruang T., 2018 Formalin contaminated in seafood and frozen meat at Somdet market, Kalasin Province. Journal of Environmental Protection 9:1286–1293.
- Wijayanti N. S., Lukitasari M., 2016 [Analysis of formalin level and organoleptic test in salty fish in Madiun Market]. Jurnal Florea 3(1):59–64. [In Indonesian].
- Wilianto W., Yudianto A., 2013 [The effect of formalin exposure on various levels for DNA psoas major by PCR Lokus CSF1PO, D5S818, D13S317, D21S11 Analysis]. Jurnal Kedokteran Forensik Indonesia 15(1):11–16. [In Indonesian].
- Yeasmin T., Reza M. S., Shikha F. H., Khan M. N. A., Kamal M., 2010 Quality changes in formalin treated rohu fish (*Labeo rohita*, Hamilton) during ice storage condition. Asian Journal of Agricultural Science 2(4):158–163.
- Yulisa N., Asni E., Azrin M., 2014 [Formaldehyde test of gurame fish in Pekanbaru Traditional Market]. Jurnal Online Mahasiswa 1(2):1-12. [In Indonesian].
- Yusuf Y., Zuki Z., Amanda R. R., 2015 [The effect of various treatmenst for reduction formalin level on fish by spectrophotometry method]. Jurnal Riset Kimia 8(2):182– 188. [In Indonesian].
- *** BPOM, Indonesian Food and Drug Authority, 2008 [Formaldehyde]. Badan POM RI, Jakarta, Indonesia. [In Indonesian].

Received: 30 September 2020. Accepted: 13 November 2020. Published online: 21 November 2020. Authors:

Isny Maulidevi, Alma Ata University, Faculty of Health Sciences, Department of Nutrition, Jl. Brawijaya 99, 55183 Yogyakarta, Indonesia, e-mail: isnymaulidevi540@gmail.com

Nurul Andriyani, Alma Ata University, Faculty of Health Sciences, Department of Nutrition, Jl. Brawijaya 99, 55183 Yogyakarta, Indonesia, e-mail: nurulandriyani96@gmail.com

Ryan Salfarino, Alma Ata University, Faculty of Health Sciences, Department of Nutrition, Jl. Brawijaya 99, 55183 Yogyakarta, Indonesia, e-mail: ryan.salfarino@gmail.com

Veriani Aprilia, Alma Ata University, Faculty of Health Sciences, Department of Nutrition, Jl. Brawijaya 99, 55183 Yogyakarta, Indonesia, e-mail: verianiaprilia@almaata.ac.id

How to cite this article:

Maulidevi I., Andriyani N., Salfarino R., Aprilia V., 2020 The residue of formalin in catfish (*Clarias gariepinus*) after processing and storage for short periods. AACL Bioflux 13(6):3441-3446.

This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution and reproduction in any medium, provided the original author and source are credited.